

# Risa: Building R objects from local ISA-Tab files

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## 1 Introduction

The Risa package is part of the ISA infrastructure software suite (<http://isa-tools.org>). It provides functionality to read ISA-Tab datasets, described in the following section. The source code and latest version can be found in the GitHub repository <https://github.com/ISA-tools/Risa>. Please, submit all 'bugs' and feature requests through <https://github.com/ISA-tools/Risa/issues>.

## 2 ISA-Tab format

The Investigation / Study / Assay (ISA) Tab-delimited (Tab) format is a general purpose framework with which to collect and communicate complex metadata (i.e. sample characteristics, technologies used, type of measurements made) from experiments employing a combination of technologies (<http://isa-tools.org>). In particular, ISA-Tab has been developed for - but not limited to - experiments using genomics, transcriptomics, proteomics or metabol/nomics techniques (the 'omics').

ISA-Tab uses three types of file to capture the experimental metadata:

- *Investigation file*
- *Study file*
- *Assay file* (with associated data files).

The Investigation file contains an overall description of an experiment while all experimental steps are described in the Study and in the Assay file(s). For each Investigation file there may be one or more Study files; for each Study file there may be one or more Assay files.

### 2.1 Investigation file

In this file, information is reported on a per-column basis and the fields are organized and divided in sections. The Investigation file is intended to meet three needs:

- to define key entities, such as factors, protocols, parameters, which may be referenced in the other files;
- to relate Assay files to Study files; and optionally,
- to relate each Study file to an Investigation (when two or more Study files need to be grouped). The declarative sections cover general information such as contacts, protocols and equipment, and also - where applicable - the description of terminologies (controlled vocabularies or ontologies) and other annotation resources that were used.

## 2.2 Study file

In this file, information is structured on a per-row basis with the first row being used for column headers. The Study file contains contextualizing information for one or more assays, for example; the subjects studied; their source(s); the sampling methodology; their characteristics; and any treatments or manipulations performed to prepare the specimens.

## 2.3 Assay file

In this file, as for the Study file, fields are organized on a per-row basis with the first row being used for column headers. The Assay file represents a portion of the experimental graph (i.e., one part of the overall structure of the workflow); each Assay file must contain assays of the same type, defined by the type of measurement (i.e. gene expression) and the technology employed (i.e. DNA microarray). Assay-related information includes protocols, additional information relating to the execution of those protocols and references to data files (whether raw or processed).

For easy transfer, ISA-Tab files and associated data files can be packaged into an ISAarchive, using a standalone Java application named ISAcreator (<http://isatab.sourceforge.net>). In order to facilitate identification of ISA-Tab components in an ISAarchive, specific extensions have been created as follows:

- *i\_iname.txt* for identifying the Investigation file
- *s\_sname.txt* for identifying Study file (s)
- *a\_aname.txt* for identifying Assay file (s)

where 'iname', 'sname', 'aname' are the user-given names for the investigation, study/ies, assay(s), respectively.

## 3 The Risa package

The Risa package is used to build R objects from an ISA archive or dataset. The output is a list of objects containing, for example, the investigation, studies and assays filenames, the contents of their files, the list of samples, among other things.

These objects can then be used by downstream Bioconductor packages for data analysis and visualization (i.e. xcms). The package currently includes the function `processAssayXcmsSet` that, for a specific mass spectrometry assay, builds an `xcmsSet` object.

### 3.1 Building an R object from a local ISA dataset

If you have your own ISA archive, you can use the function `readISAtab` to convert it into an R object. The arguments for the function `readISAtab` are:

- `path` the name of the directory containing ISAtab files. The default is the working directory.
- `verbose` a boolean indicating to show messages for the different steps, if TRUE, or not to show them, if FALSE

As an example, we can use the *faahKO* dataset, whose version 1.2.11 contains an ISA dataset describing the experiment. First, it is required to load the *Risa* package, and the *faahKO* package must have been installed.

```
> library(Risa)
> require(faahKO)
```

Then, we read the ISA-Tab data set from the *faahKO* package:

```
> faahkoISA <- readISAtab(find.package("faahKO"))
```

The object `faahkoISA` belongs to the *ISAtab* class, and contains the following elements:

- `path` - the path of the ISA-Tab dataset,
- `investigation.filename` - the name of the Investigation file
- `investigation.file` - a data frame with the contents of the Investigation file
- `study.identifiers` - the list of study identifiers
- `study.fileNames` - the names of the study files
- `study.files` - a list of data frames with the contents of the study files
- `assay.fileNames` - the names of the assay files
- `assay.fileNames.per.study` - the names of the assay files according to the study they belong to
- `assay.files` - a list of data frames with the contents of the assay files
- `assay.files.per.study` - a list of data frames with the contents of the assay files divided per study they belong to
- `assay.technology.types` - a list with the technology types corresponding to each assay
- `assay.measurement.types` - a list with the measurement types corresponding to each assay
- `data.fileNames` - a list with the names of the data files
- `samples` - a list with the names of the samples
- `samples.per.assay.filename` - the samples classified according to the assay filename they belong to
- `assay.fileNames.per.sample` - the names of the assay files classified per sample name
- `sample.to.rawdatafile` - the association between samples and raw data files
- `sample.to.assayname` - the association between samples and assay names
- `rawdatafile.to.sample` - the association between raw data files and samples
- `assayname.to.sample` - the association between assay names and samples

Additionally, the ISA dataset could be compressed in a .zip file. If that is the case, the function `readISAtab` can be used, passing the `zipfile` as parameter. The only condition is that the ISA-Tab files are contained directly into the zip file, i.e. not inside additional folders.

In this case, the parameters for the function `readISAtab` will be:

- `zipfile` a zip archive containing ISAtab files.
- `path` the name of the directory in which the files from the zip archive will be extracted. The default is the working directory.
- `verbose` a boolean indicating to show messages for the different steps, if TRUE, or not to show them, if FALSE

## Building xcmsSets for mass spectrometry assays

The function `processAssayXcmsSet` allows to build an `xcmsSet` (object defined in the `xcms` package) from the information in an assay file.

The parameters for this function are:

- `isa`: an ISA object, as retrieved by the function `readISAtab`
- `assay.filename` the name of the assay file with information about the relevant assay
- ... extra arguments that can be passed down to the `xcmsSet` function from the `xcms` package

Using the *faahKO* package as an example, we select the name of assay file, and use the `processAssayXcmsSet` to build a object of type *xcmsSet*:

```
> assay.filename <- faahkoISA["assay.names"][1]
> faahkoXset <- processAssayXcmsSet(faahkoISA, assay.filename)
```

## Augmenting the ISA-Tab dataset after analysis

The *Risa* package also provides the functionality to augment the original ISA-Tab dataset with more information after analysis.

The function `updateAssayMetadata` allows to modify the metadata in a particular assay file. The arguments are:

- `isa` An isatab object, as retrieved by the `readISAtab` function.
- `assay.filename` the filename of the assay file to be augmented/modified
- `col.name` the name of the column of the assay file to be modified
- `values` the values to be added to the column of the assay file: it could be a single value, and in this case the value is repeated across the column, or it could be a list of values (whose length must match the number of rows of the assay file)

To continue with our example using the *faahKO* data package, we will assume that the results of analysis are stored in the file *faahkoDSDF.txt*. Then, we will update the ISA-Tab dataset adding the result file into the 'Derived Spectral Data File' column of the assay file.

```
> updateAssayMetadata(faahkoISA, assay.filename, "Derived Spectral Data File", "faahkoDSDF.txt" )
```

An object of class "ISATab"

Slot "path":

```
[1] "C:/Users/biocbuild/bbs-3.11-bioc/R/library/faahKO"
```

Slot "investigation.filename":

```
[1] "i_Investigation.txt"
```

Slot "investigation.file":

	V1
1	ONTOLOGY SOURCE REFERENCE
2	Term Source Name
3	Term Source File
4	Term Source Version
5	Term Source Description
6	INVESTIGATION
7	Investigation Identifier

8	Investigation Title
9	Investigation Description
10	Investigation Submission Date
11	Investigation Public Release Date
12	Comment [Created with configuration]
13	Comment [Last Opened With Configuration]
14	INVESTIGATION PUBLICATIONS
15	Investigation PubMed ID
16	Investigation Publication DOI
17	Investigation Publication Author List
18	Investigation Publication Title
19	Investigation Publication Status
20	Investigation Publication Status Term Accession Number
21	Investigation Publication Status Term Source REF
22	INVESTIGATION CONTACTS
23	Investigation Person Last Name
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25	Investigation Person Mid Initials
26	Investigation Person Email
27	Investigation Person Phone
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29	Investigation Person Address
30	Investigation Person Affiliation
31	Investigation Person Roles
32	Investigation Person Roles Term Accession Number
33	Investigation Person Roles Term Source REF
34	STUDY
35	Study Identifier
36	Study Title
37	Study Description
38	Study Submission Date
39	Study Public Release Date
40	Study File Name
41	STUDY DESIGN DESCRIPTORS
42	Study Design Type
43	Study Design Type Term Accession Number
44	Study Design Type Term Source REF
45	STUDY PUBLICATIONS
46	Study PubMed ID
47	Study Publication DOI
48	Study Publication Author List
49	Study Publication Title
50	Study Publication Status
51	Study Publication Status Term Accession Number
52	Study Publication Status Term Source REF
53	STUDY FACTORS
54	Study Factor Name
55	Study Factor Type
56	Study Factor Type Term Accession Number
57	Study Factor Type Term Source REF
58	STUDY ASSAYS
59	Study Assay Measurement Type
60	Study Assay Measurement Type Term Source REF
61	Study Assay Measurement Type Term Accession Number

62	Study Assay Technology Type
63	Study Assay Technology Type Term Source REF
64	Study Assay Technology Type Term Accession Number
65	Study Assay Technology Platform
66	Study Assay File Name
67	STUDY PROTOCOLS
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69	Study Protocol Type
70	Study Protocol Type Term Accession Number
71	Study Protocol Type Term Source REF
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75	Study Protocol Parameters Name
76	Study Protocol Parameters Name Term Accession Number
77	Study Protocol Parameters Name Term Source REF
78	Study Protocol Components Name
79	Study Protocol Components Type
80	Study Protocol Components Type Term Accession Number
81	Study Protocol Components Type Term Source REF
82	STUDY CONTACTS
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85	Study Person Mid Initials
86	Study Person Email
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92	Study Person Roles Term Accession Number
93	Study Person Roles Term Source REF

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Slot "investigation.identifier":  
[1] ""

Slot "study.identifiers":  
[1] "Global metabolite profiling of faah(-/-) mice"

Slot "study.titles":  
[1] "Global metabolite profiling of faah(-/-) mice"

Slot "study.descriptions":  
[1] "Enzymes regulate biological processes through the conversion of specific substrates to products"

Slot "study.contacts":  
[1] " "

Slot "study.contacts.affiliations":  
[1] ""

Slot "study filenames":  
Global metabolite profiling of faah(-/-) mice  
"s\_Proteomic\_profiling\_of\_yeast\_TFs.txt"

Slot "study.files":  
\$`Global metabolite profiling of faah(-/-) mice`  
Source Name Characteristics[NEWT:Organism LC] Term Source REF  
1 Saghantelian\_1 Mus musculus (Mouse) NEWT  
2 Saghantelian\_2 Mus musculus (Mouse) NEWT  
3 Saghantelian\_3 Mus musculus (Mouse) NEWT  
4 Saghantelian\_4 Mus musculus (Mouse) NEWT  
5 Saghantelian\_5 Mus musculus (Mouse) NEWT  
6 Saghantelian\_6 Mus musculus (Mouse) NEWT  
7 Saghantelian\_7 Mus musculus (Mouse) NEWT  
8 Saghantelian\_8 Mus musculus (Mouse) NEWT  
9 Saghantelian\_9 Mus musculus (Mouse) NEWT  
10 Saghantelian\_10 Mus musculus (Mouse) NEWT  
11 Saghantelian\_11 Mus musculus (Mouse) NEWT  
12 Saghantelian\_12 Mus musculus (Mouse) NEWT

	Term	Accession	Number	Characteristics[tissue]	Term	Source	REF
1			10090	spinal cord		MA	
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7			10090	spinal cord		MA	
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9			10090	spinal cord		MA	
10			10090	spinal cord		MA	
11			10090	spinal cord		MA	
12			10090	spinal cord		MA	

Term	Accession	Number	Protocol	REF	Sample	Name	Factor	Value[Genotype]
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1	216 sample collection	K01	KO
2	216 sample collection	K02	KO
3	216 sample collection	K03	KO
4	216 sample collection	K04	KO
5	216 sample collection	K05	KO
6	216 sample collection	K06	KO
7	216 sample collection	WT1	WT
8	216 sample collection	WT2	WT
9	216 sample collection	WT3	WT
10	216 sample collection	WT4	WT
11	216 sample collection	WT5	WT
12	216 sample collection	WT6	WT

	Term	Source	REF	Term	Accession	Number
1			NA			NA
2			NA			NA
3			NA			NA
4			NA			NA
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Slot "assay.fileNames":

V2

"a\_metabolite.txt"

Slot "assay.fileNames.per.study":

\$`Global metabolite profiling of faah(-/-) mice`

\$`Global metabolite profiling of faah(-/-) mice`[[1]]

[1] "a\_metabolite.txt"

Slot "assay.files":

\$a\_metabolite.txt

	Sample	Name	Protocol	REF	Extract	Name	Protocol	REF	Labeled	Extract	Name
1		K01	extraction			K01	labeling				NA
2		K02	extraction			K02	labeling				NA
3		K03	extraction			K03	labeling				NA
4		K04	extraction			K04	labeling				NA
5		K05	extraction			K05	labeling				NA
6		K06	extraction			K06	labeling				NA
7		WT1	extraction			WT1	labeling				NA
8		WT2	extraction			WT2	labeling				NA
9		WT3	extraction			WT3	labeling				NA
10		WT4	extraction			WT4	labeling				NA
11		WT5	extraction			WT5	labeling				NA
12		WT6	extraction			WT6	labeling				NA

Label	Term	Source	REF	Term	Accession	Number	Protocol	REF
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1	NA	NA	NA	mass spectrometry
2	NA	NA	NA	mass spectrometry
3	NA	NA	NA	mass spectrometry
4	NA	NA	NA	mass spectrometry
5	NA	NA	NA	mass spectrometry
6	NA	NA	NA	mass spectrometry
7	NA	NA	NA	mass spectrometry
8	NA	NA	NA	mass spectrometry
9	NA	NA	NA	mass spectrometry
10	NA	NA	NA	mass spectrometry
11	NA	NA	NA	mass spectrometry
12	NA	NA	NA	mass spectrometry
Parameter Value[instrument] Term Source REF Term Accession Number				
1	Agilent 1100 LC-MSD SL		NA	NA
2	Agilent 1100 LC-MSD SL		NA	NA
3	Agilent 1100 LC-MSD SL		NA	NA
4	Agilent 1100 LC-MSD SL		NA	NA
5	Agilent 1100 LC-MSD SL		NA	NA
6	Agilent 1100 LC-MSD SL		NA	NA
7	Agilent 1100 LC-MSD SL		NA	NA
8	Agilent 1100 LC-MSD SL		NA	NA
9	Agilent 1100 LC-MSD SL		NA	NA
10	Agilent 1100 LC-MSD SL		NA	NA
11	Agilent 1100 LC-MSD SL		NA	NA
12	Agilent 1100 LC-MSD SL		NA	NA
Parameter Value[ion source] Term Source REF Term Accession Number				
1	electrospray ionization		MS	1000073
2	electrospray ionization		MS	1000073
3	electrospray ionization		MS	1000073
4	electrospray ionization		MS	1000073
5	electrospray ionization		MS	1000073
6	electrospray ionization		MS	1000073
7	electrospray ionization		MS	1000073
8	electrospray ionization		MS	1000073
9	electrospray ionization		MS	1000073
10	electrospray ionization		MS	1000073
11	electrospray ionization		MS	1000073
12	electrospray ionization		MS	1000073
Parameter Value[detector] Term Source REF Term Accession Number				
1	NA		NA	NA
2	NA		NA	NA
3	NA		NA	NA
4	NA		NA	NA
5	NA		NA	NA
6	NA		NA	NA
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9	NA		NA	NA
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Parameter Value[ionization mode] Term Source REF Term Accession Number				
1	positive mode		NA	NA
2	positive mode		NA	NA

3	positive mode	NA	NA
4	positive mode	NA	NA
5	positive mode	NA	NA
6	positive mode	NA	NA
7	positive mode	NA	NA
8	positive mode	NA	NA
9	positive mode	NA	NA
10	positive mode	NA	NA
11	positive mode	NA	NA
12	positive mode	NA	NA

	MS Assay Name	Raw Spectral Data File	Protocol	REF	Normalization Name
1	lc-ms-1	./cdf/KO/ko15.CDF		NA	NA
2	lc-ms-2	./cdf/KO/ko16.CDF		NA	NA
3	lc-ms-3	./cdf/KO/ko18.CDF		NA	NA
4	lc-ms-4	./cdf/KO/ko19.CDF		NA	NA
5	lc-ms-5	./cdf/KO/ko21.CDF		NA	NA
6	lc-ms-6	./cdf/KO/ko22.CDF		NA	NA
7	lc-ms-7	./cdf/WT/wt15.CDF		NA	NA
8	lc-ms-8	./cdf/WT/wt16.CDF		NA	NA
9	lc-ms-9	./cdf/WT/wt18.CDF		NA	NA
10	lc-ms-10	./cdf/WT/wt19.CDF		NA	NA
11	lc-ms-11	./cdf/WT/wt21.CDF		NA	NA
12	lc-ms-12	./cdf/WT/wt22.CDF		NA	NA

	Data Transformation Name	Derived Spectral Data File	Factor	Value[Genotype]
1	NA	faahkoDSDF.txt		KO
2	NA	faahkoDSDF.txt		KO
3	NA	faahkoDSDF.txt		KO
4	NA	faahkoDSDF.txt		KO
5	NA	faahkoDSDF.txt		KO
6	NA	faahkoDSDF.txt		KO
7	NA	faahkoDSDF.txt		WT
8	NA	faahkoDSDF.txt		WT
9	NA	faahkoDSDF.txt		WT
10	NA	faahkoDSDF.txt		WT
11	NA	faahkoDSDF.txt		WT
12	NA	faahkoDSDF.txt		WT

	Term Source	REF	Term Accession Number
1		NA	NA
2		NA	NA
3		NA	NA
4		NA	NA
5		NA	NA
6		NA	NA
7		NA	NA
8		NA	NA
9		NA	NA
10		NA	NA
11		NA	NA
12		NA	NA

```
Slot "assay.files.per.study":
$`Global metabolite profiling of faah(-/-) mice`
$`Global metabolite profiling of faah(-/-) mice`[[1]]
```

	Sample Name	Protocol	REF	Extract Name	Protocol	REF	Labeled Extract Name
1	K01	extraction		K01	labeling		NA
2	K02	extraction		K02	labeling		NA
3	K03	extraction		K03	labeling		NA
4	K04	extraction		K04	labeling		NA
5	K05	extraction		K05	labeling		NA
6	K06	extraction		K06	labeling		NA
7	WT1	extraction		WT1	labeling		NA
8	WT2	extraction		WT2	labeling		NA
9	WT3	extraction		WT3	labeling		NA
10	WT4	extraction		WT4	labeling		NA
11	WT5	extraction		WT5	labeling		NA
12	WT6	extraction		WT6	labeling		NA

	Label	Term	Source	REF	Term	Accession Number	Protocol	REF
1	NA			NA			NA mass spectrometry	
2	NA			NA			NA mass spectrometry	
3	NA			NA			NA mass spectrometry	
4	NA			NA			NA mass spectrometry	
5	NA			NA			NA mass spectrometry	
6	NA			NA			NA mass spectrometry	
7	NA			NA			NA mass spectrometry	
8	NA			NA			NA mass spectrometry	
9	NA			NA			NA mass spectrometry	
10	NA			NA			NA mass spectrometry	
11	NA			NA			NA mass spectrometry	
12	NA			NA			NA mass spectrometry	

	Parameter	Value[instrument]	Term	Source	REF	Term	Accession Number
1	Agilent	1100 LC-MSD SL			NA		NA
2	Agilent	1100 LC-MSD SL			NA		NA
3	Agilent	1100 LC-MSD SL			NA		NA
4	Agilent	1100 LC-MSD SL			NA		NA
5	Agilent	1100 LC-MSD SL			NA		NA
6	Agilent	1100 LC-MSD SL			NA		NA
7	Agilent	1100 LC-MSD SL			NA		NA
8	Agilent	1100 LC-MSD SL			NA		NA
9	Agilent	1100 LC-MSD SL			NA		NA
10	Agilent	1100 LC-MSD SL			NA		NA
11	Agilent	1100 LC-MSD SL			NA		NA
12	Agilent	1100 LC-MSD SL			NA		NA

	Parameter	Value[ion source]	Term	Source	REF	Term	Accession Number
1	electrospray	ionization			MS		1000073
2	electrospray	ionization			MS		1000073
3	electrospray	ionization			MS		1000073
4	electrospray	ionization			MS		1000073
5	electrospray	ionization			MS		1000073
6	electrospray	ionization			MS		1000073
7	electrospray	ionization			MS		1000073
8	electrospray	ionization			MS		1000073
9	electrospray	ionization			MS		1000073
10	electrospray	ionization			MS		1000073
11	electrospray	ionization			MS		1000073
12	electrospray	ionization			MS		1000073

	Parameter	Value[detector]	Term	Source	REF	Term	Accession Number
1			NA		NA		NA

2	NA	NA	NA
3	NA	NA	NA
4	NA	NA	NA
5	NA	NA	NA
6	NA	NA	NA
7	NA	NA	NA
8	NA	NA	NA
9	NA	NA	NA
10	NA	NA	NA
11	NA	NA	NA
12	NA	NA	NA

	Parameter	Value[ionization mode]	Term	Source	REF	Term	Accession	Number
1		positive mode			NA			NA
2		positive mode			NA			NA
3		positive mode			NA			NA
4		positive mode			NA			NA
5		positive mode			NA			NA
6		positive mode			NA			NA
7		positive mode			NA			NA
8		positive mode			NA			NA
9		positive mode			NA			NA
10		positive mode			NA			NA
11		positive mode			NA			NA
12		positive mode			NA			NA

	MS Assay Name	Raw	Spectral Data File	Protocol	REF	Normalization Name
1	lc-ms-1		./cdf/KO/ko15.CDF		NA	NA
2	lc-ms-2		./cdf/KO/ko16.CDF		NA	NA
3	lc-ms-3		./cdf/KO/ko18.CDF		NA	NA
4	lc-ms-4		./cdf/KO/ko19.CDF		NA	NA
5	lc-ms-5		./cdf/KO/ko21.CDF		NA	NA
6	lc-ms-6		./cdf/KO/ko22.CDF		NA	NA
7	lc-ms-7		./cdf/WT/wt15.CDF		NA	NA
8	lc-ms-8		./cdf/WT/wt16.CDF		NA	NA
9	lc-ms-9		./cdf/WT/wt18.CDF		NA	NA
10	lc-ms-10		./cdf/WT/wt19.CDF		NA	NA
11	lc-ms-11		./cdf/WT/wt21.CDF		NA	NA
12	lc-ms-12		./cdf/WT/wt22.CDF		NA	NA

	Data Transformation Name	Derived Spectral Data File	Factor Value[Genotype]
1	NA	NA	KO
2	NA	NA	KO
3	NA	NA	KO
4	NA	NA	KO
5	NA	NA	KO
6	NA	NA	KO
7	NA	NA	WT
8	NA	NA	WT
9	NA	NA	WT
10	NA	NA	WT
11	NA	NA	WT
12	NA	NA	WT

	Term	Source	REF	Term	Accession	Number
1		NA		NA		
2		NA		NA		
3		NA		NA		

4	NA	NA
5	NA	NA
6	NA	NA
7	NA	NA
8	NA	NA
9	NA	NA
10	NA	NA
11	NA	NA
12	NA	NA

Slot "assay.names":

\$a\_metabolite.txt

	MS Assay Name
1	lc-ms-1
2	lc-ms-2
3	lc-ms-3
4	lc-ms-4
5	lc-ms-5
6	lc-ms-6
7	lc-ms-7
8	lc-ms-8
9	lc-ms-9
10	lc-ms-10
11	lc-ms-11
12	lc-ms-12

Slot "assay.technology.types":

[1] "mass spectrometry"

Slot "assay.measurement.types":

[1] "metabolite profiling"

Slot "data.filenames":

\$a\_metabolite.txt

	Raw Spectral Data File	Derived Spectral Data File
1	./cdf/K0/ko15.CDF	faahkoDSDF.txt
2	./cdf/K0/ko16.CDF	faahkoDSDF.txt
3	./cdf/K0/ko18.CDF	faahkoDSDF.txt
4	./cdf/K0/ko19.CDF	faahkoDSDF.txt
5	./cdf/K0/ko21.CDF	faahkoDSDF.txt
6	./cdf/K0/ko22.CDF	faahkoDSDF.txt
7	./cdf/WT/WT15.CDF	faahkoDSDF.txt
8	./cdf/WT/WT16.CDF	faahkoDSDF.txt
9	./cdf/WT/WT18.CDF	faahkoDSDF.txt
10	./cdf/WT/WT19.CDF	faahkoDSDF.txt
11	./cdf/WT/WT21.CDF	faahkoDSDF.txt
12	./cdf/WT/WT22.CDF	faahkoDSDF.txt

Slot "samples":

[1] "K01" "K02" "K03" "K04" "K05" "K06" "WT1" "WT2" "WT3" "WT4" "WT5" "WT6"

```
Slot "samples.per.study":
$`Global metabolite profiling of faah(-/-) mice`
[1] "K01" "K02" "K03" "K04" "K05" "K06" "WT1" "WT2" "WT3" "WT4" "WT5" "WT6"
```

```
Slot "samples.per.assay.filename":
$a_metabolite.txt
[1] "K01" "K02" "K03" "K04" "K05" "K06" "WT1" "WT2" "WT3" "WT4" "WT5" "WT6"
```

```
Slot "assay filenames.per.sample":
[[1]]
[[1]][[1]]
[1] "a_metabolite.txt"
```

```
[[2]]
[[2]][[1]]
[1] "a_metabolite.txt"
```

```
[[3]]
[[3]][[1]]
[1] "a_metabolite.txt"
```

```
[[4]]
[[4]][[1]]
[1] "a_metabolite.txt"
```

```
[[5]]
[[5]][[1]]
[1] "a_metabolite.txt"
```

```
[[6]]
[[6]][[1]]
[1] "a_metabolite.txt"
```

```
[[7]]
[[7]][[1]]
[1] "a_metabolite.txt"
```

```
[[8]]
[[8]][[1]]
[1] "a_metabolite.txt"
```

```
[[9]]
[[9]][[1]]
```



```
[1] "a_metabolite.txt"
```

```
[[10]]  
[[10]][[1]]  
[1] "a_metabolite.txt"
```

```
[[11]]  
[[11]][[1]]  
[1] "a_metabolite.txt"
```

```
[[12]]  
[[12]][[1]]  
[1] "a_metabolite.txt"
```

```
Slot "sample.to.rawdatafile":
```

```
[[1]]  
  Sample Name Raw Spectral Data File  
1      KO1      ./cdf/KO/ko15.CDF  
2      KO2      ./cdf/KO/ko16.CDF  
3      KO3      ./cdf/KO/ko18.CDF  
4      KO4      ./cdf/KO/ko19.CDF  
5      KO5      ./cdf/KO/ko21.CDF  
6      KO6      ./cdf/KO/ko22.CDF  
7      WT1      ./cdf/WT/wt15.CDF  
8      WT2      ./cdf/WT/wt16.CDF  
9      WT3      ./cdf/WT/wt18.CDF  
10     WT4      ./cdf/WT/wt19.CDF  
11     WT5      ./cdf/WT/wt21.CDF  
12     WT6      ./cdf/WT/wt22.CDF
```

```
Slot "sample.to.assayname":
```

```
[[1]]  
  Sample Name MS Assay Name  
1      KO1      lc-ms-1  
2      KO2      lc-ms-2  
3      KO3      lc-ms-3  
4      KO4      lc-ms-4  
5      KO5      lc-ms-5  
6      KO6      lc-ms-6  
7      WT1      lc-ms-7  
8      WT2      lc-ms-8  
9      WT3      lc-ms-9  
10     WT4      lc-ms-10  
11     WT5      lc-ms-11  
12     WT6      lc-ms-12
```

```
Slot "rawdatafile.to.sample":
```

```
[[1]]
```

```
Raw Spectral Data File Sample Name
1      ./cdf/KO/ko15.CDF      K01
2      ./cdf/KO/ko16.CDF      K02
3      ./cdf/KO/ko18.CDF      K03
4      ./cdf/KO/ko19.CDF      K04
5      ./cdf/KO/ko21.CDF      K05
6      ./cdf/KO/ko22.CDF      K06
7      ./cdf/WT/wt15.CDF      WT1
8      ./cdf/WT/wt16.CDF      WT2
9      ./cdf/WT/wt18.CDF      WT3
10     ./cdf/WT/wt19.CDF      WT4
11     ./cdf/WT/wt21.CDF      WT5
12     ./cdf/WT/wt22.CDF      WT6
```

```
Slot "assayname.to.sample":
```

```
[[1]]
```

```
MS Assay Name Sample Name
1      lc-ms-1      K01
2      lc-ms-10     WT4
3      lc-ms-11     WT5
4      lc-ms-12     WT6
5      lc-ms-2      K02
6      lc-ms-3      K03
7      lc-ms-4      K04
8      lc-ms-5      K05
9      lc-ms-6      K06
10     lc-ms-7      WT1
11     lc-ms-8      WT2
12     lc-ms-9      WT3
```

```
Slot "factors":
```

```
[[1]]
```

```
[[1]]$`Factor Value[Genotype]`
```

```
[1] KO KO KO KO KO KO WT WT WT WT WT WT
```

```
Levels: KO WT
```

```
Slot "treatments":
```

```
$`Factor Value[Genotype]`
```

```
[1] KO WT
```

```
Levels: KO WT
```

```
Slot "groups":
```

```
[[1]]
```

```
[[1]][[1]]
```

```
[1] "K01" "K02" "K03" "K04" "K05" "K06"
```

```
[[1]][[2]]
```

```
[1] "WT1" "WT2" "WT3" "WT4" "WT5" "WT6"
```

```
Slot "assay.tabs":
[[1]]
An object of class "MSAssayTab"
Slot "path":
[1] "C:/Users/biocbuild/bbs-3.11-bioc/R/library/faahKO"
```

```
Slot "study.filename":
[1] "s-Proteomic_profiling_of_yeast_TFs.txt"
```

```
Slot "study.identifier":
[1] "Global metabolite profiling of faah(-/-) mice"
```

```
Slot "assay.filename":
[1] "a_metabolite.txt"
```

```
Slot "assay.file":
  Sample Name Protocol REF Extract Name Protocol REF Labeled Extract Name
1      KO1      extraction      KO1      labeling      NA
2      KO2      extraction      KO2      labeling      NA
3      KO3      extraction      KO3      labeling      NA
4      KO4      extraction      KO4      labeling      NA
5      KO5      extraction      KO5      labeling      NA
6      KO6      extraction      KO6      labeling      NA
7      WT1      extraction      WT1      labeling      NA
8      WT2      extraction      WT2      labeling      NA
9      WT3      extraction      WT3      labeling      NA
10     WT4      extraction      WT4      labeling      NA
11     WT5      extraction      WT5      labeling      NA
12     WT6      extraction      WT6      labeling      NA

  Label Term Source REF Term Accession Number      Protocol REF
1      NA      NA      NA      NA mass spectrometry
2      NA      NA      NA      NA mass spectrometry
3      NA      NA      NA      NA mass spectrometry
4      NA      NA      NA      NA mass spectrometry
5      NA      NA      NA      NA mass spectrometry
6      NA      NA      NA      NA mass spectrometry
7      NA      NA      NA      NA mass spectrometry
8      NA      NA      NA      NA mass spectrometry
9      NA      NA      NA      NA mass spectrometry
10     NA      NA      NA      NA mass spectrometry
11     NA      NA      NA      NA mass spectrometry
12     NA      NA      NA      NA mass spectrometry

  Parameter Value[instrument] Term Source REF Term Accession Number
1      Agilent 1100 LC-MSD SL      NA      NA
2      Agilent 1100 LC-MSD SL      NA      NA
3      Agilent 1100 LC-MSD SL      NA      NA
4      Agilent 1100 LC-MSD SL      NA      NA
5      Agilent 1100 LC-MSD SL      NA      NA
6      Agilent 1100 LC-MSD SL      NA      NA
7      Agilent 1100 LC-MSD SL      NA      NA
8      Agilent 1100 LC-MSD SL      NA      NA
```

9	Agilent 1100 LC-MSD SL		NA	NA
10	Agilent 1100 LC-MSD SL		NA	NA
11	Agilent 1100 LC-MSD SL		NA	NA
12	Agilent 1100 LC-MSD SL		NA	NA
	Parameter Value	[ion source]	Term Source REF	Term Accession Number
1	electrospray	ionization	MS	1000073
2	electrospray	ionization	MS	1000073
3	electrospray	ionization	MS	1000073
4	electrospray	ionization	MS	1000073
5	electrospray	ionization	MS	1000073
6	electrospray	ionization	MS	1000073
7	electrospray	ionization	MS	1000073
8	electrospray	ionization	MS	1000073
9	electrospray	ionization	MS	1000073
10	electrospray	ionization	MS	1000073
11	electrospray	ionization	MS	1000073
12	electrospray	ionization	MS	1000073
	Parameter Value	[detector]	Term Source REF	Term Accession Number
1		NA	NA	NA
2		NA	NA	NA
3		NA	NA	NA
4		NA	NA	NA
5		NA	NA	NA
6		NA	NA	NA
7		NA	NA	NA
8		NA	NA	NA
9		NA	NA	NA
10		NA	NA	NA
11		NA	NA	NA
12		NA	NA	NA
	Parameter Value	[ionization mode]	Term Source REF	Term Accession Number
1		positive mode	NA	NA
2		positive mode	NA	NA
3		positive mode	NA	NA
4		positive mode	NA	NA
5		positive mode	NA	NA
6		positive mode	NA	NA
7		positive mode	NA	NA
8		positive mode	NA	NA
9		positive mode	NA	NA
10		positive mode	NA	NA
11		positive mode	NA	NA
12		positive mode	NA	NA
	MS Assay Name	Raw Spectral Data File	Protocol REF	Normalization Name
1	lc-ms-1	./cdf/KO/ko15.CDF	NA	NA
2	lc-ms-2	./cdf/KO/ko16.CDF	NA	NA
3	lc-ms-3	./cdf/KO/ko18.CDF	NA	NA
4	lc-ms-4	./cdf/KO/ko19.CDF	NA	NA
5	lc-ms-5	./cdf/KO/ko21.CDF	NA	NA
6	lc-ms-6	./cdf/KO/ko22.CDF	NA	NA
7	lc-ms-7	./cdf/WT/wt15.CDF	NA	NA
8	lc-ms-8	./cdf/WT/wt16.CDF	NA	NA
9	lc-ms-9	./cdf/WT/wt18.CDF	NA	NA
10	lc-ms-10	./cdf/WT/wt19.CDF	NA	NA

11	lc-ms-11	./cdf/WT/wt21.CDF	NA	NA
12	lc-ms-12	./cdf/WT/wt22.CDF	NA	NA

	Data Transformation	Name	Derived Spectral Data File	Factor Value[Genotype]
1		NA	NA	KO
2		NA	NA	KO
3		NA	NA	KO
4		NA	NA	KO
5		NA	NA	KO
6		NA	NA	KO
7		NA	NA	WT
8		NA	NA	WT
9		NA	NA	WT
10		NA	NA	WT
11		NA	NA	WT
12		NA	NA	WT

	Term	Source REF	Term	Accession Number
1		NA		NA
2		NA		NA
3		NA		NA
4		NA		NA
5		NA		NA
6		NA		NA
7		NA		NA
8		NA		NA
9		NA		NA
10		NA		NA
11		NA		NA
12		NA		NA

Slot "assay.technology.type":  
[1] "mass spectrometry"

Slot "assay.measurement.type":  
[1] "metabolite profiling"

Slot "assay.names":

	MS Assay Name
1	lc-ms-1
2	lc-ms-2
3	lc-ms-3
4	lc-ms-4
5	lc-ms-5
6	lc-ms-6
7	lc-ms-7
8	lc-ms-8
9	lc-ms-9
10	lc-ms-10
11	lc-ms-11
12	lc-ms-12

Slot "data\_filenames":

	Raw Spectral Data File	Derived Spectral Data File
1	./cdf/KO/ko15.CDF	NA
2	./cdf/KO/ko16.CDF	NA

3	./cdf/KO/ko18.CDF	NA
4	./cdf/KO/ko19.CDF	NA
5	./cdf/KO/ko21.CDF	NA
6	./cdf/KO/ko22.CDF	NA
7	./cdf/WT/wt15.CDF	NA
8	./cdf/WT/wt16.CDF	NA
9	./cdf/WT/wt18.CDF	NA
10	./cdf/WT/wt19.CDF	NA
11	./cdf/WT/wt21.CDF	NA
12	./cdf/WT/wt22.CDF	NA

For an example for a real use case, please refer to <https://github.com/sneumann/mtbls2/>.

## Writing ISA-Tab datasets

The Risa package offers functions to write the whole ISA-Tab dataset or part of it back to disk. These functions are `write.ISAtab`, `write.investigation.file`, `write.study.file`, `write.assay.file`.

So, after updating the assay file as indicated above, we can save it back to disk, using the following command:

```
> temp = tempdir()
> write.ISAtab(faahkoISA, temp)
> #write.assay.file(faahkoISA, assay.filename, temp)
```

## Session Info

```
> toLatex(sessionInfo())
```

- R Under development (unstable) (2019-11-04 r77367), x86\_64-w64-mingw32
- Locale: LC\_COLLATE=C, LC\_CTYPE=English\_United States.1252, LC\_MONETARY=English\_United States.1252, LC\_NUMERIC=C, LC\_TIME=English\_United States.1252
- Running under: Windows Server 2012 R2 x64 (build 9600)
- Matrix products: default
- Base packages: base, datasets, grDevices, graphics, methods, parallel, stats, stats4, utils
- Other packages: Biobase 2.47.0, BiocGenerics 0.33.0, BiocParallel 1.21.0, MSnbase 2.13.0, ProtGenerics 1.19.0, Rcpp 1.0.3, Risa 1.29.0, S4Vectors 0.25.0, affy 1.65.1, biocViews 1.55.2, faahKO 1.27.0, mzR 2.21.0, xcms 3.9.0
- Loaded via a namespace (and not attached): BiocManager 1.30.9, DEoptimR 1.0-8, IRanges 2.21.1, MALDIquant 1.19.3, MASS 7.3-51.4, MassSpecWavelet 1.53.0, Matrix 1.2-17, R6 2.4.0, RANN 2.6.1, RBGL 1.63.1, RColorBrewer 1.1-2, RCurl 1.95-4.12, RUnit 0.4.32, XML 3.98-1.20, affyio 1.57.0, assertthat 0.2.1, bitops 1.0-6, codetools 0.2-16, colorspace 1.4-1, compiler 4.0.0, crayon 1.3.4, digest 0.6.22, doParallel 1.0.15, dplyr 0.8.3, foreach 1.4.7, ggplot2 3.2.1, glue 1.3.1, graph 1.65.0, grid 4.0.0, gtable 0.3.0, impute 1.61.0, iterators 1.0.12, lattice 0.20-38, lazyeval 0.2.2, limma 3.43.0, magrittr 1.5, multtest 2.43.0, munsell 0.5.0, mzID 1.25.0, ncd4 1.17, pcaMethods 1.79.1, pillar 1.4.2, pkgconfig 2.0.3, plyr 1.8.4, preprocessCore 1.49.0, purrr 0.3.3, rlang 0.4.1, robustbase 0.93-5, scales 1.0.0, snow 0.4-3, splines 4.0.0, survival 3.1-6, tibble 2.1.3, tidyselect 0.2.5, tools 4.0.0, vsn 3.55.0, zlibbioc 1.33.0

## **Further information**

For further information about the ISA software infrastructure, please visit our website <http://isa-tools.org>.