

RSVSim

an R/Bioconductor package for the simulation of structural variations

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1 Introduction

The simulation of structural variations (SV) is an important measure to assess the performance of algorithms dealing with SVs and their detection and can help with the design of sequencing experiments. A simulation generates a base exact ground truth, which can be used to test the sensitivity and precision of SV callers.

A FASTA-file with the simulated, rearranged genome can be used by common, published read simulators (like [Huang *et al.*, 2011]), [Hu *et al.*, 2012]) to generate NGS datasets from various platforms that can then be used to asses an SV algorithm . A typical workflow consists of


```

      width seq
[1]    40 AAAAAAAAAAAAAAAAAAAATTTTTTTTTTTTTTTTTTTT chr1
[2]    40 GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG chr2

```

```
> library(BSgenome.Ecoli.NCBI.20080805)
> genome = DNASTringSet(Ecoli[["NC_008253"]])
> names(genome) = "NC_008253"
```

Note, that the seeds for the randomizations in the following examples were set for demonstration purposes only. The `seed` parameter can be omitted or used to reproduce the same simulation several times.

The parameter `output` is set to `NA` in all examples to avoid wrtiting the output to disc. The parameter `verbose` is set to `FALSE` to suppress progress information about the simulation (which is enabled by default).

A segment is cut out from the genome. The following example generates three deletions of 10bp each:

```
> sim = simulateSV(output=NA, genome=genome, dels=3, sizeDels=10,
  bpSeqSize=6, seed=456, verbose=FALSE)
> sim
```

```
DNAStringSet object of length 2:
```

	width	seq	names
[1]	20	AAAAAAAAAAAAAAAATT	chr1
[2]	30	GGGGGGGGGGGGGGGGGCCCCCCCCCC	chr2

	Name	Chr	Start	End	Size	BpSeq
1	deletion1	chr1	21	30	10	ATT
2	deletion2	chr2	31	40	10	CCC
3	deletion3	chr1	19	28	10	AAATT

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2.2 Insertions

A segment is cut or copied from one chromosome A and inserted into another chromosome B. The following example generates three insertions of 5bp each:

```
> sim = simulateSV(output=NA, genome=genome, ins=3, sizeIns=5, bpSeqSize=6,
  seed=246, verbose=FALSE)
> sim
```

DNAStrngSet object of length 2:

	width	seq	names
[1]	35	AAAAAAAAAAAAAAAAATTTTTTTTTTTTTTTTTTTT	chr1
[2]	45	GAAAAAGGGGGGCCCCGGGGGGGGGGCCCCCCCCCCCC	chr2

```
> metadata(sim)
```

\$insertions

	Name	ChrA	StartA	EndA	ChrB	StartB	EndB	Size	Copied	BpSeqA	BpSeqB_5prime
1	insertion_1	chr1	13	17	chr2	2	6	5	FALSE	AAAAAA	GAAA
2	insertion_2	chr1	33	37	chr1	25	29	5	FALSE	TTTTTT	TTTTTT
3	insertion_3	chr2	34	38	chr2	9	13	5	FALSE	CCCCC	GGGCCC

BpSeqB_3prime

1	AAAGGG
2	TTTTTT
3	CCCGGG

Regarding insertion_1, for example, the 5bp segment AAAAA has been removed from chr1:14-18 and inserted into chr2:19-23. There are three breakpoint sequences reported for each insertion: the sequence at the deletion on chrA and at the 5' and 3' end of its insertion on chrB.

Setting the parameter `percCopiedIns` (range: 0-1, i.e. 0%-100%) can change the amount of "copy-and-paste-like" insertions.

The same example as before, with the difference that two of the three inserted sequences are copied:

```
> sim = simulateSV(output=NA, genome=genome, ins=3, sizeIns=5, percCopiedIns=0.66,
  bpSeqSize=6, seed=246, verbose=FALSE)
> sim
```

DNAStrngSet object of length 2:

	width	seq	names
[1]	45	AAAAAAAAAAAAAAAAAAAAAAAAATTTTTTTTTTTTTTTTTTTTTT	chr1
[2]	45	GAAAAAGGGGGGCCCCGGGGGGGGGGCCCCCCCCCCCCCCCC	chr2

```
> metadata(sim)
```

\$insertions

	Name	ChrA	StartA	EndA	ChrB	StartB	EndB	Size	Copied	BpSeqA	BpSeqB_5prime
1	insertion_1	chr1	13	17	chr2	2	6	5	TRUE		GAAA
2	insertion_2	chr1	33	37	chr1	25	29	5	TRUE		TTTTTT
3	insertion_3	chr2	34	38	chr2	9	13	5	FALSE	CCCCC	GGGCCC

BpSeqB_3prime

1	AAAGGG
2	TTTTTT
3	CCCGGG

The same sequence AAAAA from insertion_1 is now duplicated before insertion into chr2:19-23. Here, no breakpoint sequence is reported for the region on chr1, since this chromosome is not altered.

A segment is cut from one chromosome and its reverse complement is inserted at the same place without loss or a shift of sequence. The example below assigns a different size for each inversion:

```
DNAStrngSet object of length 2:
      width seq                                     names
[1]     40 AAAAAAAAAAAAAAAAAAAAAATTTTTTTTTTTTTTTT chr1
[2]     40 GGGGGGGGGGGGGGGGGGGGCCCCCGGGCCCCC      chr2

> metadata(sim)

$inversions
      Name Chr Start End Size BpSeq_3prime BpSeq_5prime
1 inversion1 chr1   21  22    2       AAATTT        AAAAAT
2 inversion2 chr2   31  34    4       GGGCCC        CCCGGG
3 inversion3 chr2   19  24    6       CCCCCC        GGGGGG
```

2.4 Tandem duplications

```
DNAStringSet object of length 2:
      width seq                      names
[1]    40 AAAAAAAAAAAAAAAAAAATTTTTTTTTTTTTTTTTT chr1
[2]    58 GGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGCCCCCCCCCCCCCCCCCCCC chr2

> metadata(sim)

$ tandemDuplications
      Name Chr Start End Size Duplications BpSeq
1 tandemDuplication1 chr2   12  17     6          3 GGGGGG
```

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A segment from the 5' or 3' end of one chromosome A is exchanged with the 5' or 3' end of another chromosome B. If it is not balanced, the segment from chromosome B will be lost, what results in a duplicated sequence from chromosome A. The parameter **percBalancedTrans** sets the amount of balanced translocation (0-1, i.e. 0%-100%); by default, all translocations will be balanced. Segments which are translocated between two different ends (5'↔3' of 3'↔5') are always inverted. After random generation of the breakpoint, the translocation spans the chromosome until the closest of both ends (which may include the centromere in the human genome).

DNAStrngSet object of length 2:

```
> metadata(sim)
```

	Name	ChrA	StartA	EndA	SizeA	ChrB	StartB	EndB	SizeB	Balanced	BpSeqA	BpSeqB	
1	translocation	1	chr2	1	3	3	chr1	37	40	4	TRUE	AAAGGG	TTTCCC

The same example in an unbalanced fashion:

DNAStrngSet object of length 2:

```
> metadata(sim)
```

	Name	ChrA	StartA	EndA	SizeA	ChrB	StartB	EndB	SizeB	Balanced	BpSeqA	BpSeqB
1	translocation_1	chr2	1	3	3	chr1	37	40	4	FALSE		TTTCCC

3 Simulation of biases towards SV formation mechanisms and repeat regions

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result of mechanisms such as nonallelic homologous recombination (NAHR), nonhomologous recombination (NHR), variable number of tandem repeats (VNTRs) and transposable element insertions (TEIs) ([Mills *et al.*, 2011], [Pang *et al.*, 2013]). These mechanisms can be further associated with repeat elements and regions of high homology such as LINEs, SINEs, Micro-/Minisatellites and segmental duplications ([Lam *et al.*, 2010]).

Using the default genome hg19 and setting `repeatBias` to `TRUE`, *RSVSim* simulates a bias of breakpoint positioning towards repeat regions. This is done in two steps:

1. Weighting SV formation mechanisms (here: NAHR, NHR, VNTR, TEI, Other) for each SV type. The type "Other" can be used for any individual weighting of repeats; by default, "Other" is associated with a random breakpoint.
2. Weighting each SV formation mechanism for each kind of repeat. The following types of repeat regions are supported: LINE/L1, LINE/L2, SINE/Alu, SINE/MIR, segmental duplications (SD), tandem repeats (TR; mainly micro-/minisatellites) and Random. The latter, "Random", means any region on the genome.

For the mechanism NAHR, both breakpoints will lie within a repeat region (with at least 50bp distance to the repeat margins), while for NHR, VNTR, TEI and Other, the repeat must make up for at least 75% of the SV region.

This feature is turned off automatically, when the user specifies his own genome (i.e. any genome other than hg19).

The default weights for SV mechanisms for deletions, insertions and duplications are based on figure 4b in [Mills *et al.*, 2011]. The weights for inversions refer to figure 3c in [Pang *et al.*, 2013]. The mechanisms and breakpoint sequences of translocations have not been studied as extensively as for other kinds of SVs. The default weights for translocations were chosen according to some exemplary publications ([Ou *et al.*, 2011], [Chen *et al.*, 2008]), so that NAHR, NHR and random breakpoint positioning contribute equally. In all cases, the results for SVs >1.000bp were used. The exact weights are:

	dels	ins	invs	dups	trans
NAHR	0.23	0.10	0.65	0.10	0.33
NHR	0.69	0.03	0.35	0.03	0.33
TEI	0.04	0.82	0.00	0.82	0.00
VNTR	0.04	0.05	0.00	0.05	0.00
Other	0.00	0.00	0.00	0.00	0.34

The default weights for repeat regions for every SV mechanism were based on the enrichment analysis in [Lam *et al.*, 2010] (see their supplemental table 5). The exact values are:

```
> data(weightsRepeats, package="RSVSim")
> show(weightsRepeats)
```

	NAHR	NHR	TEI	VNTR	Other
L1	0.59	1.04	1.66	0	0
L2	0.13	0.62	0.25	0	0
Alu	2.72	1.16	0.47	0	0
MIR	0.14	0.74	0.14	0	0
SD	5.95	2.06	0.57	0	0
TR	0.00	0.00	0.00	1	0
Random	1.00	1.00	1.00	0	1

The user may provide other weights by passing his own `data.frames`, using the function arguments `weightsMechanisms` and `weightsRepeats`. The structure of the `data.frames` has to be identical to the default ones shown above (i.e. same dimensions, column and row names). The effect of the weights

is comparable to the `prob` argument in the R function `sample`.

For example would exclude tandem repeats, segmental duplications and random regions from the simulation (except for VNTRs) by setting their weights to zero for all mechanisms. NAHRs, would be related to SINEs only. For `weightsMechanisms`, the default values will be used, because the argument is missing here. Note, that `repeatBias=TRUE` has to be set to use this feature.

This feature requires the coordinates of repeat regions for hg19. This can be handled in two ways:

- *RSVSim* downloads the coordinates once automatically from the UCSC Browser's RepeatMasker track (which may take up to 45 Minutes!).
- The user may specify the filename of a RepeatMasker output file downloaded from their homepage ([Smit *et al.*, 1996-2010]): <http://www.repeatmasker.org/species/homSap.html> (e.g. hg19.fa.out.gz). Loading this file takes only a few minutes.

In both cases, *RSVSim* saves the coordinates as `RData` object `repeats_hg19.RData` to the *RSVSim* installation directory for a faster access in the future (if write privileges allow to do so). After that, one of the two steps mentioned above is not necessary anymore and next time, *RSVSim* is going to load the coordinates automatically from the `RData` file.

When loading the repeats, neighboured ones with a distance up to 50bp will be merged, to obtain larger repeat regions and to allow SVs to span more than one repeat. But, breakpoints will only be placed within repeats of the same type (e.g. LINE/L1-LINE/L1, or SINE/MIR-SINE/MIR etc.).

4 Simulation of additional breakpoint mutations

SV breakpoints are usually not clean but tend to co-occur with other, usually smaller mutations, such as indels or SNPs. *RSVSim* allows to randomly generate additional SNPs and indels within the flanking regions of each breakpoint. Their generation can be configured by the four arguments `bpFlankSize`, `percSNPs`, `indelProb` and `maxIndelSize`, which specify the size of the flanking regions in bp (i.e. proximity of the mutations to the breakpoint), the fraction on SNPs (in %), the probability of an indel (insertions and deletions are equally likely) and the maximum size of an indel (size is selected randomly between 1 and `maxIndelSize`). Each flanking region may only contain one indel. SNPs can affect 0-100% of the region. By default, this feature is turned off.

The following example creates one deletion with 10% SNPs and 100% indel probability within 10bp up-/downstream of the breakpoint:

```
> sim = simulateSV(output=NA, genome=genome, dels=1, sizeDels=5, bpFlankSize=10,
  percSNPs=0.25, indelProb=1, maxIndelSize=3, bpSeqSize=10, seed=123, verbose=FALSE)
> sim
```

DNASTringSet object of length 2:

	width	seq	names
[1]	40	AAAAAAAAAAAAAAAAAAAAAAAAATTTTTTTTTTTTTTTTTTTT	chr1
[2]	34	GGGGGGTGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGGG	chr2

```
> metadata(sim)
```

\$deletions

	Name	Chr	Start	End	Size	BpSeq
1	deletion1	chr2	3	7	5	GGGGGGT

In addition to the 5bp deletion of the sequence CCCCC, two SNPs C→T and C→A, and a deletion of two more Cs were added up- and downstream of the breakpoint.

5 Simulation within a genome subset

It is possible to run the simulation to certain chromosomes only by specifying the chromosome names in the parameter `chrs`. It has to be taken care that these chromosome names match the names in the `DNAStrngSet` containing the genome sequences (e.g. "chr1", "chr2", ..., "chrY" for the default genome hg19 from the package *BSgenome.Hsapiens.UCSC.hg19*).

Furthermore, every SV has it's own parameter to restrict the simulation to a desired set of genomic regions: `regionsDels`, `regionsIns`, `regionsInvs`, `regionsDups` and `regionsTrans`. Each one being a `GRanges` object with a chromosome name, start- and end-position.

The following example places randomly four inversions into the second half of chr1 and the first half of chr2:

```
> regions = GRanges(IRanges(c(21,1),c(40,20)), seqnames=c("chr1","chr2"))
> regions
```

GRanges object with 2 ranges and 0 metadata columns:

```
      seqnames      ranges strand
      <Rle> <IRanges> <Rle>
[1]      chr1      21-40      *
[2]      chr2       1-20      *
```

seqinfo: 2 sequences from an unspecified genome; no seqlengths

```
> sim = simulateSV(output=NA, genome=genome, invs=4, sizeInvs=5,
  regionsInvs=regions, bpSeqSize=6, seed=2345, verbose=FALSE)
> sim
```

DNAStrngSet object of length 2:

```
      width seq                                     names
[1]     40 AAAAAAAAAAAAAAAAAAAAAATAAAAAATAAAATTTTTTTT      chr1
[2]     40 CCCCCCCCCCGGGGGGGGGCCCCCCCCCCCCCCCCCCCC      chr2
```

```
> metadata(sim)
```

\$inversions

```
      Name Chr Start End Size BpSeq_3prime BpSeq_5prime
1 inversion1 chr1    22  26    5      AAATAA      AATAAA
2 inversion2 chr1    28  32    5      AAATTT      AATAAA
3 inversion3 chr2     6  10    5      CCCGGG      CCCCCC
4 inversion4 chr2     1   5    5      CCCCCC
```

For translocations, the regions only say where to place the breakpoint, since the translocated region spans the chromosome until the closest of both ends.

Some applications may focus on certain parts of the hg19 only, like exons, introns or transcripts. The package *GenomicFeatures* provides functionalities to export such coordinates from the UCSC Genome Browser to R (see for example `makeTxDbFromUCSC`, `exonsBy`, `intronsBy`, `transcriptsBy`). In the following example, 100 deletions would be placed somewhere in the exonic regions on hg19:

```
> transcriptDB = makeTxDbFromUCSC(genome = "hg19",tablename = "knownGene")
> exons = exonsBy(transcriptDB)
> exons = unlist(exons)
> exons = GRanges(IRanges(start=start(exons), end=end(exons)), seqnames=seqnames(exons))
> simulateSV(output=NA, dels=100, regionsDels=exons, sizeDels=1000, bpSeqSize=50)
```

SVs will not be placed within unknown regions or assembly gaps denoted by the letter N. Such regions are detected and filtered automatically.

5.1 Inserting a set of SVs

The simulation allows to turn off the random generation of breakpoints and to insert a set of (for example previously detected or known) SVs. It works by using the same regions parameters and setting the parameter `random` to `FALSE`. This may also be a vector of five `TRUE/FALSE` values (in the order: deletions, insertions, inversions, tandem duplications, translocations) if some SVs shall be generated randomly and others not.

The example below inserts a deletion at chr2:16-25:

```
> knownDeletion = GRanges(IRanges(16,25), seqnames="chr2")
> names(knownDeletion) = "myDeletion"
> knownDeletion
```

GRanges object with 1 range and 0 metadata columns:

	seqnames	ranges	strand
	<Rle>	<IRanges>	<Rle>
myDeletion	chr2	16-25	*

seqinfo: 1 sequence from an unspecified genome; no seqlengths

```
> sim = simulateSV(output=NA, genome=genome, regionsDels=knownDeletion,
  bpSeqSize=10, random=FALSE, verbose=FALSE)
> sim
```

DNASTringSet object of length 2:

	width	seq	names
[1]	40	AAAAAAAAAAAAAAAAAAAAAAAAATTTTTTTTTTTTTTTTTTTT	chr1
[2]	30	GG	chr2

```
> metadata(sim)
```

\$deletions

	Name	Chr	Start	End	Size	BpSeq
myDeletion	myDeletion	chr2	16	25	10	GGGGGGGGGG

Note, that the output adopts the names, that were given the `GRanges` object of the inserted SV(s).

It's a little different for insertions and translocations, since they involve two genomic regions. Thus, the `GRanges` object for `regionsIns` has to be extended by columns `chrB` and `startB`, saying, that the sequence within ranges of the `GRanges` object will be inserted at `chrB:startB`.

The next example inserts the sequence from chr1:16:25 at chr2:26:

```
> knownInsertion = GRanges(IRanges(16,25),seqnames="chr1", chrB="chr2", startB=26)
> names(knownInsertion) = "myInsertion"
> knownInsertion
```

GRanges object with 1 range and 2 metadata columns:

	seqnames	ranges	strand	chrB	startB
	<Rle>	<IRanges>	<Rle>	<character>	<numeric>
myInsertion	chr1	16-25	*	chr2	26

seqinfo: 1 sequence from an unspecified genome; no seqlengths

```
> sim = simulateSV(output=NA, genome=genome, regionsIns=knownInsertion,
  bpSeqSize=10, random=FALSE, verbose=FALSE)
> sim
```

```

width seq names
[1] 30 AAAAAAAAAAAAAAAAAATTTTTTTTTTTTTTT chr1
[2] 50 GGGGGGGGGGGGGGGGGGGGGCCCCCAAAATTTTCCCCCCCCCCCCCCC chr2

```

\$insertions

	Name	ChrA	StartA	EndA	ChrB	StartB	EndB	Size	Copied	BpSeqA
myInsertion	myInsertion	chr1	16	25	chr2	26	35	10	FALSE	AAAAATTTTT
	BpSeqB_5prime	BpSeqB_3prime								
myInsertion	CCCCCAAAAA	TTTTTCCCCC								

The next example is a simple simulation of the translocation t(9;22) leading to the BCR-ABL fusion gene. It uses simple breakpoints within 9q34.1 and 22q11.2 for demonstration:

```
> trans_BCR_ABL = GRanges(IRanges(133000000,141213431), seqnames="chr9",
chrB="chr22", startB=23000000, endB=51304566, balanced=TRUE)
> names(trans_BCR_ABL) = "BCR_ABL"
> trans_BCR_ABL
```

seqnames	ranges	strand	chrB	startB	endB	balanced
<Rle>	<IRanges>	<Rle>	<character>	<numeric>	<numeric>	<logical>
BCR_ABL	chr9 133000000-141213431	*	chr22	23000000	51304566	TRUE

seqinfo: 1 sequence from an unspecified genome; no seqlengths

```
> sim = simulateSV(output=NA, chrs=c("chr9", "chr22"), regionsTrans=trans_BCR_ABL,
  bpSeqSize=50, random=FALSE)
```

The example is not executed here, because it requires the package *BSgenome.Hsapiens.UCSC.hg19*. Setting the argument `transInsert=20` adds up to 20 random nucleotides at both breakpoints.

It is strongly recommended to only use a set of non-overlapping SVs.

6 Comparing two sets of SVs

An overlap is defined as the overlap between the breakpoints/breakpoint regions in **simSVs**/**querySVs** up to the given tolerance in bp. Overlap does not mean the whole affected region between the start and end of the SV.

Unfortunately, there is currently no common standard format for SVs. Because the main information about SVs is their position in the genome and, sometimes, the breakpoint sequence (which depends on the SV detection algorithm), **compareSV** expects the SV detections as tables in a simple BED- or

BEDPE format (<http://code.google.com/p/bedtools>). Deletions, inversions and tandem duplications, which affect one region on the genome, can be either given in both formats. Translocations and insertions, which affect to regions on the genome, require the BEDPE-format. Eventually, the output of the SV detection format has to be converted accordingly (for example in R).

The function only compares one SV type at a time, so `querySVs` and `simSVs` may not contain a mixture of different kinds of SVs.

If the BED-tables for `querySVs` or the simulation output are saved on disk, `compareSV` also accepts their filenames and loads the tables automatically as `data.frame` in R.

The following example simulates first five 5bp deletions in the small toy genome defined above:

```
> sim = simulateSV(output=NA, genome=genome, dels=5, sizeDels=5,
  bpSeqSize=10, seed=2345, verbose=FALSE)
> simSVs = metadata(sim)$deletions
> simSVs
```

	Name	Chr	Start	End	Size	BpSeq
1	deletion1	chr1	34	38	5	TTTTTTT
2	deletion2	chr1	2	6	5	AAAAAA
3	deletion3	chr2	22	26	5	GGGGCCCCC
4	deletion4	chr2	31	35	5	CCCCCCCCC
5	deletion5	chr1	12	16	5	AAAAAAAAAT

An SV detection in BED format (the `querySVs`) may look like this: Four of five deletions were detected, two with exact and two with an approximate breakpoint. Two additional deletions were detected, which were not part of the simulation.

```
> querySVs = data.frame(
  chr=c("chr1", "chr1", "chr1", "chr2", "chr2"),
  start=c(12, 17, 32, 2, 16),
  end=c(15, 24, 36, 6, 20),
  bpSeq=c("AAAAAAAAAA", "AAAAAAATTT", "TTTTTTTTTT",
    "GGGGGGGGGG", "GGGGGGCCCC")
)
> querySVs
```

	chr	start	end	bpSeq
1	chr1	12	15	AAAAAAAAAA
2	chr1	17	24	AAAAAAATTT
3	chr1	32	36	TTTTTTTTTT
4	chr2	2	6	GGGGGGGGGG
5	chr2	16	20	GGGGGGCCCC

The column with the breakpoint sequence is optional, the column names not important (BED-files have no header).

A comparison with 0bp tolerance yields only two overlaps:

```
> compareSV(querySVs, simSVs, tol=0)
```

	Name	Chr	Start	End	Size	BpSeq	Overlap	OverlapBpSeq
1	deletion1	chr1	34	38	5	TTTTTTT		NA
2	deletion2	chr1	2	6	5	AAAAAA		NA

3	deletion3	chr2	22	26	5	GGGGCCCCC	NA
4	deletion4	chr2	31	35	5	CCCCCCCCC	NA
5	deletion5	chr1	12	16	5	AAAAAAAAAAT	NA

A higher breakpoint tolerance of +/- 3bp also includes more imprecise detections:

```
> compareSV(querySVs, simSVs, tol=3)
```

	Name	Chr	Start	End	Size	BpSeq	Overlap	OverlapBpSeq
1	deletion1	chr1	34	38	5	TTTTTTT	chr1:32-36	100
2	deletion2	chr1	2	6	5	AAAAAA		NA
3	deletion3	chr2	22	26	5	GGGGCCCCC		NA
4	deletion4	chr2	31	35	5	CCCCCCCCC		NA
5	deletion5	chr1	12	16	5	AAAAAAAAAAT	chr1:12-15	90

Note that for deletion1, the breakpoint sequence matched only by 80%.

The second example compares translocations:

```
> sim = simulateSV(output=NA, genome=genome, trans=2, percBalancedTrans=0.5,
  bpSeqSize=10, seed=246, verbose=FALSE)
> simSVs = metadata(sim)$translocations
> simSVs
```

	Name	ChrA	StartA	EndA	SizeA	ChrB	StartB	EndB	SizeB	Balanced	BpSeqA
1	translocation_1	chr1	1	13	13	chr2	1	2	2	FALSE	
2	translocation_2	chr1	37	40	4	chr2	29	40	12	TRUE	TTTTTCCCCC

BpSeqB

1	AAAAAGGGGG
2	CCCCCTTTT

Detected translocations have to be given in BEDPE-format (i.e. at least six columns chr1, start1, end1, chr2, start2, end2 for the breakpoints on both chromosomes). In this example, the breakpoints were approximated up to 1bp or 2bp, optional breakpoint sequences are missing:

```
> querySVs = data.frame(
  chr=c("chr1", "chr1", "chr2"),
  start1=c(15,32,32),
  end1=c(18,36,33),
  chr2=c("chr2", "chr2", "chr1"),
  start2=c(10,31,32),
  end2=c(12,33,36)
)
> querySVs
```

	chr	start1	end1	chr2	start2	end2
1	chr1	15	18	chr2	10	12
2	chr1	32	36	chr2	31	33
3	chr2	32	33	chr1	32	36

Here, all detected SVs span the simulated breakpoints:

```
> compareSV(querySVs, simSVs, tol=0)
```

	Name	ChrA	StartA	EndA	SizeA	ChrB	StartB	EndB	SizeB	Balanced	BpSeqA
1	translocation_1	chr1	1	13	13	chr2	1	2	2	FALSE	
2	translocation_2	chr1	37	40	4	chr2	29	40	12	TRUE	TTTTTCCCCC
	BpSeqB	Overlap	OverlapBpSeq_A	OverlapBpSeq_B							
1	AAAAAGGGGG		NA	NA							
2	CCCCCTTTT		NA	NA							

7 Setting structural variation sizes

One may specify just one size in the parameters `sizeDels`, `sizeIns`, `sizeInvs` or `sizeDups` that applies to every SV of each type. But often, it might be more realistic to assign an individual, arbitrary size to every single SV. In the simplest case, they may be uniformly distributed:

```
> sizes = sample(2:5, 5, replace=TRUE)
> sizes
```

[1] 4 4 4 2 2

```
> sim = simulateSV(output=NA, genome=genome, dels=5, sizeDels=sizes,
  bpSeqSize=6, seed=246, verbose=FALSE)
> sim
```

DNAStrngSet object of length 2:

```

width seq names
[1] 30 AAAAAAAAAAAAAATTTTTTTTTTTTTTT chr1
[2] 34 GGGGGGGGGGGGGGGGGCCCCCCCCCCCCCCCC chr2

```

```
> metadata(sim)
```

	Name	Chr	Start	End	Size	BpSeq
1	deletion1	chr1	13	16	4	AAAAAA
2	deletion2	chr2	2	5	4	GGGG
3	deletion3	chr1	32	35	4	TTTTTT
4	deletion4	chr1	8	9	2	AAAAAA
5	deletion5	chr2	29	30	2	CCCCCC

7.1 Estimating size distribution from real data

According to studies from the 1000 Genomes Project, for deletions, insertions and duplications, the amount of SVs decreases rather quickly as their size increases ([Mills *et al.*, 2011]). The function `estimateSVSizes` simulates SV sizes by fitting a beta distribution, which is flexible enough to realistically model the shape of the size distribution of all four SV types. Its two shape parameters can be derived from a given vector of SV sizes. This requires the R-package *MASS*.

The following toy example draws 1.000 SV sizes between 10bp and 1000bp from a beta distribution based on a vector of 15 SV sizes:

```
> svSizes = c(10,20,30,40,60,80,100,150,200,250,300,400,500,750,1000)
> simSizes = estimateSVSizes(n=1000, svSizes=svSizes, minSize=10, maxSize=1000, hist=TRUE)
> head(simSizes, n=20)
```

```
[1] 426 343 39 27 275 971 48 21 998 27 143 10 571 16 716 937 865 10 882 11
```

The `minSize` and `maxSize` can be omitted; they are then calculated from the given set of `svSizes`. It is recommended to use a `minSize` and `maxSize` that is consistent with the minimum/maximum values in `svSizes`.

Setting the parameter `hist=TRUE` also plots a histogram of the SV sizes to give an impression of their distribution (see Fig.1).

For deletions, insertions, inversions and tandem duplications, `estimateSVSizes` can use default parameters for the beta distribution. They were estimated from the Database of Genomic Variants (DGV) release 2012-03-29 ([Iafrate *et al.*, 2004]). Hence, no set of SV sizes is needed for fitting the distribution. In total, 1.129 deletions, 490 insertions, 202 inversions and 145 tandem duplications between 500bp and 10kb were used to estimate the shape. The parameter `default` can be set to either "deletions", "insertions", "inversions" or "tandemDuplications" to use the according set of shape parameters:

```
> delSizes = estimateSVSizes(n=10000, minSize=500, maxSize=10000,
  default="deletions", hist=TRUE)
> head(delSizes, n=15)

[1] 1838 503 2240 1071 853 751 1147 671 629 776 1814 1773 517 502 581

> delSizes = estimateSVSizes(n=10000, minSize=500, maxSize=10000,
  default="insertions", hist=TRUE)
> head(delSizes, n=15)

[1] 833 1033 3816 1334 1723 2048 2521 2874 1877 509 6090 743 1387 1796 5053

> invSizes = estimateSVSizes(n=10000, minSize=500, maxSize=10000,
  default="inversions", hist=TRUE)
> head(invSizes, n=15)

[1] 2976 1769 6532 1991 1083 1678 2841 1886 4044 1037 649 2563 771 3487 1213

> delSizes = estimateSVSizes(n=10000, minSize=500, maxSize=10000,
  default="tandemDuplications", hist=TRUE)
> head(delSizes, n=15)

[1] 6742 1260 762 5371 2623 1483 1593 1801 1388 559 1107 3039 649 2442 755
```

See Fig.2, Fig.3, Fig.4 and Fig.5 to see the estimated distribution based on the SVs in the DGV. When using these default values, it is recommended to simulate SVs that do not differ too much in size (around 500bp-10kb).

8 Runtime

The runtime of *RSVSim* mainly depends on the number of simulated breakpoints and the size of the genome.

The following test case simulates 50 SVs (10 per SV type) on the complete hg19:

```
> simulateSV(output=NA, dels=10, ins=10, inv=10, dups=10, trans=10,
  sizeDels=10000, sizeIns=10000, sizeInvs=10000, sizeDups=10000,
  repeatBias=FALSE, bpFlankSize=50, percSNPs=0.25, indelProb=0.5, maxIndelSize=10)
```

Ten repetitions of the simulation yield an average time of 6 minutes on a single Intel Xeon CPU with 2.40GHz and R version 2.15.2 (2012-10-26) including loading of the hg19 and writing of the output to

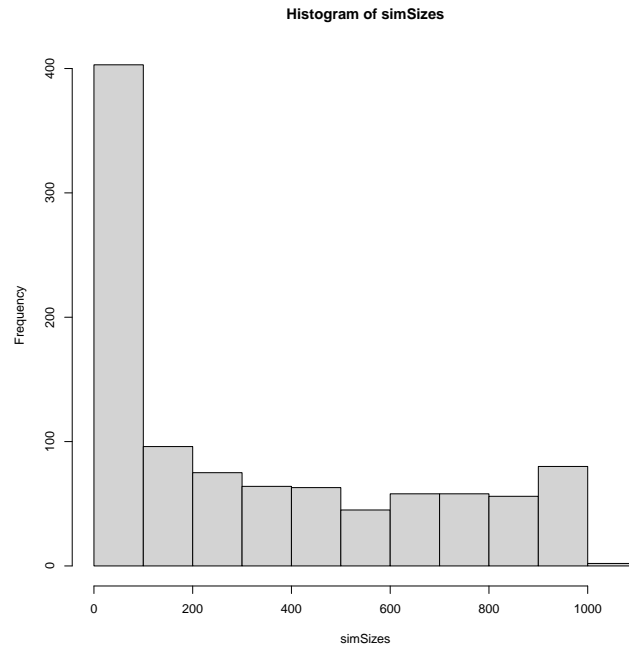


Figure 1: Distribution of 1.000 SV sizes drawn from a beta distribution using function `estimateSVSizes`.

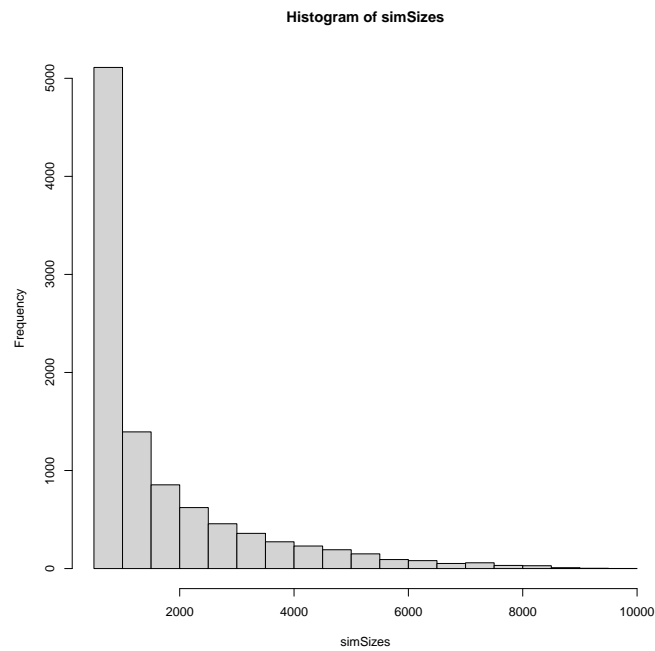


Figure 2: Distribution of 10.000 deletion sizes based on deletions from the Database of Genomic Variants.

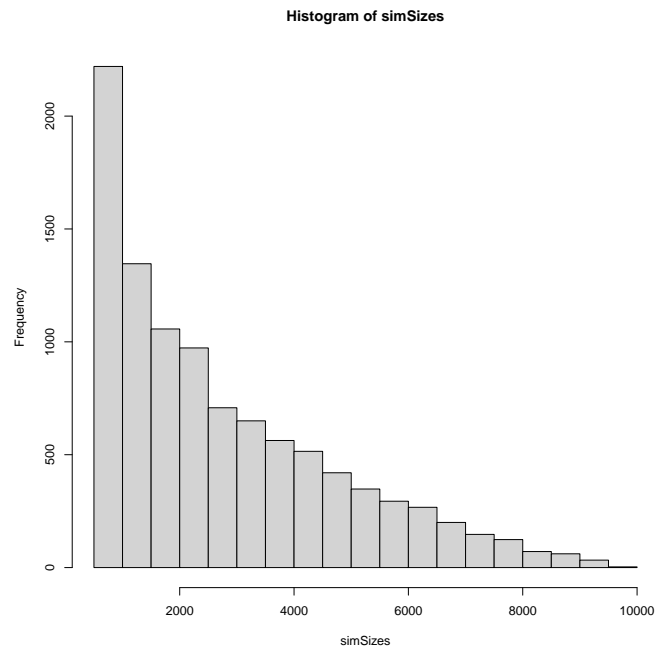


Figure 3: Distribution of 10.000 insertion sizes based on insertions from the Database of Genomic Variants.

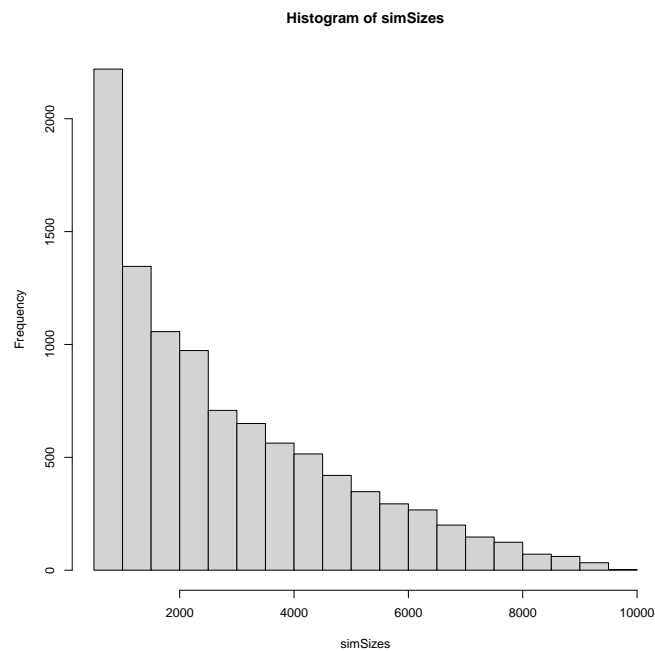


Figure 4: Distribution of 10.000 inversion sizes based on inversions from the Database of Genomic Variants.

disc.

Enabling biases for repeat regions for the same test case (i.e. `repeatBias=TRUE`), yield an average 7 minutes.

For other simulations on hg19, the runtime will scale linearly with the number of SV breakpoints.

Note, that the one-time, initial download of the repeat coordinates for hg19 from the UCSC browser may take up to 45 minutes. Alternatively, providing a RepeatMasker output file is much quicker (see section 3 for more details).

References

- [Chen *et al.*, 2008] Chen W. *et al* (2008) Mapping translocation breakpoints by next-generation sequencing, *Genome Res*, **18**(7), 1143-1149.
- [Huang *et al.*, 2011] Huang W. *et al* (2011) ART: a next-generation sequencing read simulator, *Bioinformatics*, **28** (4), 593-594.
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- [Iafrate *et al.*, 2004] Iafrate A.J. *et al* (2004) Detection of large-scale variation in the human genome, *Nat Genet.*, **36**(9), 949-951.
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- [Ou *et al.*, 2011] Ou Z. *et al* (2011) Observation and prediction of recurrent human translocations mediated by NAHR between nonhomologous chromosomes, *Genome Res*, **21**(1), 33-46.
- [Pang *et al.*, 2013] Pang A.W. *et al* (2013) Mechanisms of Formation of Structural Variation in a Fully Sequenced Human Genome, *Hum Mutat*, **34**(2), 345-354.
- [Smit *et al.*, 1996-2010] Smit A. *et al* (1996-2010) RepeatMasker Open-3.0., <<http://www.repeatmasker.org>>.

9 Session Information

R version 4.0.0 (2020-04-24)

Platform: x86_64-w64-mingw32/x64 (64-bit)

Running under: Windows Server 2012 R2 x64 (build 9600)

Matrix products: default

locale:

```
[1] LC_COLLATE=C                      LC_CTYPE=English_United States.1252
[3] LC_MONETARY=English_United States.1252 LC_NUMERIC=C
[5] LC_TIME=English_United States.1252
```

attached base packages:

```
[1] stats4      parallel  stats      graphics  grDevices  utils      datasets  methods
[9] base
```

other attached packages:

[1] MASS_7.3-51.6	RSVSim_1.28.0	GenomicRanges_1.40.0	GenomeInfoDb_1.24.0
[5] Biostrings_2.56.0	XVector_0.28.0	IRanges_2.22.0	S4Vectors_0.26.0
[9] BiocGenerics_0.34.0			

loaded via a namespace (and not attached):

[1] zlibbioc_1.34.0	GenomicAlignments_1.24.0	BiocParallel_1.22.0
[4] lattice_0.20-41	jpeg_0.1-8.1	hwriter_1.3.2
[7] tools_4.0.0	SummarizedExperiment_1.18.0	grid_4.0.0
[10] Biobase_2.48.0	png_0.1-7	latticeExtra_0.6-29
[13] matrixStats_0.56.0	crayon_1.3.4	Matrix_1.2-18
[16] GenomeInfoDbData_1.2.3	RColorBrewer_1.1-2	bitops_1.0-6
[19] RCurl_1.98-1.2	DelayedArray_0.14.0	compiler_4.0.0
[22] Rsamtools_2.4.0	ShortRead_1.46.0	

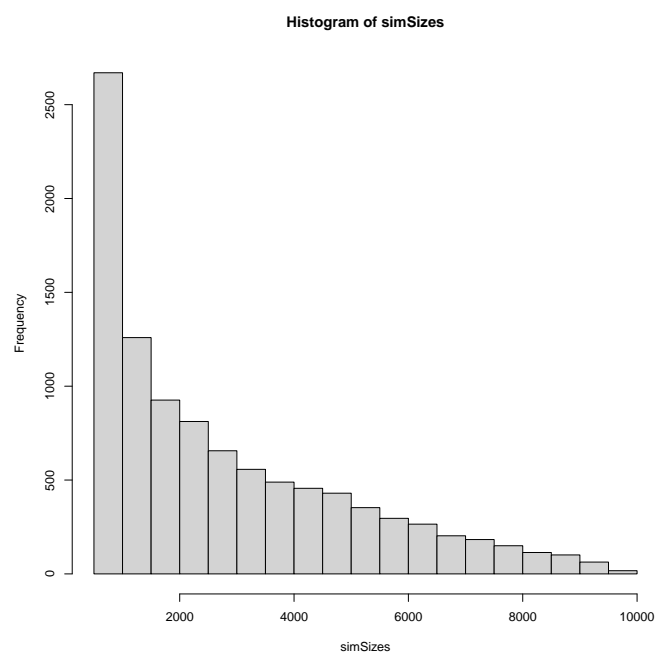


Figure 5: Distribution of 10.000 tandem duplication sizes based on tandem duplications from the Database of Genomic Variants.