

Family Based Association Tests Using the **fbat** package

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1 Introduction

The R package **fbat** can be used to test the following null hypotheses for each marker based on family pedigrees:

- H_{01} : the marker has no association and no linkage with the trait;
 H_{02} : the marker has no association with the trait in the presence of linkage.

We assume that

- the families are **nuclear** families
- there are no missing genotypes and phenotypes for children
- markers are bi-allelic.

A more general software FBAT is available as a stand-alone executable with documentation and example files from <http://www.biostat.harvard.edu/~fbat/fbat.htm>. While this R package has some important limitations as present, these will be addressed in further versions.

2 Pedigree data file format

All fields are separated by whitespace (e.g. one or more spaces).

First line : names of all markers in the sequence of the genotype data. For example, marker₁, marker₂, ..., marker_m.

Remaining lines: The remaining lines contain only non-negative integers and have the same format:

family	pid	father	mother	sex	affection	marker _{1.1}	marker _{1.2}	...	marker _{m.1}	marker _{m.2}
--------	-----	--------	--------	-----	-----------	-----------------------	-----------------------	-----	-----------------------	-----------------------

where

family: family id

pid: patient id

father: father id.

Use 0 (zero) for founders or marry-ins (parents not specified) in a pedigree. A **founder** in a pedigree is an individual who is not a child of any individuals in the pedigree.

mother: mother id.

Use 0 (zero) for founders or marry-ins (parents not specified) in a pedigree.

A **founder** in a pedigree is an individual who is not a child of any individuals in the pedigree.

sex: 1 – male; 2 – female;

affection: affection status (i.e., trait)

2 – affected; 1 – unaffected; 0 – unknown

marker_{*i,j*}: allele *j* of marker *i*, $j = 1, 2; i = 1, 2, \dots, m$.

non-missing Alleles are represented by positive integers. Missing alleles are represented by zero (0).

3 Data quality control

The R package **fbat** also provides some basic QC functions.

The function **missGFreq** checks the completeness of genotypes. This function outputs counts of missing genotypes per marker and per subject.

The function **pedHardyWeinberg** checks the assumption of the Hardy-Weinberg equilibrium for markers.

The function **checkMendelian** checks the following possible Mendelian-related errors:

1. father id = subject id;
2. mother id = subject id;
3. could not determine if an individual is a parent or a child in a family;
4. inconsistent parental sex in a family;
5. parental genotypes are not compatible with childrens' genotypes in a family;
6. all childrens' genotypes are missing in a family;
7. inconsistent sib genotypes in a family.

4 Examples

To call the functions in the R package **fbat**, we first need to load it into R:

To read the pedigree file **CAMP.ped** into R, we use the function **readGenes** in the R package **GeneticsBase**:

```
gSet<-readGenes(gfile="CAMP.ped", gformat="fbat")
```

The function `readGenes.ped` returns back an object of the R class *geneSet*.

Before we apply family based association tests, it would be good practice to check Hardy-Weinberg equilibrium for each marker based on parental data. We can use the function `pedHardyWeinberg` to do this.

```
> data(CAMP)
```

```
Reading 8 markers and 2011 subjects from `CAMP.ped` ...  
generating 'geneSet' object...
```

```
Successfully read the pedigree file `CAMP.ped`.
```

```
Number of Markers: 8  
Number of Subjects: 2011  
Number of Families: 651
```

```
Reading 12 vars from `CAMPZ.phe` ... Done.
```

```
Number of Phenotype Variables: 12  
Number of Observations      : 2011
```

```
> ch <- pedHardyWeinberg(CAMP)
```

```
converting geneSet object to numerical matrix...  
HWE test...
```

	nInfoInd	nGenotype	nHET	nHOM	nAllele	nMissing	chi2	df	p-value
m709	1269	3	4	1265	2	34	0.003	1	0.955
m654	1259	3	558	701	2	44	1.080	1	0.299
m47	1244	3	582	662	2	59	0.005	1	0.944
p46	1253	3	596	657	2	50	0.029	1	0.864
p79	1244	3	580	664	2	59	0.030	1	0.862
p252	1184	3	410	774	2	119	0.685	1	0.408
p491	1263	3	27	1236	2	40	0.147	1	0.701
p523	1271	3	405	866	2	32	0.082	1	0.775

The column `nInfoInd` means the number of informative individuals, i.e. individuals whose genotypes contain no missing alleles for the specified marker; the column `nGenotype` means number of possible genotypes; the column `nHET` means number of heterozygous genotypes; the column `nHOM` means number of homozygous genotypes; the column `nAllele` means number of alleles; the column `nMissing` means number of missing alleles; the column `chi2` means chi square test statistic; the column means `df` means degree of freedom of the chi square test statistic under the null hypothesis that Hardy-Weinberg condition holds; and the column `p-value` means pvalue of the test.

To view the statistics for individual markers, we can use the function `viewHW`. For example,

```
> viewHW(ch, "p79")

number of possible genotypes for marker p79 >>
[1] 3
genotype frequency >>
      p79.1 p79.2 freq
[1,]      1      1 488
[2,]      1      2 580
[3,]      2      2 176
allele frequency >>
      1      2
0.625 0.375
nInfoInd nGenotype      nHET      nHOM  nAllele nMissing      chi2      df
1244.000      3.000  580.000  664.000      2.000      59.000    0.030    1.000
p-value
0.862
```

To check Mendelian-related errors, we can use the function `checkMendelian`. For example,

```
> tmp <- checkMendelian(CAMP, quiet = TRUE)
> cat("For each marker, how many families contains mendelian errors?\n")

For each marker, how many families contains mendelian errors?

> print(tmp$nMerrMarker)

m709 m654  m47  p46  p79 p252 p491 p523
  19  129  128  131  134  140   25  101

> cat("For each family, how many markers contains mendelian errors?\n")

For each family, how many markers contains mendelian errors?

> cat("tmp$nMerrFamily[1:10]>>\n")

tmp$nMerrFamily[1:10]>>

> print(tmp$nMerrFamily[1:10])

family1 family2 family3 family4 family5 family6 family7 family8
      0      1      0      0      1      0      0      0
family9 family10
      0      0
```

```
> cat("For each family, how many times\n")
```

For each family, how many times

```
> cat("'father id = subject id' or 'mother id = subejct id'?\n")
```

'father id = subject id' or 'mother id = subejct id'?

```
> cat("tmp$nErrFamilySample[1:10]>>\n")
```

tmp\$nErrFamilySample[1:10]>>

```
> print(tmp$nErrFamilySample[1:10])
```

family1	family2	family3	family4	family5	family6	family7	family8
0	0	0	0	0	0	0	0
family9	family10						
0	0						

To count the number of missing genotypes for a marker or for a subject, we can use the function `missGFreq`. For example,

```
> res <- missGFreq(CAMP, founderOnly = FALSE, quiet = TRUE)
```

```
> cat("The number of missing genotypes for markers>>")
```

The number of missing genotypes for markers>>

```
> print(res$nMissMarkers)
```

	00	0*	*0
m709	55	0	0
m654	60	0	0
m47	89	0	0
p46	68	0	0
p79	90	0	0
p252	188	0	0
p491	57	0	0
p523	53	0	0

```
> cat("The number of missing genotypes for the first 10 subjects>>")
```

The number of missing genotypes for the first 10 subjects>>

```
> print(res$nMissSubjects[1:10, ])
```

	00	0*	*0
subject1	0	0	0
subject2	0	0	0
subject3	0	0	0
subject4	0	0	0
subject5	0	0	0
subject6	1	0	0
subject7	0	0	0
subject8	0	0	0
subject9	0	0	0
subject10	0	0	0

To get the family based association test statistics, we use the function `fbat`:

```
> res <- fbat(CAMP)
```

The usage of the function `fbat` is

```
fbat(geneSetObject, model="a", traitMethod=3, traitOffset=0, quiet=TRUE)
```

The function argument `model` specifies the genotype codings.

By default, we use the additive model (`model="a"`). Other available models include dominant (`model="d"`), recessive (`model="r"`), and genotype (`model="g"`) models.

The function argument `traitMethod` indicates the trait coding method. If `traitMethod` is equal to 1, then the trait is represented by `trait-offset` where `trait` is the sixth column (i.e., `affection` status) of the pedigree matrix and the value of `offset` is provided by the argument `traitOffset`. If the argument `traitMethod` takes value other than 1, then the trait is set to be 1 if the sixth column of the pedigree matrix takes value 2 and the trait is set to be 0 if the sixth column of the pedigree matrix takes value 1.

The function `fbat` returns a list. To summarize the values, degrees of freedom, and *p*-values of the test statistics for the markers, we can use the function `summaryPvalue`:

```
> summaryPvalue(res)
```

```
*****
      chisq rank      pvalue
m709  1.0000000      1 3.173105e-01
m654  1.3677298      1 2.422023e-01
m47   16.5161290      1 4.823799e-05
p46   0.1130742      1 7.366710e-01
p79   11.1838235      1 8.251356e-04
p252  37.7790698      1 7.922726e-10
p491  18.2413793      1 1.946047e-05
p523  45.7821782      1 1.321609e-11
*****
```

To adjust multiple comparisons, we can use the function `p.adjust` in the R package `base` to adjust the p -values. For example,

```
> pvals <- res$statPvalue[, 3]
> p.adjust.M <- p.adjust.methods
> p.adj <- sapply(p.adjust.M, function(meth) p.adjust(pvals, meth))
> noquote(apply(p.adj, 2, format.pval, digits = 3))
```

	holm	hochberg	hommel	bonferroni	BH	BY	fdr	none
[1,]	0.726607	0.634621	0.634621	1.000000	0.36264	0.985605	0.36264	0.317311
[2,]	0.726607	0.634621	0.484405	1.000000	0.32294	0.877695	0.32294	0.242202
[3,]	0.000241	0.000241	0.000241	0.000386	9.65e-05	0.000262	9.65e-05	4.82e-05
[4,]	0.736671	0.736671	0.736671	1.000000	0.73667	1.000000	0.73667	0.736671
[5,]	0.003301	0.003301	0.003301	0.006601	0.00132	0.003588	0.00132	0.000825
[6,]	5.55e-09	5.55e-09	5.55e-09	6.34e-09	3.17e-09	8.61e-09	3.17e-09	7.92e-10
[7,]	0.000117	0.000117	0.000117	0.000156	5.19e-05	0.000141	5.19e-05	1.95e-05
[8,]	1.06e-10	1.06e-10	1.06e-10	1.06e-10	1.06e-10	2.87e-10	1.06e-10	1.32e-11

To view summary statistics of individual marker, we can use the function `viewstat`. For example,

```
> viewstat(res, "p79")

*****
651 pedigree 2011 persons
359 informative families at marker p79
The alleles of marker p79 >>
[1] 1 2
Score for marker p79 >>
[1] 471 301
Expected score for marker p79 >>
[1] 432 340
Covariance matrix of the score for marker p79 >>
      [,1] [,2]
[1,] 136 -136
[2,] -136 136
Moore-Penrose generalized inverse of covariance matrix
      [,1] [,2]
[1,] 0.001838235 -0.001838235
[2,] -0.001838235 0.001838235
test statistics for marker p79 >>
      chisq      rank      pvalue
1.118382e+01 1.000000e+00 8.251356e-04
*****
```


Note that if the covariance matrix of the S score vector is singular, the Moore-Penrose generalized inverse is used.

Sometimes the user might want to know if a genotype is homozygous or heterozygous. The function `pedFlagHomo` can provide those information. For example,

```
> res.f <- pedFlagHomo(CAMP)
```

converting geneSet object to numerical matrix...
flag homozygotes and heterozygotes...
dim(flagHomoMat)= 1303 8
length(ped[,2])= 1303
numHomo -- number of homozygous genotypes
numHetero -- number of heterozygous genotypes
numMiss1 -- number of genotypes containing one missing allele
numMiss2 -- number of genotypes containing two missing alleles
counts>>>

	numHomo	numHetero	numMiss1	numMiss2
m709	1265	4	0	34
m654	701	558	0	44
m47	662	582	0	59
p46	657	596	0	50
p79	664	580	0	59
p252	774	410	0	119
p491	1236	27	0	40
p523	866	405	0	32

The function `pedGFFreq` gets genotype frequencies and percentages. For example,

```
> res <- pedGFFreq(CAMP)
```

converting geneSet object to numerical matrix...
counting genotype frequencies...
genotype counts>>>

	1/1	1/2	2/2
m709	1265	4	0
m654	536	558	165
m47	171	582	491
p46	197	596	460
p79	488	580	176
p252	68	410	706
p491	1236	27	0
p523	813	405	53

The function `pedAFreq` gets allele frequencies and percentages. For example,

```
> res <- pedAFreq(CAMP)
```

```
converting geneSet object to numerical matrix...
count allele frequencies...
allele frequencies and percentages>>>
```

```
      1      2      1      2
m709 2534      4 0.998 0.002
m654 1630  888 0.647 0.353
m47   924 1564 0.371 0.629
p46   990 1516 0.395 0.605
p79  1556  932 0.625 0.375
p252  546 1822 0.231 0.769
p491 2499   27 0.989 0.011
p523 2031  511 0.799 0.201
```

The functions `fbat`, `pedHardyWeinberg`, `pedFlagHomo`, `pedGFreq`, and `pedAFreq` have default forms (`fbat.default`, `pedHardyWeinberg.default`, `pedFlagHomo.default`, `pedGFreq.default`, and `pedAFreq.default`) that use a pedigree matrix as input.

Appendix

A Notation

For a given marker,

- Y_{ij} — Observed trait of the j -th offspring in family i .
- T_{ij} — A function of Y_{ij} .

$$T_{ij} = T(Y_{ij}).$$

For example

$$T_{ij} = T(Y_{ij}) = Y_{ij} - \mu_{ij},$$

where μ_{ij} is an offset.

- g_{ij} — Genotype of the j -th offspring in family i ;
- X_{ij} — A function of g_{ij} .

$$X_{ij} = X(g_{ij}).$$

- S score:

$$S = \sum_{ij} T_{ij} X_{ij} = \sum_{ij} T(Y_{ij}) X(g_{ij}).$$

- test statistic:

$$U = S - E[S|H_0, \mathcal{C}],$$

where \mathcal{C} is a condition set. When parental genotypes are complete, the condition set $\mathcal{C} = \mathcal{T} \cup \mathcal{G}$, where \mathcal{T} is the observed traits in all family members and \mathcal{G} is the parental genotypes. When parental genotypes are incomplete, the condition set $\mathcal{C} = \mathcal{T} \cup \mathcal{G}^* \cup \mathcal{G}_{\text{offspring}}$, \mathcal{G}^* is the partially observed parental genotypes and $\mathcal{G}_{\text{offspring}}$ is the set of offspring genotypes (i.e., the offspring genotype configuration).

- V – variance or covariance matrix of U under the null hypothesis H_0 . I.e.,

$$V = \text{Cov}(U|H_0, \mathcal{C}) = \text{Cov}(S|H_0, \mathcal{C}).$$

- For the univariate case,

$$Z = \frac{U}{\sqrt{V}} \Big|_{H_0, \mathcal{C}} \dot{\rightarrow} N(0, 1).$$

- For the multivariate case,

$$\chi^2 = U'V^{-1}U \Big|_{H_0, \mathcal{C}} \dot{\rightarrow} \chi_r^2,$$

where $r = \text{rank}(V)$.

B Genotype coding methods

Denote K as the number of all possible different alleles for the locus and X as the vector of genotype coding.

GEN X is a vector with length equal to the number of genotypes that are possible given the parental genotypes in the sample, a maximum of $K(K+1)/2$ genotypes, and with elements equal to 1 or 0 to indicate which of the possible genotypes is equal to the genotype g .

GDOM codes the j th element of the vector X as $x_j = 1$ if genotype g has one or two alleles of type j , otherwise $x_j = 0$. X is a vector of length K .

GREC codes the j th element of the vector X as $x_j = 1$ if genotype g has two alleles of type j , otherwise $x_j = 0$. X is a vector of length K .

GTD scores the number of alleles of a particular type by coding x_j equal to the number of alleles of type j in the genotype g (i.e., $x_j = 0, 1$, or 2 if g has 0, 1 or 2 alleles of type j). X is a vector of length K .

2-allele case

Example of different marker codings for a marker with $K = 2$ alleles, see Schaid (1996)

genotype	$X(g)$			
g	GEN	GDOM	GREC	GTDT
		(A, a)	(A, a)	(A, a)
AA	$(0,0,0)$	$(1,0)$	$(1,0)$	$(2,0)$
Aa	$(1,0,0)$	$(1,1)$	$(0,0)$	$(1,1)$
aa	$(0,1,0)$	$(0,1)$	$(0,1)$	$(0,2)$

3-allele case

Example of different marker codings for a marker with $K = 3$ alleles, see Schaid (1996) (This table is Table 4 of Horvath et al.'s report for FBAT software)

genotype	$X(g)$			
g	GEN	GDOM	GREC	GTDT
		(A, B, C)	(A, B, C)	(A, B, C)
AA	$(0,0,0,0,0)$	$(1,0,0)$	$(1,0,0)$	$(2,0,0)$
AB	$(1,0,0,0,0)$	$(1,1,0)$	$(0,0,0)$	$(1,1,0)$
AC	$(0,1,0,0,0)$	$(1,0,1)$	$(0,0,0)$	$(1,0,1)$
BB	$(0,0,1,0,0)$	$(0,1,0)$	$(0,1,0)$	$(0,2,0)$
BC	$(0,0,0,1,0)$	$(0,1,1)$	$(0,0,0)$	$(0,1,1)$
CC	$(0,0,0,0,1)$	$(0,0,1)$	$(0,0,1)$	$(0,0,2)$

C Trait coding methods

Denote Y_{ij} as the trait of the j -th child of the i th nuclear family. Y_{ij} can be dichotomous, measured (i.e., continuous?), time-to-onset (i.e., censored?)

The trait coding methods ($T_{ij} = T(Y_{ij})$) are listed below:

- $T_{ij} = 1$ if the j th child is affected; $T_{ij} = 0$ otherwise.
- $T_{ij} = Y_{ij} - \mu_{ij}$, where μ_{ij} is an offset.
- $T_{ij} = Y_{ij} - \mu_{ij}(\mathbf{x}'\boldsymbol{\beta})$, where $E(Y_{ij}|\mathbf{x}) = \mu_{ij}(\mathbf{x}'\boldsymbol{\beta})$, and \mathbf{x} are design matrix of covariates, $\boldsymbol{\beta}$ are unknown parameters.