Package 'SIM'

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Author Renee X. de Menezes and Judith M. Boer
Maintainer Renee X. de Menezes < r. menezes@nki.nl>
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SIM-p	package Statistical Integration of Microarrays	

Description

SIM is a statistical model to identify associations between two genomic datasets. Where one is assigned as dependent variable and the other as independent e.g. copy number measurements on several samples versus expression measurements on the same samples. A region of interest can be chosen to run the integrated analysis on either the same region for both dependent and independent datasets or different regions. For each dependent feature a P-value measures the association with the independent data, the contribution of each independent feature is given as Z-scores. The integrated analysis is based on the random-effect model for gene-sets as implemented in gt.

maybe something about annotation?

By default we use method.adjust = "BY" (Benjamini-Yekutieli) for multiple testing correction. This method accounts for dependence between measurements and is more conservative than "BH" (Benjamini-Hochberg). For details on the multiple testing correction methods see p.adjust. We have experienced that a rather low stringency cut-off on the BY-values of 20% allows the detection of associations for data with a low number of samples or a low frequency of abberations. False positives are rarely observed.

Make sure that the array probes are mapped to the same builds of the genome, and that the chrom.table used by the integrated.analysis is from the same build as well. See sim.update.chrom.table.

Details

Package: SIM
Type: Package
Version: 1.7.1
Date: 2010-09-14
License: Open

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Author(s)

Marten Boetzer, Melle Sieswerda, Renee X. de Menezes < R. X. Menezes@lumc.nl>

References

Menezes RX, Boetzer M, Sieswerda M, van Ommen GJ, Boer JM (2009). Integrated analysis of DNA copy number and gene expression microarray data using gene sets. *BMC Bioinformatics*, **10**, 203-.

Goeman JJ, van de Geer SA, de Kort F, van Houwelingen HC (2004). A global test for groups of genes: testing association with a clinical outcome. *Bioinformatics*, **20**, 93-109.

See Also

assemble.data, integrated.analysis, sim.plot.zscore.heatmap, sim.plot.pvals.on.region, sim.plot.pvals.on.genome, tabulate.pvals, tabulate.top.dep.features, tabulate.top.indep.features, impute.nas.by.surrounding, sim.update.chrom.table, sim.plot.overlapping.indep.dep.features, getoverlappingregions

Examples

```
#load the datasets and the samples to run the integrated analysis
data(expr.data)
data(acgh.data)
data(samples)
#assemble the data
assemble.data(dep.data = acgh.data,
              indep.data = expr.data,
              dep.ann = colnames(acgh.data)[1:4],
              indep.ann = colnames(expr.data)[1:4],
              dep.id="ID",
              dep.chr = "CHROMOSOME",
              dep.pos = "STARTPOS",
              dep.symb="Symbol",
              indep.id="ID",
              indep.chr = "CHROMOSOME",
              indep.pos = "STARTPOS",
              indep.symb="Symbol",
              overwrite = TRUE,
              run.name = "chr8q")
#run the integrated analysis
integrated.analysis(samples = samples,
                    input.regions ="8q",
                    zscores=TRUE,
                    run.name = "chr8q")
# use functions to plot the results of the integrated analysis
#plot the P-values along the genome
sim.plot.pvals.on.genome(input.regions = "8q",
                         significance = c(0.2, 0.05),
```

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```
adjust.method = "BY",
                         pdf = FALSE,
                         run.name = "chr8q")
#plot the P-values along the regions
sim.plot.pvals.on.region(input.regions = "8q",
       adjust.method="BY",
       run.name = "chr8q")
#plot the z-scores in an association heatmap
#plot the zscores in a heatmap
sim.plot.zscore.heatmap(input.regions = "8q",
                        method="full",
                        significance=0.2,
                        z.threshold=3,
                        show.names.indep=TRUE,
                        show.names.dep=TRUE,
                        adjust.method = "BY",
                        add.plot = "smooth",
                        smooth.lambda = 2,
                        pdf = FALSE,
                        run.name = "chr8q")
sim.plot.zscore.heatmap(input.regions = "8q",
                        method="full",
                        significance = 0.05,
                        z.threshold = 1,
                        show.names.indep=TRUE,
                        show.names.dep=FALSE,
                        adjust.method = "BY",
                        add.plot = "heatmap",
                        smooth.lambda = 2,
                        pdf = FALSE,
                        run.name = "chr8q")
sim.plot.zscore.heatmap(input.regions = "8q",
                        method="full",
                        significance = 0.05,
                        z.threshold = 1,
                        show.names.indep=TRUE,
                        show.names.dep=TRUE,
                        adjust.method = "BY",
                        add.plot = "none",
                        pdf = FALSE,
                        run.name = "chr8q")
#tabulate the P-values per region (prints to screen)
tabulate.pvals(input.regions = "8q",
               adjust.method="BY",
               bins=c(0.001,0.005,0.01,0.025,0.05,0.075,0.10,0.20,1.0),
               run.name = "chr8q")
table.dep <- tabulate.top.dep.features(input.regions="8q",
```

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```
adjust.method="BY",
        method="full",
        significance=0.05,
        run.name="chr8q")
head(table.dep[["8q"]])
table.indep <- tabulate.top.indep.features(input.regions="8q",</pre>
                                     adjust.method="BY",
            method="full",
            significance= 0.05,
            z.threshold=c(-1, 1),
            run.name="chr8q")
head(table.indep[["8q"]])
sim.plot.overlapping.indep.dep.features(input.regions="8q",
                                   adjust.method="BY",
          significance=0.1,
          z.threshold=c(-1,1),
          log=TRUE,
          summarize="consecutive",
          pdf=FALSE,
          method="full",
          run.name="chr8q")
```

acgh.data

Array Comparative Genomic Hybridization data

Description

Copy number data taken from Pollack et al. PNAS. 2002, 99(20): 12963-8.

Usage

```
data(acgh.data)
```

Format

A data frame with 99 observations on 45 variables. The first 4 columns are the unique identifier, symbol for the chromosome and start position of the probe the next 41 columns are the copy number measurements of 41 samples.

Details

A subset of the original data is taken namely all the probes on the long arm of chromosome 8.

Source

http://genome-www.stanford.edu/aCGH_breast/data.shtml

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References

Pollack JR, Sorlie T, Perou CM, Rees CA, Jeffrey SS, Lonning PE, Tibshirani R, Botstein D, Borresen-Dale AL, Brown PO (2002). Microarray analysis reveals a major direct role of DNA copy number alteration in the transcriptional program of human breast tumors. *Proc Natl Acad Sci USA*. **1**, 99(20), 12963-8.

Examples

```
data(acgh.data)
```

assemble.data

Assemble the data to run the integrated analysis

Description

Assembles the dependent and independent data and annotation of the both data sets.

Usage

```
assemble.data(dep.data,
  indep.data,
  dep.id = "ID",
  dep.chr = "CHROMOSOME",
  dep.pos = "STARTPOS",
  dep.ann = NULL,
  dep.symb,
  indep.id = "ID",
  indep.chr = "CHROMOSOME",
  indep.pos = "STARTPOS",
  indep.ann = NULL,
  indep.symb,
  overwrite = FALSE,
  run.name = "analysis_results")
```

Arguments

dep.data

The dependent data (data.frame), along with annotations. Each row should correspond to one feature. The following columns are expected to exist, and the column names should be inserted in the function. dep.id: A unique identifier. dep.chr: The number of the chromosome (1,2,...,22,X,Y). dep.pos: The base pair position, relative to the chromosome. dep.symb: Gene symbol (optional). dep.ann: Annotation can be multiple columns.

indep.data

data.frame The independent data, along with annotations. Each row should correspond to one feature. The following columns are expected to exist, and the column names should be inserted in the function. indep.id: A unique identifier. indep.chr: The number of the chromosome (1,2, ..., 22, X, Y). indep.pos: The base pair position, relative to the chromosome. indep.symb: Gene symbol (optional). indep.ann: Annotation can be multiple columns.

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dep.ann	vector with either the names of the columns or the column numbers in the dependent data that contain the annotation.
indep.ann	vector with either the names of the columns or the column numbers in the independent data that contain the annotation.
dep.id	vector with the column name in the dependent data that contains the ID. Will be used in the sim.plot.zscore.heatmap function. Empty ID's will be substituted by NA.
dep.chr	vector with column name in the dependent data that contains the chromosome numbers.
dep.pos	vector with the column name in the dependent data that contains the position on the chromosome in bases.
dep.symb	Optional, either missing or a single vector with the column name in the dependent data that contains the symbols. Will be used in sim.plot.zscore.heatmap as label.
indep.id	vector with the column name in the independent data that contains the ID. Will be used in the sim.plot.zscore.heatmap function. Empty ID's will be substituted by NA.
indep.chr	vector with the column name in the independent data that contains the chromosome numbers.
indep.pos	vector with the column name in the independent data that contains the position on the chromosome in bases.
indep.symb	Optional, either missing or a vector with the column name in the dependent data that contains the Symbols. Will be used in sim.plot.zscore.heatmap as label.
overwrite	\log logical, indicate when a run.name is already present, the results can be overwritten.
run.name	Name of the analysis. The results will be stored in a folder with this name in the current working directory (use getwd() to print the current working directory). If the missing, the default folder "analysis_results" will be generated.

Details

Based on the chromosome and probe position an absolute position is calculated according to chromosomenumber*1e9 + probe position. Chromosome column is converted to factor and releveled according to the levels of the chrom.table, so the only levels allowed are c(1:22, "X", "Y"). Currently only human genome support without mitochondrial DNA.

Value

No values are returned. Instead, the datasets and annotation columns are stored in separate files in the data folder in the directory specified in run.name. If assemble.data has run successfully, the integrated.analysis function can be performed.

Author(s)

Marten Boetzer, Melle Sieswerda, Renee X. de Menezes < R. X. Menezes@lumc.nl>

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See Also

SIM, integrated.analysis

Examples

```
# Generate datasets and the samples to run the integrated analysis
set.seed(53245)
ngenes <- 100
nsamples <- 100
# generate copy number measurements
x <- matrix(rnorm(n = ngenes*nsamples), nrow = ngenes, ncol = nsamples)</pre>
# add mean shift effect for half of the samples, copy gain for 2nd half of the genes
x[seq_len(ngenes/2), seq_len(nsamples/2)] <- x[seq_len(ngenes/2), seq_len(nsamples/2)] + 2
# generate gene expression with normal distribution and mean equal to gene copy number
y <- rnorm(n = ngenes*nsamples, mean = matrix(x, nrow = ngenes*nsamples, ncol = 1), sd = 0.8)
v <- matrix(v, nrow = ngenes, ncol = nsamples)</pre>
samples <- paste0("S", seq_len(nsamples))</pre>
colnames(x) \leftarrow colnames(y) \leftarrow samples
# Making data objects
acgh.data <- data.frame(ID = paste0("G", seq_len(ngenes)),</pre>
                      CHROMOSOME = rep(1, ngenes),
                      STARTPOS = seq_len(ngenes)*12*10^5,
                      Symbol = paste0("Gene", seq_len(ngenes)),
                      x)
expr.data <- data.frame(ID = paste0("G", seq_len(ngenes)),</pre>
                         CHROMOSOME = rep(1, ngenes),
                         STARTPOS = seq_len(ngenes)*12*10^5,
                         Symbol = paste0("Gene", seq_len(ngenes)),
                         y)
#assemble the data
assemble.data(dep.data = acgh.data,
              indep.data = expr.data,
               dep.ann = colnames(acgh.data)[1:4],
               indep.ann = colnames(expr.data)[1:4],
               dep.id="ID",
              dep.chr = "CHROMOSOME",
               dep.pos = "STARTPOS",
               dep.symb="Symbol",
               indep.id="ID",
               indep.chr = "CHROMOSOME",
               indep.pos = "STARTPOS",
               indep.symb="Symbol",
              overwrite = TRUE,
              run.name = "chr1p")
```

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Description

A table indicating the base positions of the beginning and end of chromosome arms and bands. Currently based on the UCSC March 2006/NCBI 36 build of the human genome.

Usage

```
data(chrom.table)
```

Format

A data frame with 862 observations on the following 6 variables. chr, arm, band, start, end, stain

Details

Possibly the chrom. table can be update by sim.update.chrom.table. Currently only human genome support without mitochondrial DNA.

See Also

sim.update.chrom.table

Examples

```
data(chrom.table)
```

expr.data

Expression data example

Description

Expression data taken from Pollack et al. PNAS. 2002, 99(20): 12963-8.

Usage

```
data(expr.data)
```

Format

A data frame with 99 observations on 45 variables. The first 4 columns are the unique identifier, symbol for the chromosome and start position of the probe the next 41 columns are the expression log-ratios of 41 samples.

Details

A subset of the original data is taken namely all the probes on the long arm of chromosome 8.

Source

http://genome-www.stanford.edu/aCGH_breast/data.shtml

References

Pollack JR, Sorlie T, Perou CM, Rees CA, Jeffrey SS, Lonning PE, Tibshirani R, Botstein D, Borresen-Dale AL, Brown PO (2002). Microarray analysis reveals a major direct role of DNA copy number alteration in the transcriptional program of human breast tumors. *Proc Natl Acad Sci USA*. **1**, 99(20), 12963-8.

Examples

```
data(expr.data)
```

getoverlappingregions Get the overlapping regions between independent and dependent regions

Description

Generates a table with overlapping regions.

Usage

Arguments

independent_regions

data.frame().Independent regions found with tab tabulate.top.indep.features.

dependent_regions

data.frame().Independent regions found with tab tabulate.top.dep.features.

method

method to estimate the overlapping regions, either "union" or "overlapping". overlapping outputs only the overlapping parts of the overlapping regions. union outputs the whole region. Say independent region = 1-10, dependent region = 5-12. The output is 1-12.

Details

Calculates the overlap between two tables.

Author(s)

Marten Boetzer, Melle Sieswerda, Renee X. de Menezes < R. X. Menezes@lumc.nl>

See Also

SIM, tabulate.top.dep.features, tabulate.top.indep.features, sim.plot.overlapping.indep.dep.features

Examples

```
#no examples yet!
```

```
impute.nas.by.surrounding
```

Impute NA's in array-CGH data

Description

Replace an NA by the median of the surrounding features in the same sample.

Usage

Arguments

dataset data.frame with the dataset to replace the NA's by the medians of the surrounding

features.

window.size numeric value, specifying of how many features around the NA the median

should be taken.

Details

This function can be used when the dependent dataset in the integrated analysis function is array-CGH data and contains probes that have an NA. To avoid loosing data by throwing away the probes with NA's, the impute nas. by surrounding function can be used which simply takes the median of the probes around an NA. The number of probes used for the imputatin is chosen by giving a value for window size. This script takes quite long to run!

Value

A data.frame is returned, containing the inserted "dataset" all NA replaced with median of the window of size "window.size" around the NA.

Author(s)

Marten Boetzer, Melle Sieswerda, Renee X. de Menezes < R. X. Menezes@lumc.nl>

See Also

SIM, assemble.data, integrated.analysis

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Examples

```
#no examples yet!
```

integrated.analysis

Integrated analysis of dependent and indepedent microarray data

Description

Runs the Integrated Analysis to test for associations between dependent and independent microarray data on the same set of samples.

Usage

Arguments

samples

vector with either the names of the columns in the dependent and independent data corresponding to the samples, or a numerical vector containing the column numbers to include in the analysis, e.g. 5:10 means columns 5 till 10. Make sure that both datasets have the same number of samples with the same column names!

input.regions

vector indicating the dependent regions to be analyzed. Can be defined in four ways: 1) predefined input region: insert a predefined input region, choices are: "all chrs", "all chrs auto", "all arms", "all arms auto" In the predefined regions "all arms" and "all arms auto" the arms 13p, 14p, 15p, 21p and 22p are left out, because in most studies there are no or few probes in these regions. To include them, just make your own vector of arms. 2) whole chromosome(s): insert a single chromosome or a list of chromosomes as a vector: c(1, 2, 3). 3) chromosome arms: insert a single chromosome arm or a list of chromosome arms like c("1q", "2p", "2q"). 4) subregions of a chromosome: insert a chromosome number followed by the start and end position like "chr1:1-1000000" These regions can also be combined, e.g. c("chr1:1-1000000", "2q", 3). See details for more information.

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input.region.indep

indicating the independent region which will be analysed in combination of the dependent region. Only one input region can given using the same format as the dependent input region.

zscores

logical indicates whether the Z-scores are calculated (takes longer time to run). If zscores = FALSE, only P-values are calculated.

method

either one of "full", "window", "overlap" or "smooth". This defines how the data is used for theintegrated.analysis. full: the whole dependent data region is taken. window: takes the middle of the dependent probe and does the integration on the independent probes that are within the window given at window-size given by window. overlap: does the integration on the independent probes that are within the start and end of the dependent probes given at dep. end. smooth: does smooth on the dependent probes with smoothing factor given at smooth. lambda, finds the value of smooth for each independent probe and does the integration on them. Only needed when method = "smooth", de-

fault smooth.lambda = 2

dep.end

numeric or character either the name of the column "end" in the dependent data or, when not available, an numeric value which indicates the end deviating from the start. When a numeric value is inserted, the function will do: start +dep.end = end. Only needed when method = "window" or "overlap".

window

numeric values. Window to search for overlapping independent features per dependent probe. First value is the number of positions to the left from the middle of the probe, the second value is the number of positions to the right from the middle of the probe. Only needed when method = "window".

smooth.lambda

numeric factor used for smoothing the dependent data. Only needed when method = "smooth". See quantsmooth for more information. By default the segment = min(nrow(dep.data), 100).

adjust

formula a formula like ~gender, where gender is a vector of the same size as samples. The regression models is correct for the gender effect, see gt.

run.name

character name of the analysis. The results will be stored in a folder with this name in the current working directory (use getwd() to print the current working directory). If missing the default folder "analysis_results" will be generated.

additional arguments for gt e.g. model="logistic" or when permutations > 0 the null distribution is estimated using permutations, see gt. See Details.

Details

The Integrated Analysis is a regression of the independent data on the dependent features. The regression itself is done using the gt, which means that the genes in a region (e.g. a chromosome arm) are tested as a gene set. The individual associations between each dependent and each independent feature are calculated as Z-scores (standardized influences, see ?gt).

This function splits the datasets into separate sets for each region (as specified by the input.regions) and runs the analysis for each region separately.

When running the Integrated Analysis for a predefined input region, like "all arms" and "all chrs", output can be obtained for all input regions, as well as subsets of it. But note that the genomic 14 integrated.analysis

unit must be the same: if integrated.analysis was run using chromosomes as units, any of the functions and plots must also use chromosomes as units, and not chromosome arms. Similarly, if integrated analysis was run using chromosome arms as units, these units must also be used to produce plots and outputs. For example if the input.regions = "all arms" was used, P-value plots (see sim.plot.pvals.on.region can be produced by inserting the input.regions = "all arms", but also for instance "1p" or "20q". However, to produce a plot of the whole chromosome, for example chromosome 1, the integrated should be re-run with input.region=1. The same goes for "all chrs": P-value plots etc. can be produced for chromosome 1,2 and so on... but to produce plots for an arm, the integrated.analysis should be re-run for that region. This also goes for subregions of the chromosome like "chr1:1-10000000".

By default the gt uses a "linear" model, only when the dependent data is a logical matrix containing TRUE and FALSE a "logistic" model is selected. All other models need model = "", see gt for available models.

Value

No values are returned. Instead, the results of the analysis are stored in the subdirectories of the directory specified in run.name. E.g. the z-score matrices are saved in subfolder method.

The following functions can be used to visualize the data:

1)	sim.plot.zscore.heatmap (only possible when zscores = TRUE)
2)	sim.plot.pyals.on.region

- 3) sim.plot.pvals.on.genome
- 4) sim.plot.overlapping.indep.dep.features

Other functions can be used to tabulate the results:

- 1) tabulate.pvals
- 2) tabulate.top.dep.features
- 3) tabulate.top.indep.features (only possible when zscores = TRUE
- 4) getoverlappingregions (only possible when tablulate.top.dep.features and tabulate.top.indep.features were run.

Author(s)

Marten Boetzer, Melle Sieswerda, Renee X. de Menezes < R. X. Menezes@lumc.nl>

References

Menezes RX, Boetzer M, Sieswerda M, van Ommen GJ, Boer JM (2009). Integrated analysis of DNA copy number and gene expression microarray data using gene sets. *BMC Bioinformatics*, **10**, 203-.

Goeman JJ, van de Geer SA, de Kort F, van Houwelingen HC (2004). A global test for groups of genes: testing association with a clinical outcome. *Bioinformatics*, **20**, 93-109.

See Also

SIM, sim.plot.zscore.heatmap, sim.plot.pvals.on.region, sim.plot.pvals.on.genome, tabulate.pvals, tabulate.top.dep.features, tabulate.top.indep.features, getoverlappingregions, sim.plot.overlapping.indep.dep.features, gt

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Examples

```
#first run example(assemble.data)
data(samples)
#perform integrated analysis without Z-scores using the method = "full"
integrated.analysis(samples=samples,
    input.regions="8q",
    zscores=FALSE,
    method="full",
    run.name="chr8q")
```

link.metadata

Link a metadata annotation file to expression ID

Description

Get annotation out of a AnnotationData package and link them to the expression data using the expression probe ID's

Usage

Arguments

data	data.frame with expression data including an expression probe ID column.
col.ID.link	numeric value, specifying the column of data that contains the ${\rm ID}$ to link with the poslist.
chr	list specifying the metadata annotation of the chromosome location on the genome.
chrloc	list specifying the metadata annotation of the location of the probe on the chromosome.
symbol	list specifying the metadata annotation of the symbol corresponding to the probe.

Details

Often, the annotation for expression array probes lack chromosome position information. Therefore, this function adds the two required columns to run the integrated analysis: "CHROMOSOME" and "STARTPOS". In addition, the optional column, "Symbol" is added.

Value

A data. frame is returned, containing a dataset with annotation columns which can be used forintegrated.analysis.

Author(s)

Marten Boetzer, Melle Sieswerda, Renee x Menezes <R.X.Menezes@lumc.nl>

See Also

RESOURCERER.annotation.to.ID

Examples

```
# first download and install the AnnotationData package for your expression array platform
# for example
## Not run: library(hgu133plus2)
## Not run: expr.data <- link.metadata(data, col.ID.link = 1, chr = as.list(hgu133plus2CHR),
chrloc = as.list(hgu133plus2CHRLOC), symbol = as.list(hgu133plus2SYMBOL))
## End(Not run)</pre>
```

RESOURCERER.annotation.to.ID

Link RESOURCERER annotation file to expression ID

Description

Get annotation out of the RESOURCERER annotation file and link them to expression data with help of expression ID's

Usage

```
RESOURCERER.annotation.to.ID(data, poslist, col.ID.link = 1, col.poslist.link = 1)
```

Arguments

data data frame with expression data including an expression ID column.

poslist data.frame containing the RESOURCERER annotation file

col.ID.link numeric value, specifying the column of data that contains the ID to link with

the poslist.

col.poslist.link

numeric value, specifying the column of poslist that contains the ID to link

with the data.

Details

This function will output the inserted dataset, including the necessary, for integrated.analysis, a nnotation columns: "CHROMOSOME", "STARTPOS" and "Symbol" out of the inserted RESOURCE RER annotation file poslist.

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Value

A data.frame is returned, containing a dataset with annotation columns which can be used for integrated.analysis

Author(s)

Marten Boetzer, Melle Sieswerda, Renee x Menezes < R. X. Menezes@lumc.nl>

See Also

link.metadata

Examples

```
# download expression array annotation from RESOURCERER
# ftp://occams.dfci.harvard.edu/pub/bio/tgi/data/Resourcerer
# it may be necessary to remove the first row, which states the genome build used for mapping
## Not run: read.an <- read.delim("affy_U133Plus2.txt", sep="\t", header=T)

# get physical mapping columns
## Not run: expr.data <- RESOURCERER_annotation_to_ID(data = read.expr, poslist = read.an, col.ID.link = 1, col.pos</pre>
```

samples

Samples for example data

Description

Vector of sample names corresponding to the column headers containing the data in both the copy number (acgh.data) and expression (expr.data) example datasets.

Usage

data(samples)

Format

A character vector.

Source

http://genome-www.stanford.edu/aCGH_breast/data.shtml

References

Pollack JR, Sorlie T, Perou CM, Rees CA, Jeffrey SS, Lonning PE, Tibshirani R, Botstein D, Borresen-Dale AL, Brown PO (2002). Microarray analysis reveals a major direct role of DNA copy number alteration in the transcriptional program of human breast tumors. *Proc Natl Acad Sci USA*. **1**, 99(20), 12963-8.

Examples

```
data(samples)
```

```
sim.plot.overlapping.indep.dep.features
```

P-value plot and mean-zscore plots with indication of overlapping features.

Description

Generates three plots: The first plot contains the P-values along the region, with the cut-off displayed. The second plot contains the mean-zscores along the region, with the cut-offs displayed. The third plots generates the cytobands of the region.

Usage

```
sim.plot.overlapping.indep.dep.features(input.regions,
input.region.indep = NULL,
adjust.method = "BY",
log = FALSE,
significance = 0.2,
max.pow = 5,
z.threshold = c(-3,3),
summarize = c("consecutive", "stretch", "window"),
stretch = 10,
window = 1e6,
percentage = 0.5,
xlim=NULL,
pdf = FALSE,
method = c("full", "smooth", "window", "overlap"),
run.name = "analysis_results", ...)
```

Arguments

input.regions

vector indicating the dependent regions to be analyzed. Can be defined in four ways: 1) predefined input region: insert a predefined input region, choices are: "all chrs", "all chrs auto", "all arms", "all arms auto" In the predefined regions "all arms" and "all arms auto" the arms 13p, 14p, 15p, 21p and 22p are left out, because in most studies there are no or few probes in these regions. To include them, just make your own vector of arms. 2) whole chromosome(s): insert a single chromosome or a list of chromosomes as a vector: c(1, 2, 3). 3) chromosome arms: insert a single chromosome arm or a list of chromosome arms like c("1q", "2p", "2q"). 4) subregions of a chromosome: insert a chromosome number followed by the start and end position like "chr1:1-1000000" These regions can also be combined, e.g. c("chr1:1-1000000", "2q", 3). See integrated.analysis for more information.

input.region.indep

indicating the independent region which will be analysed in combination of the dependent region. Only one input region can given using the same format as the

dependent input region.

adjust.method Method used to adjust the P-values for multiple testing, see p.adjust. Default is

"BY" recommended when copy number is used as dependent data. See SIM for

more information about adjusting P-values.

log logical default log = FALSE, if log = TRUE P-values are plotting on log[10]

scale.

significance The threshold for selecting significant P-values.

max.pow numeric only when log = TRUE scale of the y-axis.

z.threshold Threshold to display a green or red bar in the color bar on top of the heatmap

for independent features with mean z-scores above z. threshold (high positive

association) or below -z. threshold (high negative association).

summarize either one of "consecutive", "stretch", "window" which visualizes the subre-

gions according to the selected summarization a) "consecutive" shows the consecutive significant regions b) "stretch" shows a percentage percentage signif-

icant stretch of size stretch

and c) "window" a percentage percentage significant window of length window

stretch integer length of stretch, default stretch = 10 window integer length of window, default window = 1e6

percentage number between [0,1] given the percentage of significance in either

the "stretch" of "window"

xlim c(min, max) scale of the x-axis. Can be used for zooming in on a region.

pdf logical indicate whether to generate a pdf of the plots in the current working

directory or not.

method this must be the either full, window, overlap or smooth but the data should gen-

erated by the same method in integrated.analysis.

run.name This must be the same a given to integrated.analysis

... not used in this version

Details

details: Cytobands plot adapted from SNPChip

Value

No values are returned. The results are stored in a subdirectory of run. name as pdf.

Author(s)

Marten Boetzer, Melle Sieswerda, Renee X. de Menezes < R. X. Menezes@lumc.nl>

See Also

SIM, tabulate.top.dep.features, tabulate.top.indep.features, getoverlappingregions

Examples

sim.plot.pvals.on.genome

Plot the P-values in whole genome overview

Description

Generates a plot of the analyzed dependent data probe positions and their significance on all chromosomes.

Usage

Arguments

input.regions

vector indicating the dependent regions to be analyzed. Can be defined in four ways: 1) predefined input region: insert a predefined input region, choices are: "all chrs", "all chrs auto", "all arms", "all arms auto" In the predefined regions "all arms" and "all arms auto" the arms 13p, 14p, 15p, 21p and 22p are left out, because in most studies there are no or few probes in these regions. To include them, just make your own vector of arms. 2) whole

significance

adjust.method

method

run.name

pdf

main

ylab

ann

chromosome(s): insert a single chromosome or a list of chromosomes as a vector: c(1, 2, 3). 3) chromosome arms: insert a single chromosome arm or a list of chromosome arms like c("1q", "2p", "2q"). 4) subregions of a chromosome: insert a chromosome number followed by the start and end position like "chr1:1-1000000" These regions can also be combined, e.g. c("chr1:1-1000000", "2q", 3). See integrated.analysis for more information. Two values that categorize the P-values on the selected chromosomes. These margins are indicated by different colors shown in the legend. These values can be defined, e.g. pval.sig = c(0.3, 0.075)Method used to adjust the P-values for multiple testing. see p.adjust Default is "BY" recommended when copy number is used as dependent data. See SIM for more information about adjusting P-values. this must be the either full, window, overlap or smooth but the data should generated by the same method in integrated.analysis. Boolean. Indicate whether to generate a pdf of the plots in the current working directory or not. This must be the same a given to integrated. analysis the usual graphical parameter for the caption of the plot. the usual graphical parameter for the y-axis label of the plot.

Details

Grey vertical lines indicate unsignificant probes on top the significant ones are plotted. A purple dot indicates the centromere and a organe line the input region.

Arguments to be passed to pdf when pdf = TRUE, see Details.

the usual graphical parameter for annotation of the plot.

Sometimes it is useful to make the genome-plot as A4 landscape-format, add the following parameters to the sim.plot.pvals.on.genome(..., paper='a4r', width=0, height=0)

Value

No values are returned. The results are stored in the folder "pvalue.plots" in directory run. name as pdf.

Author(s)

Marten Boetzer, Melle Sieswerda, Renee X. de Menezes < R. X. Menezes@lumc.nl>

See Also

SIM, sim.plot.zscore.heatmap, sim.plot.pvals.on.region

Examples

sim.plot.pvals.on.region

P-value histograms and P-values along the genome per region

Description

Generates two plots of the P-values for an analyzed region. The first plot contains the distribution of the raw P-values and ranked plots of the raw and adjusted P-values. The second plot contains the P-values along the genome of analyzed input regions.

Usage

Arguments

input.regions

vector indicating the dependent regions to be analyzed. Can be defined in four ways: 1) predefined input region: insert a predefined input region, choices are: "all chrs", "all chrs auto", "all arms", "all arms auto" In the predefined regions "all arms" and "all arms auto" the arms 13p, 14p, 15p, 21p and 22p are left out, because in most studies there are no or few probes in these regions. To include them, just make your own vector of arms. 2) whole chromosome(s): insert a single chromosome or a list of chromosomes as a vector: c(1, 2, 3). 3) chromosome arms: insert a single chromosome arm or a list of chromosome arms like c("1q", "2p", "2q"). 4) subregions of a chromosome: insert a chromosome number followed by the start and end position like "chr1:1-1000000" These regions can also be combined, e.g. c("chr1:1-1000000", "2q", 3). See integrated analysis for more information.

 ${\tt significance}$

The threshold for selecting significant P-values.

adjust.method

Method used to adjust the P-values for multiple testing. see p.adjust Default is "BY" recommended when copy number is used as dependent data. See SIM for more information about adjusting P-values.

sim.plot.zoom.in 23

method this must be the either full, window, overlap or smooth but the data should gen-

erated by the same method in integrated.analysis.

run.name This must be the same a given to integrated.analysis

... Arguments to be passed to pdf.

Details

This function returns a pdf containing the P-value plots. The second plot contains the multiple testing corrected P-values plotted along the chromosome (arm). On the x-axis, the start positions of the dependent features are displayed. On the y-axis, the P-value levels are displayed. Two dotted lines indicate P-value levels 0.2 and 0.1. In general, P-values below 0.2 are said to be "significant".

Value

No values are returned. The results are stored in a subdirectory of run. name as pdf.

Author(s)

Marten Boetzer, Melle Sieswerda, Renee X. de Menezes < R. X. Menezes@lumc.nl>

See Also

SIM, sim.plot.pvals.on.genome

Examples

sim.plot.zoom.in

Zoom in on heatmap

Description

Zoom in on pervious produced heatmap by sim.plot.zscore.heatmap

Usage

```
sim.plot.zoom.in(call)
```

Arguments

call

language, function call of sim.plot.zscore.heatmap

Details

sim.plot.zscore.heatmap returns (invisible) the function call and as attribute the local function environment. So by adjusting the the xlim and ylim a zoom in is created.

Value

an additional plot is created, on the previous plot a rectangle indicating the zoomed region.

Author(s)

Marten Boetzer, Melle Sieswerda, Renee X. de Menezes < R. X. Menezes@lumc.nl>

See Also

SIM, sim.plot.zscore.heatmap

Examples

```
\verb|sim.plot.zscore.heatmap| \\
```

Association heatmap from z-scores

Description

Produces an association heatmap that shows the association (standardized influence) of each independent feature (expression measurement) with each dependent feature (copy number measurement). A P-value bar on the left indicates test signficance. A color bar on top indicates genes with mean z-scores across the signficant copy number probes above a set threshold. A summary of the copy number data helps to identify what copy number alterations are present in a region of association with expression. Positive association can mean copy number gain and increased expression, or deletion and decreased expression. The heatmaps can also be used in an exploratory analysis, looking for very local effects of copy number changes (usually small amplifications) on gene expression, that do not lead to a significant test result.

Usage

```
sim.plot.zscore.heatmap(input.regions = "all chrs",
  input.region.indep = NULL,
  method = c("full", "smooth", "window", "overlap"),
  adjust = ~1,
  significance = 0.2,
  z.threshold = 3,
  colRamp = colorRampPalette(c("red", "black", "green")),
  add.colRamp = colorRampPalette(c("blue", "black", "yellow"))(7),
  show.names.indep = FALSE,
  show.names.dep = FALSE,
  adjust.method = "BY",
  scale,
  add.scale,
  add.plot = c("smooth", "none", "heatmap"),
  smooth.lambda = 2,
  pdf = TRUE,
  run.name = "analysis_results",...)
```

Arguments

input.regions

vector indicating the dependent regions to be analyzed. Can be defined in four ways: 1) predefined input region: insert a predefined input region, choices are: "all chrs", "all chrs auto", "all arms", "all arms auto" In the predefined regions "all arms" and "all arms auto" the arms 13p, 14p, 15p, 21p and 22p are left out, because in most studies there are no or few probes in these regions. To include them, just make your own vector of arms. 2) whole chromosome(s): insert a single chromosome or a list of chromosomes as a vector: c(1, 2, 3). 3) chromosome arms: insert a single chromosome arm or a list of chromosome arms like c("1q", "2p", "2q"). 4) subregions of a chromosome: insert a chromosome number followed by the start and end position like "chr1:1-1000000" These regions can also be combined, e.g. c("chr1:1-1000000", "2q", 3). See integrated analysis for more information.

input.region.indep

indicating the independent region which will be analysed in combination of the dependent region. Only one input region can given using the same format as the dependent input region.

method

this must be the either full, window, overlap or smooth but the data should generated by the same method in integrated.analysis.

adjust

This variable must be a vector with the same length as samples or FALSE. The vector will be transformed to a factor and the levels of this will be coloured according to their subtype. When subtype=FALSE, all the samples will be coloured black.

significance

The threshold for selecting significant P-values.

z.threshold

Threshold to display a green or red bar in the color bar on top of the heatmap for independent features with mean z-scores above z.threshold (high positive association) or below -z.threshold (high negative association).

colRamp Palette of colors to be used in the heatmap.

add.colRamp Palette of colors to be used in the added plot.

show.names.indep

logical if set to TRUE, displays the names (indep.id and in dep.symb entered in the assemble.data) of the independent features with mean z-scores above or below the z.threshold in the heatmap.

show.names.dep logical if set to TRUE, displays the names (dep.id and dep.sy mb entered in

the assemble.data) of the significant dependent features in the heatmap.

adjust.method Method used to adjust the P-values for multiple testing, see p.adjust. Default is

"BY" recommended when copy number is used as dependent data. See SIM for

more information about adjusting P-values.

scale Vector specifying the color scale in the heatmap. If scale="auto", the maximum

and minimum value of all z-scores will be calculated and set as the limits for all analyzed regions. Another option is to define a custom scale, e.g. scale =

c(-5,5).

add. scale Vector specifying the color scale in the left plot near the heatmap. If scale="auto",

the maximum and minimum value of all the values will be calculated and set as the limits for all analyzed regions. Another option is to define a custom scale,

e.g. scale = c(-5,5).

add.plot Summary plot of copy number data in left panel. Either "smooth", "heatmap",

or "none". The "smooth" plot smoothes the copy number log ratios per sample, see quantsmooth for more details. The "heatmap" method produces an aCGH heatmap where green indicates gain, and red loss. The scale of the aCGH heatmap is automatically set to the min and max of the aCGH measurements of the analyzed regions. Default is plot.method = "none", no additional plot will

be drawn.

smooth.lambda Numeric value, specifying the quantile smoothing parameter for plot.method="smooth".

See quantsmooth and references for more information.

pdf logical; indicate whether to generate a pdf of the plots in the current working

directory or not.

run.name This must be the same a given to integrated.analysis

... not used in this version

Details

The sim.plot.zscore.heatmap function can only run after the integrated.analysis is run with zscores = TRUE.

The results are returned as a single-page pdf containing an association heatmap of the regions listed in input.regions. For high-density arrays large files will be produced, both demanding more memory available from your computer to produce them as well as being heavier to open on screen. To avoid this, analyze chromosome arms as units instead of chromosomes, both here and in input.regions = "all arms".

The heatmap contains the z-scores generated by the function integrated analysis with zscores=TRUE. The dependent features are plotted from bottom to top, the independent features from left to right. Positive associations are shown in green, negative associations in red (color scale on the right).

At the left side of the heatmap a color bar represents the multiple testing corrected P-values of the probes in the dependent data (copy number), also with a color legend. Dependening on which plot.method is used, a summary of copy number changes is shown on the left. At the top of the heatmap is a color bar corresponding to the mean z-scores of the independent features (expression data) that are above or below the z.threshold. If show.names.indep is set to TRUE, labels will be drawn for the probes with mean z-scores greater than z.threshold or lower than -z.threshold at the bottom of the heatmap. If show.names.dep is set to TRUE, labels will be drawn for the significant dependent probes lower than significance to the right of the heatmap.

Value

No values are returned. The results are stored in a subdirectory of run.name as pdf.

Author(s)

Marten Boetzer, Melle Sieswerda, Renee X. de Menezes < R. X. Menezes@lumc.nl>

References

Eilers PH, de Menezes RX. 2005 Apr 1, Quantile smoothing of array CGH data. *Bioinformatics*, **21**(7):1146-53.

Wang P, Kim Y, Pollack J, Narasimhan B, Tibshirani R. 2005, A method for calling gains and losses in array CGH data. *Biostatistics*, **6**:45-58.

See Also

SIM, tabulate.pvals, tabulate.top.dep.features, tabulate.top.indep.features, sim.plot.overlapping.indep.dep.features

Examples

```
#first run example(assemble.data)
#and example(integrated.analysis)
#plot the zscores in a heatmap
sim.plot.zscore.heatmap(input.regions = "8q", adjust.method = "BY",
                        run.name = "chr8q", pdf = FALSE)
sim.plot.zscore.heatmap(input.regions = "8q",
                        method="full",
                        significance = 0.05,
                        z.threshold = 1,
                        colRamp = colorRampPalette(c("red", "black",
                           "green"))(15),
                        show.names.indep=TRUE,
                        show.names.dep=TRUE,
                        adjust.method = "holm"
                        add.plot = "heatmap",
                        smooth.lambda = 2,
                        pdf = FALSE,
                        run.name = "chr8q")
```

sim.update.chrom.table

Update the chromomosome table

Description

A function to update the genomic positions of chromosome arms. Base locations of the start and end of chromosome arms should be used from the same organism and build of genome as the location provided as annotation with the datasets.

Usage

```
sim.update.chrom.table(db = "homo_sapiens_core_40_36b")
```

Arguments

db

database name

Details

This functions requires library RMySQL. Currently SIM only supports integrated analysis on the human genome without mitochondrial DNA.

Value

Chromosome table chrom.table.

Author(s)

Marten Boetzer, Renee X. de Menezes < R. X. Menezes@lumc.nl>

References

```
http://www.ensembl.org/info/data/mysql.html
```

See Also

SIM, chrom.table

tabulate.pvals 29

Examples

```
#youn need internet connection for this!
#sim.update.chrom.table(db = "homo_sapiens_core_40_36b")
```

tabulate.pvals

Sums significant P-values for the analyzed regions

Description

Generates a data. frame with the significance of P-values in the analyzed regions, dividing them into bins.

Usage

Arguments

input.regions

vector indicating the dependent regions to be analyzed. Can be defined in four ways: 1) predefined input region: insert a predefined input region, choices are: "all chrs", "all chrs auto", "all arms", "all arms auto" In the predefined regions "all arms" and "all arms auto" the arms 13p, 14p, 15p, 21p and 22p are left out, because in most studies there are no or few probes in these regions. To include them, just make your own vector of arms. 2) whole chromosome(s): insert a single chromosome or a list of chromosomes as a vector: c(1, 2, 3). 3) chromosome arms: insert a single chromosome arm or a list of chromosome arms like c("1q", "2p", "2q"). 4) subregions of a chromosome: insert a chromosome number followed by the start and end position like "chr1:1-1000000" These regions can also be combined, e.g. c("chr1:1-1000000", "2q", 3). See integrated analysis for more information.

adjust.method

Method used to adjust the P-values for multiple testing, see p.adjust. Default is "BY" recommended when copy number is used as dependent data. See SIM for more information about adjusting P-values.

bins

vector of significance thresholds. This function will calculate the number of features having a P-value lower than the bin.

significance.idx

Index of "bins" to use when computing the percentage of significant P-values. Defaults to 8 (i.e. the first entry in "bins"), in this case 0.20.

order.by Column used for sorting the table. Defaults to "%" (i.e. the percentage of sig-

nificant p-values).

decreasing Direction used for sorting. Defaults to TRUE (i.e. highest values on top).

method this must be the either full, window, overlap or smooth but the data should gen-

erated by the same method in integrated.analysis.

run.name This must be the same a given to integrated.analysis

Value

Returns a data. frame. Each row corresponds to a chromosome and has as many entries as entries in bins, plus 1. Each entry contains the number of P-values that is smaller or equal to the corresponding entry in bins.

The last entry holds the percentage of P-values that is smaller than or equal to the bin identified by significance.idx.

Author(s)

Marten Boetzer, Melle Sieswerda, Renee X. de Menezes < R. X. Menezes@lumc.nl>

See Also

SIM, tabulate.top.dep.features, tabulate.top.indep.features

Examples

tabulate.top.dep.features

Lists the P-values for the dependent features

Description

Lists the integrated analysis P-values for the dependent features in the analyzed regions, together with the available annotation.

Usage

Arguments

input.regions vector indicating the dependent regions to be analyzed. Can be defined in

four ways: 1) predefined input region: insert a predefined input region, choices are: "all chrs", "all chrs auto", "all arms", "all arms auto" In the predefined regions "all arms" and "all arms auto" the arms 13p, 14p, 15p, 21p and 22p are left out, because in most studies there are no or few probes in these regions. To include them, just make your own vector of arms. 2) whole chromosome(s): insert a single chromosome or a list of chromosomes as a vector: c(1, 2, 3). 3) chromosome arms: insert a single chromosome arm or a list of chromosome arms like c("1q", "2p", "2q"). 4) subregions of a chromosome: insert a chromosome number followed by the start and end posi-

tion like "chr1:1-1000000" These regions can also be combined, e.g. c("chr1:1-1000000", "2q",

3). See integrated.analysis for more information.

adjust.method Method used to adjust the P-values for multiple testing, see p.adjust. Default is

"BY" recommended when copy number is used as dependent data. See SIM for

more information about adjusting P-values.

method this must be the either one of "full", "window", "overlap" or "smooth" but the

data should generated by the same method in integrated.analysis.

significance threshold used to select the significant dependent features. Pvalues below this

threshold will be used to estimate regions.

run.name This must be the same a given to integrated.analysis

Details

Output is a .txt file containing a table with sorted integrated analysis P-values of the dependent features. It includes the ann.dep columns that were read in the assemble.data function. Additionally it returns a .txt file containing the significant P-value rich regions. No P-value rich regions are returned when zscores.diag = "all".

Value

Returns a list of data. frame's for each input region. Significant P-value rich regions are returned as a data. frame. This data.frame can be used as an input for getoverlapping regions. Additionally, the results are stored in a subdirectory of run.name as txt.

Author(s)

Marten Boetzer, Melle Sieswerda, Renee X. de Menezes < R. X. Menezes@lumc.nl>

See Also

SIM, tabulate.pvals, tabulate.top.indep.features

Examples

```
#first run example(assemble.data)
#and example(integrated.analysis)
#get the top dependent features sorted by p-value
```

tabulate.top.indep.features

Lists the mean z-scores for the independent features

Description

Lists the mean z-scores for independent features in the analyzed regions, calculated across the significant dependent features. Gives insight in the expression levels most strongly associated with copy number changes.

Usage

Arguments

input.regions

vector indicating the dependent regions to be analyzed. Can be defined in four ways: 1) predefined input region: insert a predefined input region, choices are: "all chrs", "all chrs auto", "all arms", "all arms auto" In the predefined regions "all arms" and "all arms auto" the arms 13p, 14p, 15p, 21p and 22p are left out, because in most studies there are no or few probes in these regions. To include them, just make your own vector of arms. 2) whole chromosome(s): insert a single chromosome or a list of chromosomes as a vector: c(1, 2, 3). 3) chromosome arms: insert a single chromosome arm or a list of chromosome arms like c("1q", "2p", "2q"). 4) subregions of a chromosome: insert a chromosome number followed by the start and end position like "chr1:1-1000000" These regions can also be combined, e.g. c("chr1:1-1000000", "2q", 3). See integrated analysis for more information.

input.region.indep

fill in

method

this must be the either one of "full", "window", "overlap" or "smooth" but the data should generated by the same method in integrated analysis.

adjust.method Method used to adjust the P-values for multiple testing, see p.adjust. Default is

"BY" recommended when copy number is used as dependent data. See SIM for

more information about adjusting P-values.

significance threshold used to select the significant dependent features. Only the z-scores

with these features are used to calculate the mean z-scores across the indepen-

dent features.

decreasing fill in z.threshold fill in

run.name This must be the same a given to integrated.analysis

Details

tabulate.top.indep.features can only be run after integrated.analysis with zscores = TRUE.

Output is a .txt file containing a table with the mean z-scores of all independent features per analyzed region. It includes the ann. indep columns that were read in the assemble.data function.

Additionally it returns a .txt file containing the significant zscores rich regions.

Depending on the argument "adjust.method", the P-values are first corrected for multiple testing. Next, the z-scores are filtered to include only those entries that correspond to significant (P-value < "significance") dependent features to calculate the mean z-scores.

The dependent table can not be generated for diagonal integrated runs.

Value

Returns a list of data. frame's for each input region. Significant P-value rich regions are returned as a data.frame. This data.frame can be used as an input for getoverlappingregions. Additionally, the results are stored in a subdirectory of run.name as txt.

Author(s)

Marten Boetzer, Melle Sieswerda, Renee X. de Menezes < R. X. Menezes@lumc.nl>

See Also

SIM, tabulate.pvals, tabulate.top.dep.features

Examples

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