

Introduction to RBM package

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1 Overview

This document provides an introduction to the RBM package. The RBM package executes the resampling-based empirical Bayes approach using either permutation or bootstrap tests based on moderated t-statistics through the following steps.

- Firstly, the RBM package computes the moderated t-statistics based on the observed data set for each feature using the `lmFit` and `eBayes` function.
- Secondly, the original data are permuted or bootstrapped in a way that matches the null hypothesis to generate permuted or bootstrapped resamples, and the reference distribution is constructed using the resampled moderated t-statistics calculated from permutation or bootstrap resamples.
- Finally, the p-values from permutation or bootstrap tests are calculated based on the proportion of the permuted or bootstrapped moderated t-statistics that are as extreme as, or more extreme than, the observed moderated t-statistics.

Additional detailed information regarding resampling-based empirical Bayes approach can be found elsewhere (Li et al., 2013).

2 Getting started

The RBM package can be installed and loaded through the following R code.
Install the RBM package with:

```
> if (!requireNamespace("BiocManager", quietly=TRUE))
+   install.packages("BiocManager")
> BiocManager::install("RBM")
```

Load the RBM package with:

```
> library(RBM)
```

3 RBM_T and RBM_F functions

There are two functions in the RBM package: `RBM_T` and `RBM_F`. Both functions require input data in the matrix format with rows denoting features and columns denoting samples. `RBM_T` is used for two-group comparisons such as study designs with a treatment group and a control group. `RBM_F` can be used for more complex study designs such as more than two groups or time-course studies. Both functions need a vector for group notation, i.e., "1" denotes the treatment group and "0" denotes the control group. For the `RBM_F` function, a contrast vector need to be provided by users to perform pairwise comparisons between groups. For example, if the design has three groups (0, 1, 2), the `aContrast` parameter will be a vector such as ("X1-X0", "X2-X1", "X2-X0") to denote all pairwise comparisons. Users just need to add an extra "X" before the group labels to do the contrasts.

- Examples using the `RBM_T` function: `normdata` simulates a standardized gene expression data and `unifdata` simulates a methylation microarray data. The p -values from the `RBM_T` function could be further adjusted using the `p.adjust` function in the `stats` package through the Benjamini-Hochberg method.

```
> library(RBM)
> normdata <- matrix(rnorm(1000*6, 0, 1),1000,6)
> mydesign <- c(0,0,0,1,1,1)
> myresult <- RBM_T(normdata,mydesign,100,0.05)
> summary(myresult)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(myresult$permutation_p<=0.05)
```

```

[1] 31

> which(myresult$permutation_p<=0.05)

[1] 19 28 43 92 113 144 200 252 254 304 334 375 381 388 392 432 538 572 630
[20] 636 685 687 744 775 787 826 889 891 904 944 995

> sum(myresult$bootstrap_p<=0.05)

[1] 9

> which(myresult$bootstrap_p<=0.05)

[1] 92 202 208 236 353 477 525 598 634

> permutation_adj_p <- p.adjust(myresult$permutation_p, "BH")
> sum(permutation_adj_p<=0.05)

[1] 5

> bootstrap_adj_p <- p.adjust(myresult$bootstrap_p, "BH")
> sum(bootstrap_adj_p<=0.05)

[1] 0

> unifdata <- matrix(runif(1000*7,0.10, 0.95), 1000, 7)
> mydesign2 <- c(0,0,0, 1,1,1,1)
> myresult2 <- RBM_T(unifdata,mydesign2,100,0.05)
> sum(myresult2$permutatioin_p<=0.05)

[1] 0

> sum(myresult2$bootstrap_p<=0.05)

[1] 10

> which(myresult2$bootstrap_p<=0.05)

[1] 94 162 199 210 233 419 614 676 848 920

> bootstrap2_adj_p <- p.adjust(myresult2$bootstrap_p, "BH")
> sum(bootstrap2_adj_p<=0.05)

[1] 0

```

- Examples using the RBM_F function: normdata_F simulates a standardized gene expression data and unifdata_F simulates a methylation microarray data. In both examples, we were interested in pairwise comparisons.

```

> normdata_F <- matrix(rnorm(1000*9,0,2), 1000, 9)
> mydesign_F <- c(0, 0, 0, 1, 1, 1, 2, 2, 2)
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult_F <- RBM_F(normdata_F, mydesign_F, aContrast, 100, 0.05)
> summary(myresult_F)

              Length Class  Mode
ordfit_t      3000   -none-  numeric
ordfit_pvalue 3000   -none-  numeric
ordfit_beta1   3000   -none-  numeric
permutation_p 3000   -none-  numeric
bootstrap_p    3000   -none-  numeric

> sum(myresult_F$permutation_p[, 1]<=0.05)

[1] 74

> sum(myresult_F$permutation_p[, 2]<=0.05)

[1] 49

> sum(myresult_F$permutation_p[, 3]<=0.05)

[1] 87

> which(myresult_F$permutation_p[, 1]<=0.05)

[1] 17 29 30 48 78 114 184 189 191 202 214 217 225 249 261 281 282 288 310
[20] 328 348 350 362 368 379 388 402 405 413 421 436 438 460 466 474 509 518 525
[39] 532 538 542 557 575 578 586 595 597 635 636 638 645 655 670 676 691 694 721
[58] 733 757 801 814 822 857 861 864 879 896 906 920 936 948 965 966 986

> which(myresult_F$permutation_p[, 2]<=0.05)

[1] 17 29 114 182 189 202 225 249 261 281 288 310 328 348 350 362 368 388 402
[20] 421 436 460 511 518 532 538 575 597 615 636 638 655 670 672 676 683 691 721
[39] 733 757 801 814 822 829 864 896 920 948 966

> which(myresult_F$permutation_p[, 3]<=0.05)

[1] 17 30 48 78 114 168 184 189 202 214 217 225 249 261 281 288 307 310 328
[20] 332 348 350 362 364 368 379 388 402 405 413 421 436 438 460 466 474 503 509
[39] 511 518 525 528 532 538 542 557 575 576 578 595 597 611 615 635 636 638 645
[58] 652 655 670 672 676 691 694 721 733 737 757 778 801 807 814 822 857 861 864
[77] 879 896 906 907 912 920 936 948 965 966 986

> con1_adj_p <- p.adjust(myresult_F$permutation_p[, 1], "BH")
> sum(con1_adj_p<=0.05/3)

```

```

[1] 16

> con2_adjp <- p.adjust(myresult_F$permutation_p[, 2], "BH")
> sum(con2_adjp<=0.05/3)

[1] 0

> con3_adjp <- p.adjust(myresult_F$permutation_p[, 3], "BH")
> sum(con3_adjp<=0.05/3)

[1] 20

> which(con2_adjp<=0.05/3)

integer(0)

> which(con3_adjp<=0.05/3)

[1] 17 189 225 328 362 368 402 405 421 532 542 575 597 636 655 676 721 733 801
[20] 822

> unifdata_F <- matrix(runif(1000*18, 0.15, 0.98), 1000, 18)
> mydesign2_F <- c(rep(0, 6), rep(1, 6), rep(2, 6))
> aContrast <- c("X1-X0", "X2-X1", "X2-X0")
> myresult2_F <- RBM_F(unifdata_F, mydesign2_F, aContrast, 100, 0.05)
> summary(myresult2_F)

              Length Class  Mode
ordfit_t      3000   -none-  numeric
ordfit_pvalue 3000   -none-  numeric
ordfit_beta1  3000   -none-  numeric
permutation_p 3000   -none-  numeric
bootstrap_p   3000   -none-  numeric

> sum(myresult2_F$bootstrap_p[, 1]<=0.05)

[1] 55

> sum(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 53

> sum(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 57

> which(myresult2_F$bootstrap_p[, 1]<=0.05)

```

```

[1] 15 22 32 64 145 147 181 192 193 231 233 261 280 282 284 289 294 295 306
[20] 361 378 380 403 427 454 460 521 533 538 539 555 557 560 581 618 670 702 707
[39] 727 739 766 785 816 817 825 842 849 850 865 875 927 931 935 963 976

> which(myresult2_F$bootstrap_p[, 2]<=0.05)

[1] 15 22 29 32 104 110 133 145 181 192 193 195 204 233 234 261 270 282 289
[20] 294 295 306 361 380 454 460 521 533 538 539 557 560 581 618 651 670 707 727
[39] 739 766 785 817 825 850 875 898 927 931 935 942 963 972 976

> which(myresult2_F$bootstrap_p[, 3]<=0.05)

[1] 15 22 32 64 82 104 110 133 145 147 193 195 204 231 233 270 282 294 295
[20] 306 361 378 380 392 427 454 474 521 533 538 539 555 560 581 618 638 646 651
[39] 670 686 707 727 739 766 785 825 829 849 850 865 875 927 931 963 972 976 986

> con21_adjp <- p.adjust(myresult2_F$bootstrap_p[, 1], "BH")
> sum(con21_adjp<=0.05/3)

[1] 4

> con22_adjp <- p.adjust(myresult2_F$bootstrap_p[, 2], "BH")
> sum(con22_adjp<=0.05/3)

[1] 4

> con23_adjp <- p.adjust(myresult2_F$bootstrap_p[, 3], "BH")
> sum(con23_adjp<=0.05/3)

[1] 6

```

4 Ovarian cancer methylation example using the RBM_T function

Two-group comparisons are the most common contrast in biological and biomedical field. The ovarian cancer methylation example is used to illustrate the application of RBM_T in identifying differentially methylated loci. The ovarian cancer methylation example is taken from the genome-wide DNA methylation profiling of United Kingdom Ovarian Cancer Population Study (UKOPS). This study used Illumina Infinium 27k Human DNA methylation Beadchip v1.2 to obtain DNA methylation profiles on over 27,000 CpGs in whole blood cells from 266 ovarian cancer women and 274 age-matched healthy controls. The data are downloaded from the NCBI GEO website with access number GSE19711. For illustration purpose, we chose the first 1000 loci in 8 randomly selected women with 4 ovarian cancer cases (pre-treatment) and 4 healthy controls. The following codes show the process of generating significant differential DNA methylation loci using the RBM_T function and presenting the results for further validation and investigations.

```
> system.file("data", package = "RBM")
```

```
[1] "C:/Users/biocbuild/bbs-3.12-bioc/tmpdir/RtmpIh9P10/Rinst261067977c07/RBM/data"
```

```
> data(ovarian_cancer_methylation)
> summary(ovarian_cancer_methylation)
```

IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]
cg00000292: 1	Min. :0.01058	Min. :0.01187	Min. :0.009103
cg00002426: 1	1st Qu.:0.04111	1st Qu.:0.04407	1st Qu.:0.041543
cg00003994: 1	Median :0.08284	Median :0.09531	Median :0.087042
cg00005847: 1	Mean :0.27397	Mean :0.28872	Mean :0.283729
cg00006414: 1	3rd Qu.:0.52135	3rd Qu.:0.59032	3rd Qu.:0.558575
cg00007981: 1	Max. :0.97069	Max. :0.96937	Max. :0.970155
(Other) :994		NA's :4	

exmdata4[, 2]	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]
Min. :0.01019	Min. :0.01108	Min. :0.01937	Min. :0.01278
1st Qu.:0.04092	1st Qu.:0.04059	1st Qu.:0.05060	1st Qu.:0.04260
Median :0.09042	Median :0.08527	Median :0.09502	Median :0.09362
Mean :0.28508	Mean :0.28482	Mean :0.27348	Mean :0.27563
3rd Qu.:0.57502	3rd Qu.:0.57300	3rd Qu.:0.52099	3rd Qu.:0.52240
Max. :0.96658	Max. :0.97516	Max. :0.96681	Max. :0.95974
	NA's :1		


```
exmdata8[, 2]
Min. :0.01357
1st Qu.:0.04387
Median :0.09282
Mean :0.28679
3rd Qu.:0.57217
Max. :0.96268
```

```
> ovarian_cancer_data <- ovarian_cancer_methylation[, -1]
> label <- c(1, 1, 0, 0, 1, 1, 0, 0)
> diff_results <- RBM_T(aData=ovarian_cancer_data, vec_trt=label, repetition=100, alpha=0.05)
> summary(diff_results)
```

	Length	Class	Mode
ordfit_t	1000	-none-	numeric
ordfit_pvalue	1000	-none-	numeric
ordfit_beta0	1000	-none-	numeric
ordfit_beta1	1000	-none-	numeric
permutation_p	1000	-none-	numeric
bootstrap_p	1000	-none-	numeric

```
> sum(diff_results$ordfit_pvalue<=0.05)
```

```
[1] 45
```

```
> sum(diff_results$permutation_p<=0.05)
```

```
[1] 73
```

```
> sum(diff_results$bootstrap_p<=0.05)
```

```
[1] 44
```

```
> ordfit_adj_p <- p.adjust(diff_results$ordfit_pvalue, "BH")
```

```
> sum(ordfit_adj_p<=0.05)
```

```
[1] 0
```

```
> perm_adj_p <- p.adjust(diff_results$permutation_p, "BH")
```

```
> sum(perm_adj_p<=0.05)
```

```
[1] 15
```

```
> boot_adj_p <- p.adjust(diff_results$bootstrap_p, "BH")
```

```
> sum(boot_adj_p<=0.05)
```

```
[1] 0
```

```
> diff_list_perm <- which(perm_adj_p<=0.05)
```

```
> diff_list_boot <- which(boot_adj_p<=0.05)
```

```
> sig_results_perm <- cbind(ovarian_cancer_methylation[diff_list_perm, ], diff_results$ordfit_t)
```

```
> print(sig_results_perm)
```

	IlmnID	Beta	exmdata2[, 2]	exmdata3[, 2]	exmdata4[, 2]
19	cg00016968	0.80628480	NA	0.81440820	0.83623180
83	cg00072216	0.04505377	0.04598964	0.04000674	0.03231534
103	cg00094319	0.73784280	0.73532960	0.75574900	0.73830220
131	cg00121904	0.15449580	0.17949750	0.23608110	0.24354150
245	cg00224508	0.04479948	0.04972043	0.04152814	0.04189373
259	cg00234961	0.04192170	0.04321576	0.05707140	0.05327565
280	cg00260778	0.64319890	0.60488960	0.56735060	0.53150910
627	cg00612467	0.04777553	0.03783457	0.05380982	0.05582291
764	cg00730260	0.90471270	0.90542290	0.91002680	0.91258610
848	cg00826384	0.05721674	0.05612171	0.06644259	0.06358381
851	cg00830029	0.58362500	0.59397870	0.64739610	0.67269640
887	cg00862290	0.43640520	0.54047160	0.60786800	0.56325950
911	cg00888479	0.07388961	0.07361080	0.10149800	0.09985076
928	cg00901493	0.03737166	0.03903724	0.04684618	0.04981432
979	cg00945507	0.13432250	0.23854600	0.34749760	0.28903340
	exmdata5[, 2]	exmdata6[, 2]	exmdata7[, 2]	exmdata8[, 2]	
19	0.80831380	0.73306440	0.82968340	0.84917800	
83	0.04965089	0.04833366	0.03466159	0.04390894	
103	0.67349260	0.73510200	0.75715920	0.78981220	
131	0.17352980	0.12564280	0.18193170	0.20847670	

245	0.04208405	0.05284988	0.03775905	0.03955271
259	0.04030003	0.03996053	0.05086962	0.05445672
280	0.61920530	0.61925200	0.46753250	0.55632410
627	0.04740551	0.05332965	0.05775211	0.05579710
764	0.90575890	0.88760470	0.90756300	0.90946790
848	0.05230160	0.06119713	0.06542751	0.06240686
851	0.50820240	0.34657470	0.66276570	0.64634510
887	0.50259740	0.40111730	0.56646700	0.54552980
911	0.08633986	0.06765189	0.09070268	0.12417730
928	0.04490690	0.04204062	0.05050039	0.05268215
979	0.11848510	0.16653850	0.30718420	0.26624740

diff_results\$ordfit_t[diff_list_perm]

19	-2.446404
83	2.514109
103	-2.268711
131	-3.451679
245	1.962457
259	-4.052697
280	4.170347
627	-2.239498
764	-1.808081
848	-2.314412
851	-2.841244
887	-3.217939
911	-3.621731
928	-2.716443
979	-4.750997

diff_results\$permutation_p[diff_list_perm]

19	0
83	0
103	0
131	0
245	0
259	0
280	0
627	0
764	0
848	0
851	0
887	0
911	0
928	0
979	0

```
> sig_results_boot <- cbind(ovarian_cancer_methylation[diff_list_boot, ], diff_results$ordfit_t)
> print(sig_results_boot)
```

```
[1] IlmnID
[2] Beta
[3] exmdata2[, 2]
[4] exmdata3[, 2]
[5] exmdata4[, 2]
[6] exmdata5[, 2]
[7] exmdata6[, 2]
[8] exmdata7[, 2]
[9] exmdata8[, 2]
[10] diff_results$ordfit_t[diff_list_boot]
[11] diff_results$bootstrap_p[diff_list_boot]
<0 rows> (or 0-length row.names)
```