

# Package ‘GeoTcgaData’

November 2, 2025

**Type** Package

**Title** Processing Various Types of Data on GEO and TCGA

**Version** 2.11.0

**Description** Gene Expression Omnibus(GEO) and The Cancer Genome Atlas (TCGA) provide us with a wealth of data, such as RNA-seq, DNA Methylation, SNP and Copy number variation data. It's easy to download data from TCGA using the gdc tool, but processing these data into a format suitable for bioinformatics analysis requires more work. This R package was developed to handle these data.

**Depends** R (>= 4.2.0)

**License** Artistic-2.0

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**RoxygenNote** 7.2.3

**Suggests** knitr, rmarkdown, DESeq2, S4Vectors, ChAMP, impute, tidyr, clusterProfiler, org.Hs.eg.db, edgeR, limma, quantreg, minfi, IlluminaHumanMethylation450kanno.ilmn12.hg19, dearseq, NOISeq, testthat (>= 3.0.0), CATT, TCGAbiolinks, enrichplot, GEOquery, BiocGenerics

**VignetteBuilder** knitr

**Imports** utils, data.table, plyr, cqn, topconfects, stats, SummarizedExperiment, methods

**Language** en-US

**URL** <https://github.com/YuLab-SMU/GeoTcgaData>

**BugReports** <https://github.com/YuLab-SMU/GeoTcgaData/issues>

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## Contents

|                                 |    |
|---------------------------------|----|
| array_preprocess . . . . .      | 3  |
| cal_mean_module . . . . .       | 3  |
| cluster_array . . . . .         | 4  |
| combine_pvalue . . . . .        | 5  |
| countToFpk . . . . .            | 5  |
| countToTpm . . . . .            | 6  |
| differential_array . . . . .    | 7  |
| differential_CNV . . . . .      | 8  |
| differential_limma . . . . .    | 9  |
| differential_methy . . . . .    | 10 |
| differential_RNA . . . . .      | 12 |
| differential_SNP . . . . .      | 15 |
| differential_SNP_GEO . . . . .  | 16 |
| differential_SNP_tcga . . . . . | 17 |
| fpkmToTpm . . . . .             | 18 |
| geneExpress . . . . .           | 18 |
| gene_ave . . . . .              | 19 |
| gene_cov . . . . .              | 19 |
| get_geo_array . . . . .         | 20 |
| GSE66705_sample2 . . . . .      | 20 |
| id_conversion_TCGA . . . . .    | 21 |
| kegg_liver . . . . .            | 21 |
| Merge_methy_tcga . . . . .      | 22 |
| module . . . . .                | 22 |
| prepare_chi . . . . .           | 23 |
| profile . . . . .               | 23 |
| repAssign . . . . .             | 24 |
| repRemove . . . . .             | 25 |
| SNP_QC . . . . .                | 25 |
| ventricle . . . . .             | 26 |

|              |           |
|--------------|-----------|
| <b>Index</b> | <b>27</b> |
|--------------|-----------|

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|                  |                                      |
|------------------|--------------------------------------|
| array_preprocess | <i>Preprocess of Microarray data</i> |
|------------------|--------------------------------------|

---

**Description**

Preprocess of Microarray data

**Usage**

```
array_preprocess(x, missing_value = "knn", string = " /// ")
```

**Arguments**

|               |                                                                             |
|---------------|-----------------------------------------------------------------------------|
| x             | matrix of Microarray data, each column is a sample, and each row is a gene. |
| missing_value | Method to impute missing expression data, one of "zero" and "knn".          |
| string        | a string, sep of the gene                                                   |

**Value**

matrix

**Examples**

```
arraylist <- get_geo_array("GSE781")
arraylist <- lapply(arraylist, array_preprocess)
```

---

|                 |                                                       |
|-----------------|-------------------------------------------------------|
| cal_mean_module | <i>Find the mean value of the gene in each module</i> |
|-----------------|-------------------------------------------------------|

---

**Description**

Find the mean value of the gene in each module

**Usage**

```
cal_mean_module(geneExpress, module)
```

**Arguments**

|             |                                                                                                          |
|-------------|----------------------------------------------------------------------------------------------------------|
| geneExpress | a data.frame of gene expression data. Each column is a sample, and each row is a gene.                   |
| module      | a data.frame of two column. The first column is module name, the second column are genes in this module. |

**Value**

a data.frame, means the mean of gene expression value in the same module

**Examples**

```
data(geneExpress)
data(module)
result <- cal_mean_module(geneExpress, module)
```

---

|               |                                          |
|---------------|------------------------------------------|
| cluster_array | <i>cluster probes of Microarray data</i> |
|---------------|------------------------------------------|

---

**Description**

cluster probes of Microarray data

**Usage**

```
cluster_array(x, clusterCutoff = 0.7)
```

**Arguments**

**x** matrix of Microarray data, the first is the name of the gene, and the others are the expression value.

**clusterCutoff** Pearson correlation threshold to cut off the hierarchical tree.

**Value**

data.frame

**Examples**

```
arraylist <- get_geo_array("GSE781")
arraylist <- lapply(arraylist, array_preprocess)
arraylist_cluster <- lapply(arraylist, cluster_array)
```

---

|                |                                                          |
|----------------|----------------------------------------------------------|
| combine_pvalue | <i>combine pvalues of SNP difference analysis result</i> |
|----------------|----------------------------------------------------------|

---

**Description**

combine pvalues of SNP difference analysis result

**Usage**

```
combine_pvalue(snpResult, snp2gene, combineMethod = min)
```

**Arguments**

|               |                                                           |
|---------------|-----------------------------------------------------------|
| snpResult     | data.frame of SNP difference analysis result.             |
| snp2gene      | data frame of two column: snp and gene.                   |
| combineMethod | Method of combining the pvalue of multiple snp in a gene. |

**Value**

data.frame

**Examples**

```
snpResult <- data.frame(pvalue = runif(100), estimate = runif(100))
rownames(snpResult) <- paste0("snp", seq_len(100))
snp2gene <- data.frame(snp = rownames(snpResult),
  gene = rep(paste0("gene", seq_len(20)), 5))
result <- combine_pvalue(snpResult, snp2gene)
```

---

|             |                              |
|-------------|------------------------------|
| countToFpkm | <i>Convert count to FPKM</i> |
|-------------|------------------------------|

---

**Description**

Convert count to FPKM

**Usage**

```
countToFpkm(counts_matrix, keyType = "SYMBOL", gene_cov)
```

**Arguments**

|               |                                                                                                 |
|---------------|-------------------------------------------------------------------------------------------------|
| counts_matrix | a matrix, colnames of counts_matrix are sample name, rownames of counts_matrix are gene symbols |
| keyType       | keyType, one of keytypes(org.Hs.eg.db).                                                         |
| gene_cov      | data.frame of two column, the first column is gene length, the second column is gene GC content |

Value

a matrix

Examples

```
data(gene_cov)
lung_squ_count2 <- matrix(c(1, 2, 3, 4, 5, 6, 7, 8, 9), ncol = 3)
rownames(lung_squ_count2) <- c("DISC1", "TCOF1", "SPPL3")
colnames(lung_squ_count2) <- c("sample1", "sample2", "sample3")
result <- countToFpm(lung_squ_count2,
  keyType = "SYMBOL",
  gene_cov = gene_cov
)
```

---

|            |                             |
|------------|-----------------------------|
| countToTpm | <i>Convert count to Tpm</i> |
|------------|-----------------------------|

---

Description

Convert count to Tpm

Usage

```
countToTpm(counts_matrix, keyType = "SYMBOL", gene_cov)
```

Arguments

- counts\_matrix    a matrix, colnames of counts\_matrix are sample name, rownames of counts\_matrix are gene symbols
- keyType            keyType, one of keytypes(org.Hs.eg.db).
- gene\_cov           data.frame of two column, the first column is gene length, the second column is gene GC content

Value

a matrix

Examples

```
data(gene_cov)
lung_squ_count2 <- matrix(c(1, 2, 3, 4, 5, 6, 7, 8, 9), ncol = 3)
rownames(lung_squ_count2) <- c("DISC1", "TCOF1", "SPPL3")
colnames(lung_squ_count2) <- c("sample1", "sample2", "sample3")
result <- countToTpm(lung_squ_count2,
  keyType = "SYMBOL",
  gene_cov = gene_cov
)
```

---

differential\_array      *Differential analysis of Microarray data*


---

## Description

Differential analysis of Microarray data

## Usage

```
differential_array(df, group, method = "limma", adjust.method = "BH")
```

## Arguments

|               |                                                                                                  |
|---------------|--------------------------------------------------------------------------------------------------|
| df            | data.frame of the omic data, each column is a sample, and each row is a gene.                    |
| group         | a vector, group of samples.                                                                      |
| method        | method to do differential analysis, one of "limma", "ttest", "wilcox".                           |
| adjust.method | adjust.method, one of "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", and "none". |

## Value

data.frame

## Examples

```
library(GeoTcgaData)
library(data.table)
# Use real GEO data as example
arrayData <- read.table("GSE54807_series_matrix.txt.gz",
  sep = "\t", header = TRUE,
  fill=TRUE, comment.char = "!", check.names=FALSE)
gpl <- fread("GPL6244-17930.txt", sep = "\t", header = TRUE)
gpl <- gpl[, c("ID", "gene_assignment")]
class(gpl) <- "data.frame"

for (i in seq_len(nrow(gpl))) {
  aa <- strsplit(gpl[i, 2], " // ")[[1]][5]
  gpl[i, 2] <- as.character(strsplit(aa, " /// ")[[1]][1])
}
gpl[,1] <- as.character(gpl[,1])
arrayData[, 1] <- as.character(arrayData[, 1])
rownames(gpl) <- gpl[, 1]
arrayData[, 1] <- gpl[arrayData[, 1], 2]

arrayData <- repRemove(arrayData," /// ")

# Remove rows that do not correspond to genes
```

```

arrayData <- arrayData[!is.na(arrayData[, 1]), ]
arrayData <- arrayData[!arrayData[, 1] == "", ]
arrayData <- arrayData[!arrayData[, 1] == "---", ]

arrayData <- arrayData[order(arrayData[, 1]), ]
arrayData <- gene_ave(arrayData, 1)

keep <- apply(arrayData, 1, function(x) sum(x < 1) < (length(x)/2))
arrayData <- arrayData[keep, ]

group <- c(rep("group1", 12), rep("group2", 12))
result <- differential_array(df = arrayData, group = group)

# Use random data as example
arrayData <- matrix(runif(200), 25, 8)
rownames(arrayData) <- paste0("gene", 1:25)
colnames(arrayData) <- paste0("sample", 1:8)
group <- c(rep("group1", 4), rep("group2", 4))
names(group) <- colnames(arrayData)
result <- differential_array(df = arrayData, group = group)

```

---

differential\_CNV

*Do difference analysis of gene level copy number variation data*


---

## Description

Do difference analysis of gene level copy number variation data

## Usage

```

differential_CNV(
  cnvData,
  sampleGroup,
  method = "Chisquare",
  adjust.method = "BH",
  ...
)

```

## Arguments

|               |                                                                                                          |
|---------------|----------------------------------------------------------------------------------------------------------|
| cnvData       | data.frame of CNV data, each column is a sample, and each row is a CNV.                                  |
| sampleGroup   | vector of sample group                                                                                   |
| method        | method to do diffenental analysis, one of "Chisquare", "fisher", and "CATT"(Cochran-Armitage trend test) |
| adjust.method | adjust.method, one of "holm", "hochberg", "hommel", "bonferroni", "BH", "BY", "fdr", and "none".         |
| ...           | parameters for "Chisquare", "fisher", and "CATT"(Cochran-Armitage trend test)                            |

**Value**

data.frame with pvalue and estimate

**Examples**

```
# use TCGAbiolinks data as example
library(TCGAbiolinks)
query <- GDCquery(
  project = "TCGA-ACC",
  data.category = "Copy Number Variation",
  data.type = "Gene Level Copy Number",
  access = "open"
)
GDCdownload(query)
cnvData <- GDCprepare(query)
aa <- assays(cnvData)$copy_number
bb <- aa
aa[bb == 2] <- 0
aa[bb < 2] <- -1
aa[bb > 2] <- 1
sampleGroup <- sample(c("A", "B"), ncol(cnvData), replace = TRUE)
diffCnv <- differential_CNV(aa, sampleGroup)

# Use sangerbox CNV data as example
cnvData <- fread("Merge_GeneLevelCopyNumber.txt")
class(cnvData) <- "data.frame"
rownames(cnvData) <- cnvData[, 1]
cnvData <- cnvData[, -c(1, 2, 3)]
sampleGroup <- sample(c("A", "B"), ncol(cnvData), replace = TRUE)
diffCnv <- differential_CNV(cnvData, sampleGroup)

# use random data as example
aa <- matrix(sample(c(0, 1, -1), 200, replace = TRUE), 25, 8)
rownames(aa) <- paste0("gene", 1:25)
colnames(aa) <- paste0("sample", 1:8)
sampleGroup <- sample(c("A", "B"), ncol(aa), replace = TRUE)
diffCnv <- differential_CNV(aa, sampleGroup)
```

---

differential\_limma      *differential\_limma*

---

**Description**

differential\_limma

**Usage**

```
differential_limma(df, group, adjust.method = "BH")
```

**Arguments**

df                      data.frame of the omic data

group                  a vector, group of samples.

adjust.method        adjust.method.

**Value**

data.frame

**Examples**

```
df <- matrix(runif(200), 25, 8)
df <- as.data.frame(df)
rownames(df) <- paste0("gene", 1:25)
colnames(df) <- paste0("sample", 1:8)
group <- sample(c("group1", "group2"), 8, replace = TRUE)
result <- differential_limma(df = df, group = group)
```

---

differential\_methy      *differential\_methy*

---

**Description**

Get methylation difference gene

**Usage**

```
differential_methy(
  cpaData,
  sampleGroup,
  groupCol,
  combineMethod = "stouffer",
  missing_value = "knn",
  cpg2gene = NULL,
  normMethod = "PBC",
  region = "TSS1500",
  model = "gene",
  adjust.method = "BH",
  adjPvalCutoff = 0.05,
  ucscData = FALSE
)
```

**Arguments**

|               |                                                                                                                                                                               |
|---------------|-------------------------------------------------------------------------------------------------------------------------------------------------------------------------------|
| cpgData       | data.frame of cpg beta value, , or SummarizedExperiment object                                                                                                                |
| sampleGroup   | vector of sample group                                                                                                                                                        |
| groupCol      | group column                                                                                                                                                                  |
| combineMethod | method to combine the cpg pvalues, a function or one of "stouffer", "fisher" and "rhoScores".                                                                                 |
| missing_value | Method to impute missing expression data, one of "zero" and "knn".                                                                                                            |
| cpg2gene      | data.frame to annotate cpg locus to gene                                                                                                                                      |
| normMethod    | Method to do normalization: "PBC" or "BMIQ".                                                                                                                                  |
| region        | region of genes, one of "Body", "TSS1500", "TSS200", "3'UTR", "1stExon", "5'UTR", and "IGR". Only used when cpg2gene is NULL.                                                 |
| model         | if "cpg", step1: calculate difference cpgs; step2: calculate difference genes. if "gene", step1: calculate the methylation level of genes; step2: calculate difference genes. |
| adjust.method | character string specifying the method used to adjust p-values for multiple testing. See <a href="#">p.adjust</a> for possible values.                                        |
| adjPvalCutoff | adjusted pvalue cutoff                                                                                                                                                        |
| ucscData      | Logical, whether the data comes from UCSC Xena.                                                                                                                               |

**Value**

data.frame

**Examples**

```
# use TCGAbiolinks data
library(TCGAbiolinks)
query <- GDCquery(project = "TCGA-ACC",
  data.category = "DNA Methylation",
  data.type = "Methylation Beta Value",
  platform = "Illumina Human Methylation 450")
GDCdownload(query, method = "api", files.per.chunk = 5,
  directory = Your_Path)
merge_result <- Merge_methy_tcga(Your_Path_to_DNA_Methylation_data)
library(ChAMP) # To avoid reporting errors
differential_gene <- differential_methy(cpgData = merge_result,
  sampleGroup = sample(c("C","T"),
  ncol(merge_result[[1]]), replace = TRUE))

# use user defined data
library(ChAMP)
cpgData <- matrix(runif(2000), nrow = 200, ncol = 10)
rownames(cpgData) <- paste0("cpg", seq_len(200))
colnames(cpgData) <- paste0("sample", seq_len(10))
sampleGroup <- c(rep("group1", 5), rep("group2", 5))
names(sampleGroup) <- colnames(cpgData)
cpg2gene <- data.frame(cpg = rownames(cpgData),
```

```

    gene = rep(paste0("gene", seq_len(20)), 10))
result <- differential_methy(cpgData, sampleGroup,
    cpg2gene = cpg2gene, normMethod = NULL)
# use SummarizedExperiment object input
library(ChAMP)
cpgData <- matrix(runif(2000), nrow = 200, ncol = 10)
rownames(cpgData) <- paste0("cpg", seq_len(200))
colnames(cpgData) <- paste0("sample", seq_len(10))
sampleGroup <- c(rep("group1", 5), rep("group2", 5))
names(sampleGroup) <- colnames(cpgData)
cpg2gene <- data.frame(cpg = rownames(cpgData),
    gene = rep(paste0("gene", seq_len(20)), 10))
colData <- S4Vectors::DataFrame(
    row.names = colnames(cpgData),
    group = sampleGroup
)
data <- SummarizedExperiment::SummarizedExperiment(
    assays=S4Vectors::SimpleList(counts=cpgData),
    colData = colData)
result <- differential_methy(cpgData = data,
    groupCol = "group", normMethod = NULL,
    cpg2gene = cpg2gene)

```

---

|                  |                         |
|------------------|-------------------------|
| differential_RNA | <i>differential_RNA</i> |
|------------------|-------------------------|

---

## Description

Do difference analysis of RNA-seq data

## Usage

```

differential_RNA(
  counts,
  group,
  groupCol,
  method = "limma",
  geneLength = NULL,
  gccontent = NULL,
  filter = TRUE,
  edgeRNorm = TRUE,
  adjust.method = "BH",
  useTopconfects = TRUE,
  ucscData = FALSE
)

```

**Arguments**

|                |                                                                                                                                        |
|----------------|----------------------------------------------------------------------------------------------------------------------------------------|
| counts         | a dataframe or numeric matrix of raw counts data, or SummarizedExperiment object                                                       |
| group          | sample groups                                                                                                                          |
| groupCol       | group column                                                                                                                           |
| method         | one of "DESeq2", "edgeR" , "limma", "dearseq", "NOISeq", "Wilcoxon", and "auto".                                                       |
| geneLength     | a vector of gene length.                                                                                                               |
| gccontent      | a vector of gene GC content.                                                                                                           |
| filter         | if TRUE, use filterByExpr to filter genes.                                                                                             |
| edgeRNorm      | if TRUE, use edgeR to do normalization for dearseq method.                                                                             |
| adjust.method  | character string specifying the method used to adjust p-values for multiple testing. See <a href="#">p.adjust</a> for possible values. |
| useTopconfects | if TRUE, use topconfects to provide a more biologically useful ranked gene list.                                                       |
| ucscData       | Logical, whether the data comes from UCSC Xena.                                                                                        |

**Value**

data.frame

**Examples**

```
library(TCGAbiolinks)

query <- GDCquery(
  project = "TCGA-ACC",
  data.category = "Transcriptome Profiling",
  data.type = "Gene Expression Quantification",
  workflow.type = "STAR - Counts"
)

GDCdownload(query,
  method = "api", files.per.chunk = 3,
  directory = Your_Path
)

dataRNA <- GDCprepare(
  query = query, directory = Your_Path,
  save = TRUE, save.filename = "dataRNA.RData"
)
## get raw count matrix
dataPrep <- TCGAanalyze_Preprocessing(
  object = dataRNA,
  cor.cut = 0.6,
  datatype = "STAR - Counts"
)
```

```

# Use `differential_RNA` to do difference analysis.
# We provide the data of human gene length and GC content in `gene_cov`.
group <- sample(c("grp1", "grp2"), ncol(dataPrep), replace = TRUE)
library(cqn) # To avoid reporting errors: there is no function "rq"
## get gene length and GC content
library(org.Hs.eg.db)
genes_bitr <- bitr(rownames(gene_cov),
  fromType = "ENTREZID", toType = "ENSEMBL",
  OrgDb = org.Hs.eg.db, drop = TRUE
)
genes_bitr <- genes_bitr[!duplicated(genes_bitr[, 2]), ]
gene_cov2 <- gene_cov[genes_bitr$ENTREZID, ]
rownames(gene_cov2) <- genes_bitr$ENSEMBL
genes <- intersect(rownames(dataPrep), rownames(gene_cov2))
dataPrep <- dataPrep[genes, ]
geneLength <- gene_cov2(genes, "length")
gccontent <- gene_cov2(genes, "GC")
names(geneLength) <- names(gccontent) <- genes
## Difference analysis
DEGall <- differential_RNA(
  counts = dataPrep, group = group,
  geneLength = geneLength, gccontent = gccontent
)
# Use `clusterProfiler` to do enrichment analytics:
diffGenes <- DEGall$logFC
names(diffGenes) <- rownames(DEGall)
diffGenes <- sort(diffGenes, decreasing = TRUE)
library(clusterProfiler)
library(enrichplot)
library(org.Hs.eg.db)
gsego <- gseGO(gene = diffGenes, OrgDb = org.Hs.eg.db, keyType = "ENSEMBL")
dotplot(gsego)

# use user-defined data
df <- matrix(rnbinom(400, mu = 4, size = 10), 25, 16)
df <- as.data.frame(df)
rownames(df) <- paste0("gene", 1:25)
colnames(df) <- paste0("sample", 1:16)
group <- sample(c("group1", "group2"), 16, replace = TRUE)
result <- differential_RNA(counts = df, group = group,
  filte = FALSE, method = "Wilcoxon")
# use SummarizedExperiment object input
df <- matrix(rnbinom(400, mu = 4, size = 10), 25, 16)
rownames(df) <- paste0("gene", 1:25)
colnames(df) <- paste0("sample", 1:16)
group <- sample(c("group1", "group2"), 16, replace = TRUE)

nrows <- 200; ncols <- 20
counts <- matrix(
  runif(nrows * ncols, 1, 1e4), nrows,
  dimnames = list(paste0("cg", 1:200), paste0("S", 1:20))
)

```

```
colData <- S4Vectors::DataFrame(
  row.names = paste0("sample", 1:16),
  group = group
)
data <- SummarizedExperiment::SummarizedExperiment(
  assays=S4Vectors::SimpleList(counts=df),
  colData = colData)

result <- differential_RNA(counts = data, groupCol = "group",
  filte = FALSE, method = "Wilcoxon")
```

---

|                  |                                           |
|------------------|-------------------------------------------|
| differential_SNP | <i>Do difference analysis of SNP data</i> |
|------------------|-------------------------------------------|

---

## Description

Do difference analysis of SNP data

## Usage

```
differential_SNP(snpDf, sampleGroup, combineMethod = min)
```

## Arguments

|               |                                                                         |
|---------------|-------------------------------------------------------------------------|
| snpDf         | data.frame of SNP data, each column is a sample, and each row is a SNP. |
| sampleGroup   | vector of sample group.                                                 |
| combineMethod | Method of combining the pvalue of multiple snp in a gene.               |

## Value

data.frame

## Examples

```
library(TCGAbiolinks)
query <- GDCquery(
  project = "TCGA-CHOL",
  data.category = "Simple Nucleotide Variation",
  access = "open",
  legacy = FALSE,
  data.type = "Masked Somatic Mutation",
  workflow.type = "Aliquot Ensemble Somatic Variant Merging and Masking"
)
GDCdownload(query)
data_snp <- GDCprepare(query)
samples <- unique(data_snp$Tumor_Sample_Barcode)
sampleGroup <- sample(c("A", "B"), length(samples), replace = TRUE)
names(sampleGroup) <- samples
pvalue <- differential_SNP_tcga(snpData = data_snp,
```

```

sampleGroup = sampleGroup)

# use demo data
snpDf <- matrix(sample(c("mutation", NA), 100, replace = TRUE), 10, 10)
snpDf <- as.data.frame(snpDf)
sampleGroup <- sample(c("A", "B"), 10, replace = TRUE)
result <- differential_SNP(snpDf, sampleGroup)

```

---

differential\_SNP\_GEO    *Do difference analysis of SNP data downloaded from GEO*

---

## Description

Do difference analysis of SNP data downloaded from GEO

## Usage

```
differential_SNP_GEO(snpData, sampleGroup, method = "Chisquare")
```

## Arguments

|             |                                                                       |
|-------------|-----------------------------------------------------------------------|
| snpData     | data.frame of SNP data downloaded from GEO                            |
| sampleGroup | vector of sample group                                                |
| method      | one of "Chisquare", "fisher", and "CATT"(Cochran-Armitage trend test) |

## Value

data.frame

## Examples

```

file1 <- read.table("GSE66903_series_matrix.txt.gz",
  fill=TRUE, comment.char="!", header = TRUE)
rownames(file1) <- file1[, 1]
snpData <- file1[, -1]
sampleGroup <- sample(c("A", "B"), ncol(snpData ), replace = TRUE)
names(sampleGroup) <- colnames(snpData)
snpData <- SNP_QC(snpData)
sampleGroup <- sample(c("A", "B"), ncol(snpData ), replace = TRUE)
result1 <- differential_SNP_GEO(snpData = snpData,
  sampleGroup = sampleGroup, method = "Chisquare")

# use demo data
snpDf <- matrix(sample(c("AA", "Aa", "aa"), 100, replace = TRUE), 10, 10)
snpDf <- as.data.frame(snpDf)
sampleGroup <- sample(c("A", "B"), 10, replace = TRUE)
result <- differential_SNP_GEO(snpDf, sampleGroup, method = "fisher")

```

---

differential\_SNP\_tcga *Do difference analysis of SNP data downloaded from TCGAbiolinks*

---

## Description

Do difference analysis of SNP data downloaded from TCGAbiolinks

## Usage

```
differential_SNP_tcga(snpData, sampleGroup, combineMethod = NULL)
```

## Arguments

|               |                                                           |
|---------------|-----------------------------------------------------------|
| snpData       | data.frame of SNP data downloaded from TCGAbiolinks       |
| sampleGroup   | vector of sample group                                    |
| combineMethod | Method of combining the pvalue of multiple snp in a gene. |

## Value

data.frame

## Examples

```
library(TCGAbiolinks)
query <- GDCquery(
  project = "TCGA-CHOL",
  data.category = "Simple Nucleotide Variation",
  access = "open",
  legacy = FALSE,
  data.type = "Masked Somatic Mutation",
  workflow.type = "Aliquot Ensemble Somatic Variant Merging and Masking"
)
GDCdownload(query)
data_snp <- GDCprepare(query)
samples <- unique(data_snp$Tumor_Sample_Barcode)
sampleGroup <- sample(c("A", "B"), length(samples), replace = TRUE)
names(sampleGroup) <- samples
pvalue <- differential_SNP_tcga(snpData = data_snp,
  sampleGroup = sampleGroup)

# use demo data
snpDf <- matrix(sample(c("mutation", NA), 100, replace = TRUE), 10, 10)
snpDf <- as.data.frame(snpDf)
sampleGroup <- sample(c("A", "B"), 10, replace = TRUE)
result <- differential_SNP(snpDf, sampleGroup)
```

---

|           |                            |
|-----------|----------------------------|
| fpkmToTpm | <i>Convert fpkm to Tpm</i> |
|-----------|----------------------------|

---

**Description**

Convert fpkm to Tpm

**Usage**

```
fpkmToTpm(fpkm_matrix)
```

**Arguments**

fpkm\_matrix      a matrix, colnames of fpkm\_matrix are sample name, rownames of fpkm\_matrix are genes

**Value**

a matrix

**Examples**

```
lung_squ_count2 <- matrix(c(0.11, 0.22, 0.43, 0.14, 0.875,
  0.66, 0.77, 0.18, 0.29), ncol = 3)
rownames(lung_squ_count2) <- c("DISC1", "TCOF1", "SPPL3")
colnames(lung_squ_count2) <- c("sample1", "sample2", "sample3")
result <- fpkmToTpm(lung_squ_count2)
```

---

|             |                                             |
|-------------|---------------------------------------------|
| geneExpress | <i>a data.frame of gene expression data</i> |
|-------------|---------------------------------------------|

---

**Description**

It is a randomly generated expression data used as an example of functions in this package. the rowname is gene symbols the columns are gene expression values

**Usage**

```
geneExpress
```

**Format**

A data.frame with 10779 rows and 2 column

---

|          |                                                                    |
|----------|--------------------------------------------------------------------|
| gene_ave | <i>Average the values of same genes in gene expression profile</i> |
|----------|--------------------------------------------------------------------|

---

**Description**

Average the values of same genes in gene expression profile

**Usage**

```
gene_ave(file_gene_ave, k = 1)
```

**Arguments**

file\_gene\_ave    a data.frame of gene expression data, each column is a sample, and each row is a gene.

k                a number, indicates which is the gene column.

**Value**

a data.frame, the values of same genes in gene expression profile

**Examples**

```
aa <- c("MARCH1", "MARCH1", "MARCH1", "MARCH1", "MARCH1")
bb <- c(2.969058399, 4.722410064, 8.165514853, 8.24243893, 8.60815086)
cc <- c(3.969058399, 5.722410064, 7.165514853, 6.24243893, 7.60815086)
file_gene_ave <- data.frame(aa = aa, bb = bb, cc = cc)
colnames(file_gene_ave) <- c("Gene", "GSM1629982", "GSM1629983")

result <- gene_ave(file_gene_ave, 1)
```

---

|          |                                                   |
|----------|---------------------------------------------------|
| gene_cov | <i>a data.frame of gene length and GC content</i> |
|----------|---------------------------------------------------|

---

**Description**

the gene length and GC content data comes from TxDb.Hsapiens.UCSC.hg38.knownGene and BSgenome.Hsapiens.UCSC.hg38

**Usage**

```
gene_cov
```

**Format**

A data.frame with 27341 rows and 2 column

---

|               |                                            |
|---------------|--------------------------------------------|
| get_geo_array | <i>Get Microarray matrix data from GEO</i> |
|---------------|--------------------------------------------|

---

**Description**

Get Microarray matrix data from GEO

**Usage**

```
get_geo_array(gse)
```

**Arguments**

gse                    GSE number, such as GSE781.

**Value**

a list of matrix

**Examples**

```
arraylist <- get_geo_array("GSE781")
```

---

|                  |                                                |
|------------------|------------------------------------------------|
| GSE66705_sample2 | <i>a matrix of gene expression data in GEO</i> |
|------------------|------------------------------------------------|

---

**Description**

the first column represents the gene symbol

**Usage**

```
GSE66705_sample2
```

**Format**

A matrix with 999 rows and 3 column

**Details**

the other columns represent the expression of genes

---

|                    |                                                       |
|--------------------|-------------------------------------------------------|
| id_conversion_TCGA | <i>Convert ENSEMBL gene id to gene Symbol in TCGA</i> |
|--------------------|-------------------------------------------------------|

---

**Description**

Convert ENSEMBL gene id to gene Symbol in TCGA

**Usage**

```
id_conversion_TCGA(profiles, toType = "SYMBOL")
```

**Arguments**

|          |                                                                                        |
|----------|----------------------------------------------------------------------------------------|
| profiles | a data.frame of gene expression data, each column is a sample, and each row is a gene. |
| toType   | one of 'keytypes(org.Hs.eg.db)'                                                        |

**Value**

a data.frame, gene symbols and their expression value

**Examples**

```
library(org.Hs.eg.db)
data(profile)
result <- id_conversion_TCGA(profile)
```

---

|            |                                                 |
|------------|-------------------------------------------------|
| kegg_liver | <i>a matrix of gene expression data in TCGA</i> |
|------------|-------------------------------------------------|

---

**Description**

It is a randomly generated expression data used as an example of functions in this package. the first column represents the gene symbol

**Usage**

```
kegg_liver
```

**Format**

A matrix with 100 rows and 150 column

**Details**

the other columns represent the expression(count) of genes

---

|                  |                                                    |
|------------------|----------------------------------------------------|
| Merge_methy_tcga | <i>Merge methylation data downloaded from TCGA</i> |
|------------------|----------------------------------------------------|

---

### Description

When the methylation data is downloaded from TCGA, each sample is saved in a folder, which contains the methylation value file and the descriptive file. This function can directly extract and consolidate all folders.

### Usage

```
Merge_methy_tcga(dirr = NULL)
```

### Arguments

|      |                                                                                       |
|------|---------------------------------------------------------------------------------------|
| dirr | a string for the directory of methylation data download from tcga using the tools gdc |
|------|---------------------------------------------------------------------------------------|

### Value

a matrix, a combined methylation expression spectrum matrix

### Examples

```
merge_result <- Merge_methy_tcga(system.file(file.path("extdata", "methy"),
  package = "GeoTcgaData"))
```

---

|        |                                                                              |
|--------|------------------------------------------------------------------------------|
| module | <i>a matrix of module name, gene symbols, and the number of gene symbols</i> |
|--------|------------------------------------------------------------------------------|

---

### Description

It is a randomly generated expression data used as an example of functions in this package.

### Usage

```
module
```

### Format

A matrix with 176 rows and 3 column

---

|             |                                          |
|-------------|------------------------------------------|
| prepare_chi | <i>Preparer file for chi-square test</i> |
|-------------|------------------------------------------|

---

**Description**

Preparer file for chi-square test

**Usage**

```
prepare_chi(cnv)
```

**Arguments**

cnv                      result of ann\_merge()

**Value**

a matrix

**Examples**

```
cnv <- matrix(c(
  -1.09150, -1.47120, -0.87050, -0.50880,
  -0.50880, 2.0, 2.0, 2.0, 2.0, 2.0, 2.601962, 2.621332, 2.621332,
  2.621332, 2.621332, 2.0, 2.0, 2.0, 2.0, 2.0, 2.0, 2.0, 2.0,
  2.0, 2.0, 2.0, 2.0, 2.0, 2.0, 2.0),
  nrow = 5)
cnv <- as.data.frame(cnv)
rownames(cnv) <- c("AJAP1", "FHAD1", "CLCNKB", "CROCCP2", "AL137798.3")
colnames(cnv) <- c(
  "TCGA-DD-A4NS-10A-01D-A30U-01", "TCGA-ED-A82E-01A-11D-A34Y-01",
  "TCGA-WQ-A9G7-01A-11D-A36W-01", "TCGA-DD-AADN-01A-11D-A40Q-01",
  "TCGA-ZS-A9CD-10A-01D-A36Z-01", "TCGA-DD-A1EB-11A-11D-A12Y-01"
)
cnv_chi_file <- prepare_chi(cnv)
```

---

|         |                                                 |
|---------|-------------------------------------------------|
| profile | <i>a matrix of gene expression data in TCGA</i> |
|---------|-------------------------------------------------|

---

**Description**

It is a randomly generated expression data used as an example of functions in this package. the first column represents the gene symbol

**Usage**

```
profile
```

**Format**

A matrix with 10 rows and 10 column

**Details**

the other columns represent the expression(FPKM) of genes

---

|           |                                                                   |
|-----------|-------------------------------------------------------------------|
| repAssign | <i>Handle the case where one id corresponds to multiple genes</i> |
|-----------|-------------------------------------------------------------------|

---

**Description**

Handle the case where one id corresponds to multiple genes

**Usage**

```
repAssign(input_file, string)
```

**Arguments**

|            |                                                                         |
|------------|-------------------------------------------------------------------------|
| input_file | input file, a data.frame or a matrix, the first column should be genes. |
| string     | a string, sep of the gene                                               |

**Value**

a data.frame, when an id corresponds to multiple genes, the expression value is assigned to each gene

**Examples**

```
aa <- c("MARCH1 /// MMA", "MARC1", "MARCH2 /// MARCH3",
        "MARCH3 /// MARCH4", "MARCH1")
bb <- c("2.969058399", "4.722410064", "8.165514853",
        "8.24243893", "8.60815086")
cc <- c("3.969058399", "5.722410064", "7.165514853",
        "6.24243893", "7.60815086")
input_file <- data.frame(aa = aa, bb = bb, cc = cc)

repAssign_result <- repAssign(input_file, " /// ")
```

---

|           |                                                                   |
|-----------|-------------------------------------------------------------------|
| repRemove | <i>Handle the case where one id corresponds to multiple genes</i> |
|-----------|-------------------------------------------------------------------|

---

**Description**

Handle the case where one id corresponds to multiple genes

**Usage**

```
repRemove(input_file, string)
```

**Arguments**

|            |                                                                         |
|------------|-------------------------------------------------------------------------|
| input_file | input file, a data.frame or a matrix, the first column should be genes. |
| string     | a string, sep of the gene                                               |

**Value**

a data.frame, when an id corresponds to multiple genes, the expression value is deleted

**Examples**

```
aa <- c("MARCH1 /// MMA", "MARC1", "MARCH2 /// MARCH3",
        "MARCH3 /// MARCH4", "MARCH1")
bb <- c("2.969058399", "4.722410064", "8.165514853",
        "8.24243893", "8.60815086")
cc <- c("3.969058399", "5.722410064", "7.165514853",
        "6.24243893", "7.60815086")
input_file <- data.frame(aa = aa, bb = bb, cc = cc)
repRemove_result <- repRemove(input_file, " /// ")
```

---

|        |                                                                    |
|--------|--------------------------------------------------------------------|
| SNP_QC | <i>Do quality control of SNP data downloaded from TCGAbiolinks</i> |
|--------|--------------------------------------------------------------------|

---

**Description**

Do quality control of SNP data downloaded from TCGAbiolinks

**Usage**

```
SNP_QC(
  snpData,
  geon = 0.02,
  mind = 0.02,
  maf = 0.05,
  hwe = 1e-06,
  miss = "NoCall"
)
```

Arguments

|         |                                                                                                                |
|---------|----------------------------------------------------------------------------------------------------------------|
| snpData | data.frame of SNP data downloaded from TCGAbiolinks                                                            |
| geon    | filters out all variants with missing call rates exceeding the provided value (default 0.02) to be removed     |
| mind    | filters out all samples with missing call rates exceeding the provided value (default 0.02) to be removed      |
| maf     | filters out all variants with minor allele frequency below the provided threshold                              |
| hwe     | filters out all variants which have Hardy-Weinberg equilibrium exact test p-value below the provided threshold |
| miss    | character of miss value                                                                                        |

Value

data.frame

Examples

```
# use demo data
snpDf <- matrix(sample(c("AA", "Aa", "aa"), 100, replace = TRUE), 10, 10)
snpDf <- as.data.frame(snpDf)
sampleGroup <- sample(c("A", "B"), 10, replace = TRUE)
result <- SNP_QC(snpDf)
```

---

|           |                                                |
|-----------|------------------------------------------------|
| ventricle | <i>a matrix of gene expression data in GEO</i> |
|-----------|------------------------------------------------|

---

Description

It is a randomly generated expression data used as an example of functions in this package. the first column represents the gene symbol

Usage

ventricle

Format

A matrix with 32 rows and 20 column

Details

the other columns represent the expression of genes

# Index

- \* **datasets**
  - gene\_cov, [19](#)
  - geneExpress, [18](#)
  - GSE66705\_sample2, [20](#)
  - kegg\_liver, [21](#)
  - module, [22](#)
  - profile, [23](#)
  - ventricle, [26](#)
- array\_preprocess, [3](#)
- cal\_mean\_module, [3](#)
- cluster\_array, [4](#)
- combine\_pvalue, [5](#)
- countToFpk, [5](#)
- countToTpm, [6](#)
- differential\_array, [7](#)
- differential\_CNV, [8](#)
- differential\_limma, [9](#)
- differential\_methy, [10](#)
- differential\_RNA, [12](#)
- differential\_SNP, [15](#)
- differential\_SNP\_GEO, [16](#)
- differential\_SNP\_tcga, [17](#)
- fpkmToTpm, [18](#)
- gene\_ave, [19](#)
- gene\_cov, [19](#)
- geneExpress, [18](#)
- get\_geo\_array, [20](#)
- GSE66705\_sample2, [20](#)
- id\_conversion\_TCGA, [21](#)
- kegg\_liver, [21](#)
- Merge\_methy\_tcga, [22](#)
- module, [22](#)
- p.adjust, [11](#), [13](#)
- prepare\_chi, [23](#)
- profile, [23](#)
- repAssign, [24](#)
- repRemove, [25](#)
- SNP\_QC, [25](#)
- ventricle, [26](#)