

Bayesian Analysis of ChIP-chip

The BAC package

Raphael Gottardo

April 24, 2017

Contents

I	Licensing	2
II	Introduction	2
III	Loading the BAC Package	2
IV	Detecting bound regions	3

Part I

Licensing

This software is distributed under the Artistic License 2.0. In addition, we would appreciate if you cite the following papers when using this software for publication.

W. E. Johnson, Li, W., Meyer, C. A., Gottardo, R., Carroll, J. S., Brown, M., and Liu, X. S. (2006). Model-based analysis of tiling-arrays for ChIP-chip. PNAS 103:12457-12462.

Gottardo et al. A Flexible and Powerful Bayesian Hierarchical Model for ChIP-Chip Experiments. Biometrics (2007) In press.

Then if you use the ER data (part of it is included in this package), you should cite the following paper:

Carroll et al 2006 <http://www.cell.com/content/article/abstract?uid=PIIS0092867405004538>

Part II

Introduction

In our guide, we include example of codes that we hope will help you when using the BAC package. The codes are kept at the basic level for ease of understanding. Some of the options in the functions have been set by default. To learn more about the exact parameters and usage of each function, you may type `help(FUNCTION_NAME)` of the function of interest in R after the BAC package is loaded.

The common goal in analyzing this ChIP-chip data is to detect DNA-protein interactions from ChIP-chip experiments. As of now, the BAC package has mainly be tested with Affymetrix tiling array data. However, we expect it to work with other platforms (e.g. Agilent, Nimblegen, cDNA, etc.). In order to use the BAC package you will need both treatment (IP) and control conditions (e.g. Mock IP) with replicates under each condition. Note that BAC does not deal with normalization, so you will have to normalize your data before hands. For Affymetrix arrays, we refer you to the MATR package which contains efficient normalization procedures.

Part III

Loading the BAC Package

To load the BAC package in R, we type

```
> library(BAC)
```

Part IV

Detecting bound regions

We first load the estrogen receptor data (Carroll et al. 2006).

```
> data(ER)
```

then we calculate the (joint) posterior probabilities of enrichment based on 50 iterations for speed up, you should use more when you run the BAC function (see default parameters)

```
> # Please uncomment the following line to test the code
> # BAConER<-BAC(ER[,5:7], ER[,2:4], B=50,verbose=FALSE,w=5)
> # This load the resulting data obtained after executing the line above
> load("bac.rda")
```

where w=5 is the window size, see parameter description for more details. Now you can have a look at these posterior probabilities to see where enriched regions might be, see Figure 1.

```
> plot(ER[,1],BAConER$jointPP,pch="+",xlab="Genomic Position",ylab="Posterior probabilities")
```



Now regions can be called putative regions using the `CallRegions` function by applying a 0.5 threshold (other thresholds can be used)

```
> ERregions<-CallRegions(ER[,1],BAConER$jointPP,cutoff=0.5,maxGap=500)
```

Finally, once we are happy with the regions detected we can easily create a BED file, which can be read and visualize in the UCSC genome browser.

```
> # Create the BED file
> nRegions<-max(ERregions)
> BED<-matrix(0,nRegions,4)
> for(i in 1:nRegions)
+ {
+   BED[i,2:3]<-range(ER[ERregions==i,1])
+   #The score should be between 0 and 1000
+   BED[i,4]<-max(BAConER$jointPP[ERregions==i])*1000
+ }
> BED<-data.frame(BED)
> # The ER data is a subset of chr 21
> BED[,1]<-"chr21"
> names(BED)<-c("chrom","chromStart","chromEnd","Score")
> # print it
> print(BED)
```

	chrom	chromStart	chromEnd	Score
1	chr21	14600350	14600679	1000
2	chr21	15171823	15172238	1000
3	chr21	15299545	15299909	980
4	chr21	15467160	15467873	1000
5	chr21	15493634	15495171	1000
6	chr21	15497944	15498912	1000
7	chr21	15503734	15504989	1000
8	chr21	15505697	15506008	920
9	chr21	15636775	15637099	1000
10	chr21	15738481	15738680	900
11	chr21	15880913	15881171	1000